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Authors: Lancaster, Zachary D., Norsworthy, Jason K., and Scott, Robert C.

Source: Weed Technology, 32(4) : 364-370

Published By: Weed Science Society of America

URL: https://doi.org/10.1017/wet.2018.13

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Weed Technology

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Residual Activity of ACCase-Inhibiting Herbicides on Monocot Crops and Weeds

Zachary D. Lancaster[1], Jason K. Norsworthy[2] and Robert C. Scott[3]

[1]Graduate Research Assistant, Department of Crop, Soil, and Environmental Sciences, University of Arkansas, Fayetteville, AR, USA, [2]Professor and Elms Farming Chair of Weed Science, Department of Crop, Soil, and Environmental Sciences, University of Arkansas, Fayetteville, AR, USA and [3]Professor, Department of Crop, Soil, and Environmental Sciences, University of Arkansas, Fayetteville, AR, USA

Abstract

Field experiments were conducted in 2014 and 2015 in Fayetteville, Arkansas, to evaluate the residual activity of acetyl-CoA carboxylase (ACCase)–inhibiting herbicides for monocot crop injury and weed control. Conventional rice, quizalofop-resistant rice, grain sorghum, and corn crops were evaluated for tolerance to soil applications of six herbicides (quizalofop at 80 and 160 g ai ha^{-1}, clethodim at 68 and 136 g ai ha^{-1}, fenoxaprop at 122 g ai ha^{-1}, cyhalofop at 313 g ai ha^{-1}, fluazifop at 210 and 420 g ai ha^{-1}, and sethoxydim at 140 and 280 g ai ha^{-1}). Overhead sprinkler irrigation of 1.3 cm was applied immediately after treatment to half of the plots, and the crops planted into the treated plots at 0, 7, and 14 d after herbicide treatment. In 2014, injury from herbicide treatments increased with activation for all crops evaluated, except for quizalofop-resistant rice. At 14 d after treatment (DAT) in 2014, corn and grain sorghum were injured 19% and 20%, respectively, from the higher rate of sethoxydim with irrigation activation averaged over plant-back dates. Conventional rice was injured 13% by the higher rate of fluazifop in 2014. Quizalofop-resistant rice was injured no more than 4% by any of the graminicides evaluated in either year. In 2015, a rainfall event occurred within 24 h of initiating the experiment; thus, there were no differences between activation via irrigation or by rainfall. However, as in 2014, grain sorghum and corn were injured 16% and 13%, respectively, by the higher rate of sethoxydim, averaged over plant-back dates. All herbicides provided little residual control of grass weeds, mainly broadleaf signalgrass and barnyardgrass. These findings indicate the need to continue allowing a plant-back interval to rice following a graminicide application, unless quizalofop-resistant rice is to be planted. The plant-back interval will vary by graminicide and the amount of moisture received following the application.

Weed Management-Major Crops

Cite this article: Lancaster ZD, Norsworthy JK, Scott RC (2018) Residual Activity of ACCase-Inhibiting Herbicides on Monocot Crops and Weeds. Weed Technol 32:364–370. doi: 10.1017/wet.2018.13

Received: 1 November 2017
Accepted: 26 January 2018

Associate Editor:
William Johnson, Purdue University

Nomenclature:
Clethodim; cyhalofop; fenoxaprop; fluazifop; quizalofop; sethoxydim; barnyardgrass, *Echinochloa crus-galli* (L.) Beauv.; broadleaf signalgrass, *Urochloa platyphylla* (Munro ex C. Wright) R.D. Webster; corn, *Zea mays* L.; grain sorghum, *Sorghum bicolor* (L.) Moench.; rice, *Oryza sativa* L.

Key words:
Residual herbicide; carryover; plant-back interval; graminicide; crop tolerance

Author for correspondence:
Zachary D. Lancaster, Department of Crop, Soil, and Environmental Sciences, University of Arkansas, Fayetteville, AR 72704.
(Email: zdlancas@email.uark.edu)

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Introduction

Rice is one of the most important crops grown in Arkansas, with over 639,000 ha planted in 2016. The state of Arkansas produces most of the rice grown in the United States, with more than twice the acreage of California, the second-place state (NASS 2016). Weed control is a major obstacle to rice production; the major weeds of Arkansas rice are barnyardgrass, sprangletops (*Leptochloa* spp.), red rice (*Oryza sativa* L.), northern jointvetch [*Aeschynomene virginica* (L.) Britton & al.], Palmer amaranth (*Amaranthus palmeri* S. Wats.), and broadleaf signalgrass (Norsworthy et al. 2013). Herbicide resistance adds to the difficulty of achieving adequate control of barnyardgrass and red rice. In Arkansas, barnyardgrass has evolved resistance to propanil, quinclorac, clomazone, and acetoacetate synthase (ALS)–inhibiting herbicides (Talbert and Burgos 2007; Norsworthy et al. 2013).

To combat the pressure placed by herbicide-resistant weeds on current production systems, growers need new technologies. Rice with resistance to quizalofop, an acetyl-CoA carboxylase (ACCase)–inhibiting herbicide, is expected to be commercialized in the United States in 2018. Provisia™, the name of this technology, will be associated with both the herbicide-resistant rice trait and the commercial quizalofop product labeled for use. The use rate for quizalofop in quizalofop-resistant rice will range from 100 to 138 g ai ha^{-1} for single applications and 240 g ai ha^{-1} for maximum yearly application (Anonymous 2017). Quizalofop, a systemic herbicide, is most notably used in soybean [*Glycine max* (L.) Merr.] for POST control of annual and perennial grasses; however, it can provide moderate residual grass control (Shaner 2014). It is anticipated that quizalofop will be restricted solely to POST applications in quizalofop-resistant rice (Youman et al. 2016).

ACCase-inhibiting herbicides are commonly used in multiple crops to selectively control annual and perennial grass species. These graminicides inhibit the enzyme acetyl-CoA carboxylase, an integral step in fatty acid synthesis. Eventually, this inhibition blocks the production of phospholipids needed for cell growth (Shaner 2014). Sethoxydim, clethodim,

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fluazifop, and quizalofop are commonly used in broadleaf crops (Anonymous 2003a,b, 2009, 2015a,b), mainly because broadleaf plants are naturally tolerant to ACCase-inhibiting herbicides (Konishi and Sasaki 1994). Broadleaf species owe this tolerance to their possession of the herbicide-tolerant prokaryote form of ACCase from the *accD* gene; lacking this gene, grass species are sensitive to ACCase-inhibiting herbicides (Konishi and Sasaki 1994). Although ACCase-inhibiting herbicides have shown high efficacy against grasses, differing levels of tolerance across grass species have been observed. These differences have led to the labeling of cyhalofop and fenoxaprop for POST use in rice (Anonymous 2003b, 2016). The tolerance in rice to cyhalofop and fenoxaprop is due to reduced absorption through the cuticle and enhanced metabolism of the herbicide compared to other susceptible grass species (Ruiz-Santaella et al. 2006).

Although graminicides are generally not applied PRE or for residual weed control, it is known that they do have limited residual activity (Barber et al. 2015). Persistence and efficacy of an herbicide in soil largely dictate the length of a plant-back interval following application. Herbicide persistence in soil can have an effect on prolonged weed suppression or can cause carryover effects in a subsequent crop (Ogle and Warren 1954). The length and extent of activity of residual herbicides may depend on both soil moisture and soil texture, among other soil chemical properties. Generally, soil-applied herbicides need 1.3 to 1.9 cm of precipitation for optimum activation (Riar et al. 2012). Activation is the movement of an herbicide into the soil profile, where it can come into contact with the germinating seed (Knake et al. 1967). Smith et al. (2016) determined that efficacy of *S*-metolachlor on Palmer amaranth was greatest when 0.6 and 1.3 cm of irrigation were applied compared to a nonirrigated check. Specific herbicides with high water solubility can move with water through the soil in the presence of rainfall or irrigation. Hence, it is possible to lose an herbicide via runoff or leaching if too much water is present (Friesen 1965). However, this movement is also influenced by an herbicide's K_d (soil sorption) and K_{oc} (soil organic carbon sorption), which can bind an herbicide to soil particles and organic matter (Wauchope et al. 2002).

Generally, plant-back intervals to monocot (grass) crops range from 30 to 120 d following most ACCase-inhibiting herbicide applications (Barber et al. 2015; Anonymous 2003a,b, 2009, 2015a,b, 2016). However, previous research on ACCase-inhibiting herbicides has shown no significant residual herbicidal activity onto subsequent grass crop plantings (Mahoney et al. 2016; Spader et al. 2012). Planting within graminicide plant-back intervals would be unlikely in the Mid-South; however, crop failure after a graminicide application could limit subsequent planting options. Likewise, the occurrence of glyphosate-resistant grass weeds in the Mid-South could also cause a decreased time between ACCase-inhibiting herbicide application and the planting of a sensitive crop. Glyphosate-resistant ryegrass [*Lolium perenne* ssp. *multiflorum* (Lam.) Husn.] was confirmed in Arkansas in 1995 (Heap 2017), glyphosate-resistant goosegrass [*Eleusine indica* (L.) Gaertn.] (Mueller et al. 2011) in Tennessee in 2011; glyphosate resistance in barnyardgrass was recently documented in Tennessee and Mississippi (Steckel et al. 2017). As a result of glyphosate resistance and the consequent reduced efficacy, many producers have begun to add graminicides to glyphosate applied prior to planting, causing reduced time between application and grass crop planting (Steckel et al. 2017). Furthermore, there has been little research to document how precipitation or irrigation could influence the residual activity of ACCase-inhibiting herbicides. Thus, we report on research conducted to determine the residual activity of ACCase-inhibiting herbicides on grass weeds and crops, with and without use of irrigation for activation. We hypothesized that graminicides evaluated would have some residual activity and thus may cause injury to corn, grain sorghum, and rice not resistant to quizalofop planted soon after application.

Materials and Methods

Experiments were conducted in 2014 and 2015 to determine the length of residual activity that could be expected on grass crops and grass weeds following ACCase-inhibiting herbicide application. The field experiment was conducted at the Agricultural Research and Extension Center in Fayetteville, Arkansas (36.4° N 94.9° W) on a Leaf silt-loam soil (fine, mixed, active, thermic Typic Albaquults) with a pH of 5.2 and organic matter content of 1.8%. Experiments were initiated June 13, 2014 and June 18, 2015. The experiment was set up as a split-split plot design, with the whole-plot factor being means of activation (irrigation immediately after application vs rainfall), the split-plot factor being plant-back interval (0, 7, and 14 d after application), and the split-split plot factor being herbicide treatment (six graminicides evaluated at multiple rates). Whole plots (refers to the factor of activation assigned to these plots) had either a 1.3-cm overhead irrigation applied with a traveling gun sprinkler system (Water Reel™, Smith Irrigation Equipment, Kensington, KS) or no irrigation (herbicide activation due to rainfall occurring just after spraying). Irrigation equipment was pre-calibrated with multiple rainfall gauges to ensure that accurate irrigation amounts were achieved.

Conventional rice, quizalofop-resistant rice, grain sorghum, and corn were planted in single rows perpendicular to the treated plots across each of the four replications of the experiment at the aforementioned three intervals. The conventional rice cultivar 'Roy J' and an experimental quizalofop-resistant variety (Provisia™ rice, BASF Corp., Research Triangle Park, NC) were planted at a seeding rate of 68 seeds m^{-1} of row. For grain sorghum, DeKalb™ hybrid DKS53-67 was planted at a seeding rate of 20 seeds m^{-1} of row, and a Smartstax™ (glyphosate/glufosinate-resistant) corn hybrid 'Croplan 6274SS' was planted at a seeding rate of 13 seeds m^{-1} of row. Herbicides were applied to a tilled, bare soil prior to planting crops using a CO_2-pressurized backpack sprayer calibrated to deliver 143 L ha^{-1} at 276 kPa. Herbicide treatments evaluated are listed in Table 1, with some herbicides applied at two rates. The split-split plot to which herbicides were applied was 1.8 by 7.6 m. The plots were over-sprayed with 2,4-D at 533 g ae ha^{-1} (Weedar™ herbicide, Nufarm Americas Inc., Alsip, IL) at 2 and 4 wks after initiating the experiment to control broadleaf weeds.

Stand counts from 1 m of row for each crop were recorded 14 d after planting (DAP). Visual observations were collected for crop injury and weed control on a scale of 0 to 100, with 0 being no injury or weed control and 100 being complete crop death or weed control. Biomass from 1 m of row for all crops and a random 1 m^2 for a natural population of broadleaf signalgrass and barnyardgrass were collected at 35 d after each separate planting. Biomass samples were oven-dried at 65 C for 14 d.

All data were analyzed with JMP Pro 12.1 (SAS Institute Inc., Cary, NC) using the Fit Model procedure. For data that met the assumptions for ANOVA, means were separated using Fisher's protected LSD ($\alpha = 0.05$). Because of differing environmental conditions, years were analyzed independently. Unlike crop response, barnyardgrass and broadleaf signalgrass measurements

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Table 1. Herbicide treatments applied before first planting at Fayetteville, Arkansas.

Herbicide treatments	Rate	Trade name	Manufacturer	Address
	g ai ha^{-1}			
Quizalofop	80	Targa	Gowan Co.	Yuma, AZ
Quizalofop	160			
Clethodim	68	SelectMax	Valent USA Corp.	Longwood, FL
Clethodim	136			
Fenoxaprop	122	Ricestar HT	Bayer CropScience LP	Research Triangle Park, NC
Cyhalofop	313	Clincher	Dow AgroSciences LLC	Indianapolis, IN
Fluazifop	210	Fusilade DX	Syngenta Crop Protection LLC	Greensboro, NC
Fluazifop	420			
Sethoxydim	140	Poast	BASF Corp.	Research Triangle Park, NC
Sethoxydim	280			

were analyzed as a split-plot design, because the weed species evaluated were a natural population; thus, there were no multiple plant-back intervals.

Results and Discussion

Overall, significant interactions and main factor effects (i.e., activation, herbicide treatment, and plant-back interval) occurred with year; thus, 2014 and 2015 data were analyzed and are presented independently. Probably, this fact can be attributed to the differing rainfall patterns between years. For 2014, ideal conditions for this experiment occurred, as there was minimal rainfall after initiation of the experiment (Figure 1), with the first appreciable rainfall of 1.2 cm 10 d after treatment (DAT). This rain-free period allowed differentiation between activation treatments through use of irrigation. Thus, the main effect and interactions with activation were generally significant for the parameters evaluated. However, in 2015 a rainfall event began within 24 h of herbicide application, with 10.4 cm of rainfall occurring within 72 h of initiating the experiment (Figure 1), resulting in no difference between activation treatments.

Crop densities at 14 DAP resulted in no significant herbicide interactions or main effects for either year (data not shown). Although a significant main effect was observed for plant-back interval for multiple crop stand counts both years, within a plant-back interval no differences between treated and nontreated plots were observed either year; thus, differences may be due to conditions that affected germination at planting. Graminicides did not appear to have an effect on stand establishment of any crop evaluated.

All crops exhibited a negative response from residual activity of the evaluated herbicides, except for quizalofop-resistant rice. Quizalofop-resistant rice showed no significant effect from any of the applied herbicides, with no more than 4% injury observed in 2014 and 3% injury in 2015 (data not shown).

Grain Sorghum Response

In 2014, a significant herbicide treatment-by-activation interaction occurred for visible injury and biomass production of grain sorghum. The greatest injury resulted from the higher rate of sethoxydim with irrigation activation (20% injury), which was significantly greater than all other herbicide treatments except the higher rate of fluazifop with irrigation (15%) (Table 2). Greater

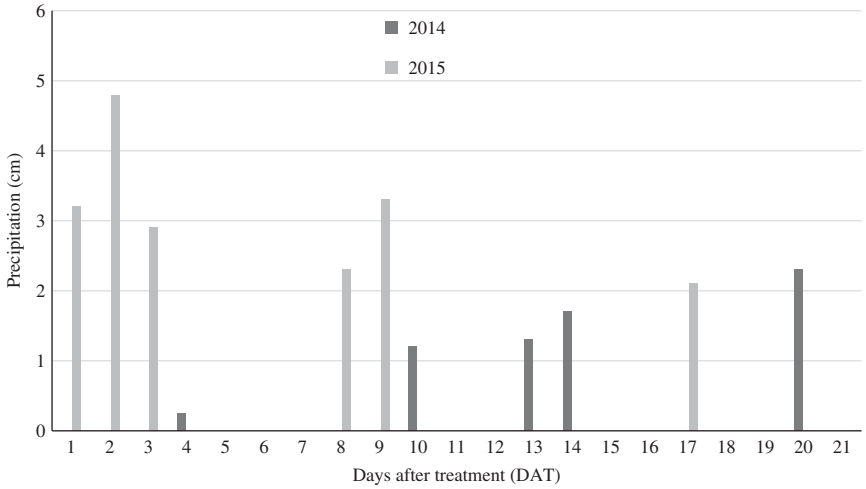


Figure 1. Precipitation 21 d after herbicide treatment for Fayetteville, Arkansas in 2014 and 2015. Experiments were initiated on June 13, 2014 and June 18, 2015.

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Table 2. Injury (14 DAP) and biomass (35 DAP) of grain sorghum, corn, and conventional rice as influenced by the residual activity of ACCase-inhibiting herbicides with and without irrigation activation in 2014 at Fayetteville, AR.[a,b]

			Grain sorghum		Corn		Conventional rice	
Activation	Herbicide	Rate	Injury[c,d]	Biomass[e,f]	Injury	Biomass	Injury[f]	Biomass[g]
		g ai ha^{-1}	----------% of nontreated----------					
Yes	Quizalofop	80	13 bc*	89 b*	9 bc	97 ab	6 bc	98
	Quizalofop	160	14 b*	86 b*	11 bc*	96 ab	5 bc	99
	Clethodim	68	13 bc*	90 ab	6 cd	97 ab	3 c	100
	Clethodim	136	14 b	88 b	5 cd	96 ab	3 c	100
	Fenoxaprop	122	13 bc*	92 ab	7 c	95 ab	3 c	99
	Cyhalofop	313	7 c	93 ab	4 cd	98 a	0	101
	Fluazifop	210	13 bc	90 ab	12 bc	95 ab	3 c	98
	Fluazifop	420	15 ab*	87 b*	13 bc*	95 ab	11 a	97
	Sethoxydim	140	9 c	94 a	5 cd	96 ab	1 cd	101
	Sethoxydim	280	20 a*	85 b*	19 a*	86 c	11 a*	98
No	Quizalofop	80	1 b	103 a	2 b	101 a	0	101
	Quizalofop	160	3 ab	101 ab	1 b	101 a	0	100
	Clethodim	68	4 ab	100 ab	3 ab	101 a	3 b	99
	Clethodim	136	7 a	99 ab	4 ab	98 b	4 ab	102
	Fenoxaprop	122	3 ab	102 ab	1 b	100 ab	0	101
	Cyhalofop	313	5 ab	100 ab	3 ab	101 a	0	99
	Fluazifop	210	5 ab	102 ab	6 a	97 b	5 ab	98
	Fluazifop	420	7 a	99 ab	6 a	97 b	8 a	101
	Sethoxydim	140	3 ab	99 ab	1 b	98 b	0	100
	Sethoxydim	280	7 a	95 b	2 b	98 b	0	98

[a]Abbreviations: ACCase, acetyl-CoA carboxylase; DAP, d after planting.
[b]Means within a column and activation level followed by the same lowercase letter are not different.
[c]Injury data expressed as percent relative to the nontreated control.
[d]Asterisk denotes increased injury with activation compared to no activation within an herbicide treatment.
[e]Biomass data expressed as percent relative to a nontreated control. Nontreated control resulted in 285, 296, and 38 g m^{-1} of row oven-dried biomass for grain sorghum, corn, and conventional rice, respectively.
[f]Treatments averaging 0 were removed from analysis for conventional rice injury due to violating the assumptions of ANOVA (homogeneity of variance).
[g]Conventional rice biomass resulted in no significant difference between treatments ($P > 0.05$).

injury from sethoxydim can most likely be attributed to having lower K_d and K_{oc} compared to other herbicides evaluated (Table 3), which led to greater availability of the herbicide in the soil. Likewise, although fluazifop is tightly bound to the soil, it rapidly degrades to fluazifop-*p*-acid, which is highly mobile in the soil and probably led to greater injury to grain sorghum (Martens 2014). Quizalofop (low and high), clethodim (low), fenoxaprop, fluazifop (high), and sethoxydim (high) resulted in greater injury when activated by sprinkler irrigation compared to the same herbicide without irrigation activation, averaged across plant-back intervals. Without irrigation for activation, injury was much lower; the highest injury was only 7% from multiple treatments, with few differences between treatments. Likewise, biomass for grain sorghum followed a trend similar to that of injury, with the lowest biomass resulting when sethoxydim was applied at a high rate with irrigation activation (85%) (Table 2); however, the sethoxydim (high) treatment with activation was only different from sethoxydim (low) with activation for relative biomass. Relative biomass was significantly reduced for quizalofop (low and high), fluazifop (high), and sethoxydim (high) with herbicide activation compared to nonactivated treatments (Table 2). Plant-back timing did not have a significant effect on either injury or relative biomass.

In 2015, with increased rainfall soon after test initiation, grain sorghum injury did not respond to activation treatment. As in 2014, sethoxydim (high) showed the greatest injury of 16% (Table 4); similarly, sethoxydim (high) produced the lowest relative biomass of 92%. Unlike 2014, a significant main effect for plant-back timing occurred in 2015 for relative grain sorghum biomass. At the plant-back timings of 0 and 7 DAT, relative biomass was 96% of the nontreated control averaged across herbicides and activation. However, at 14 DAT, relative biomass increased to 98%, thus showing an overall decrease in residual activity of the herbicides due to plant-back interval (Table 4).

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Table 3. Adsorption to soil particles (K_d), adsorption to soil organic carbon (K_{oc}), and solubility in water of ACCase-inhibiting herbicides.[a]

Herbicide	K_d	K_{oc}	Solubility in water	Source
	–––––ml g^{-1}––––		ml L^{-1}	
Clethodim	0.08–1.6	8,000	0.5–0.23	FAO 1999; Shaner 2014
Cyhalofop	265.38	2,092	0.46	Sondhia and Khare. 2014
Fenoxaprop	0.187	11,354	0.78	Anonymous 2015c; Shaner 2014
Fluazifop	0.79	5,700	1.1	Shaner 2014
Fluazifop-*p*-acid	N/A	50	780	Martens 2014
Sethoxydim	0.09–0.68	100	257	EPA 1996; Shaner 2014
Quizalofop	8.61	510	0.3	Kamrin and Montgomery 2000; Shaner 2014

[a]Abbreviations: ACCase, acetyl-CoA carboxylase; N/A, not available.

Research has shown that even though rainfall or irrigation is sometimes required to activate a herbicide in the soil, excessive rainfall can accelerate degradation of an herbicide, or cause a loss from runoff or leaching. This can reduce the length of residual activity of an herbicide (Heatherly and Hodges 1998; Splittsoesser and Derscheid 1962).

Corn Response

Like grain sorghum, a significant herbicide treatment-by-activation interaction occurred for visible injury and reduced corn biomass in 2014. Greatest injury resulted from sethoxydim (high) with activation of 19% (Table 2), which was higher than any other treatment. Herbicide treatments without activation resulted in much lower injury; the highest injury of any treatment was only 6%. Injury from quizalofop (high), fluazifop (high), and sethoxydim (high) increased when irrigation was applied, over no activation treatments. Corn biomass showed similar results, with sethoxydim (high) with activation having the lowest relative biomass of 86%, which was lower than other treatments (Table 2). Similarly, relative biomass decreased with herbicide activation for quizalofop (high) and sethoxydim (high) compared to treatments without activation.

In 2015, only herbicide treatments and plant-back intervals were significant for corn injury or relative biomass. Similar to 2014, the herbicide sethoxydim (high) produced the greatest visual injury of 13% in 2015, which was greater than any other treatment (Table 4). Sethoxydim (high) also resulted in the lowest relative biomass (93%) of any herbicide. Plant-back timing had a significant effect on corn injury, with the 0 and 7-DAT timings resulting in 7% injury averaged across herbicides and activation, whereas the 14-DAT timing resulted in lower injury at 5%.

Table 4. Herbicide treatments and plant-back interval effects on injury (14 DAP) and biomass (35 DAP) of grain sorghum, corn, and conventional rice at Fayetteville, AR, in 2015.[a,b]

		Grain sorghum		Corn		Conventional rice	
Herbicide	Rate	Injury[c]	Biomass[d]	Injury	Biomass	Injury	Biomass[e]
	g ai ha^{-1}	––% of nontreated––					
Quizalofop	80	5 de	98 bc	5 bc	98 bcd	4 cd	102
Quizalofop	160	8 bc	96 de	6 b	96 d	5 bc	100
Clethodim	68	6 cde	98 bc	3 c	100 a	2 de	98
Clethodim	136	6 cde	97 cd	3 c	99 b	3 de	103
Fenoxaprop	122	4 e	100 a	3 c	100 a	2 e	102
Cyhalofop	313	6 cde	99 ab	6 b	99 b	1 e	97
Fluazifop	210	6 cde	97 cd	6 b	98 bc	6 b	102
Fluazifop	420	9 b	95 e	7 b	97 cd	12 a	101
Sethoxydim	140	6 cde	97 cd	6 b	98 bc	4 cd	98
Sethoxydim	280	16 a	92 f	13 a	93 e	11 a	98
Plant-back interval							
0 DAT[e]			96 b	7 a			
7 DAT			96 b	7 a			
14 DAT			98 a	5 b			

[a]Abbreviations: DAP, d after planting; DAT, d after treatment.
[b]Means within a column followed by the same lowercase letter are not different according to Fisher's protected LSD ($\alpha = 0.05$).
[c]Injury data expressed as percent relative to the nontreated control.
[d]Biomass data expressed as percent relative to nontreated control. Nontreated control resulted in 276, 291, and 42 g m^{-1} of row oven-dried biomass for grain sorghum, corn, and conventional rice, respectively.
[e]Conventional rice biomass resulted in no interactions or main effects.

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Conventional Rice Response

Conventional rice showed results similar to those of grain sorghum and corn, but with generally lower levels of injury. In 2014, conventional rice injury was 11% following fluazifop (high) and sethoxydim (high) with activation (Table 2). Activation treatment only increased the injury of sethoxydim (high) from 0 without to 11% with activation. Little difference was observed between activation treatments for fluazifop (high); visual injury was 8% even without activation. Biomass of conventional rice did not show any significant interactions or main effects. In 2015, herbicide treatment was the only factor that showed significant differences in crop injury for conventional rice. Overall, injury observed in 2015 was very similar to 2014 for those treatments with activation due to the rainfall events in 2015. Fluazifop (high) and sethoxydim (high) resulted in the greatest injury to conventional rice of 12% and 11%, respectively (Table 4).

Grass Weed Control

Control of grass weeds was evaluated both years, with broadleaf signalgrass (15 plants m^{-2}) and barnyardgrass (3 plants m^{-2}) being the predominant grasses in both years. Overall, little residual weed control was observed from any ACCase-inhibiting herbicide evaluated, with only the herbicide treatment being significant for the 14-DAT rating of broadleaf signalgrass (data not shown). Subsequent control rating and relative biomass at 35 DAT did not result in any significant interactions or mains effects for broadleaf signalgrass or barnyardgrass. Because of the low level of residual injury to grass crops evaluated, little residual control of grass weeds was expected from ACCase-inhibiting herbicides.

Practical Implications

The results from this research primarily help determine plant-back intervals for ACCase-inhibiting herbicides to grass crops. The experimental outcome demonstrates that quizalofop-resistant rice is tolerant to PRE applications of ACCase-inhibiting herbicides, the cyclohexanediones and aryloxyphenoxy propionic acids, with the greatest injury reaching only 4%. Thus, quizalofop-resistant rice can be planted immediately following a graminicide application without risk of injury. Injury to conventional rice can occur if planted in close proximity to an ACCase-inhibiting herbicide application but was generally less sensitive than grain sorghum or corn. Caution must be taken with subsequent planting of grain sorghum or corn after an ACCase-inhibiting herbicide application, especially with sethoxydim. No strong impact of plant-back interval (0 to 14 DAT) on grass crop response was apparent for either year, confirming that, although the residual activity was relatively low, many of the herbicides persisted in the soil past 14 d. Timing and amount of rainfall following application of an ACCase herbicide will affect the risk for injury to a subsequent crop or the length of time between application and planting of a grass crop. A rainfall event after herbicide application can increase the residual activity of ACCase herbicides; however, large rainfall events can decrease the persistence of the herbicide in the soil. Particle runoff is the likely cause (Wauchope 1978) because of the generally high adsorption to soil particles (K_d), high adsorption to soil organic carbon (K_{oc}), and low water solubility of most ACCase-inhibiting herbicides (Table 3). In the same sense, increased microbial degradation from greater soil water availability (Parker and Doxtader 1983) could reduce residual activity of ACCase-inhibiting herbicides, which are in large part degraded by soil microbes (Shaner et al. 2014). Overall, the ACCase-inhibiting herbicides studied produced little residual grass weed control and thus should only be relied on for POST control.

References

Anonymous (2003a) Targa® herbicide product label. Gowan Co. Publication No. 03-R0415. Yuma, AZ: Gowan Co. 16 p

Anonymous (2003b) Ricestar® HT herbicide product label. Bayer CropScience Publication No. 264-682. Research Triangle Park, NC: Bayer CropScience. 5 p

Anonymous (2009) Poast Plus® herbicide product label. BASF Corp. Publication No. NVA 2008-04-026-0356. Research Triangle Park, NC: BASF Corporation. 15 p

Anonymous (2015a) Select Max® herbicide product label. Valent U.S.A. Corp. Publication No. 2015-SMAX-0001. Walnut Creek, CA: Valent U.S.A. Corp. 45 p

Anonymous (2015b) Fusilade® DX herbicide product label. Syngenta Crop Protection Publication No. SCP 1070A-L1G 0815. Greensboro, NC: Syngenta Crop Protection, LLC. 39 p

Anonymous (2015c) Ricestar® HT safety data sheet. SDS No. 102000008377. Research Triangle Park, NC: Bayer CropScience. 13 p

Anonymous (2016) Clincher® herbicide product label. Dow AgroSciences Publication No. D02-161-008. Indianapolis, IN: Dow AgroSciences LLC. 5 p

Anonymous (2017) Provisia™ herbicide product label. BASF Corp. Publication No. NVA 2017-04-522-0004. Research Triangle Park, NC: BASF Corporation. 12 p

Barber LT, Norsworthy JK, Scott RC (2015) Row crop plant-back intervals for common herbicides. Fayetteville, AR: University of Arkansas Division of Agriculture Weed Science Program. MP519

[EPA] United States Environmental Protection Agency (1996) Time-limited extensions on tolerance for residues of sethoxydim in/on asparagus, cranberries, endive, carrots, and mint. EPA series 361.Washington, DC: Environmental Protection Agency. 26 p

[FAO] Food and Agriculture Organization of the United Nations (1999) Pesticide residues in food—1999. Rome: World Health Organization. P 126

Friesen HA (1965) The movement and persistence of dicamba in the soil. Weeds 13:30–33

Heap I (2017) The International Survey of Herbicide Resistant Weeds. Available www.weedscience.org. Accessed: February 26, 2017

Heatherly LG, Hodges HF (1998) Soybean production in the Midsouth. 1st edn. Boca Raton, FL: CRC Press. 180 p

Kamrin AK, Montgomery JH (2000) Agrochemical and Pesticide Desk Reference. Boca Raton, FL: CRC Press. 349 p

Knake EL, Appleby AP, Furtick WR (1967) Soil incorporation and site of uptake of preemergence herbicides. Weeds 15:228–232

Konishi T, Sasaki Y (1994) Compartmentalization of two forms of acetyl-CoA carboxylase in plants and the origin of their tolerance toward herbicides. Proc Natl Acad Sci 91:3598–3601

Mahoney KJ, Shropshire C, Sikkema PH (2016) Examining the plant-back interval for glyphosate/glufosinate-resistant corn after the application of ACCase inhibitors. Can J Plant Sci 96:6–10

Martens J (2014) Registration of the major change in labeling for the product Fusilade DX herbicide (EPA Reg. No. 100-1070) containing the active ingredient fluazifop-*p*-butyl (chemical code 122809). Albany, NY: New York State Department of Environmental Conservation. 25 p

Mueller TC, Barnett KA, Brosnan JT, Steckel LE (2011) Glyphosate-resistant goosegrass (*Eleusine indica*) confirmed in Tennessee. Weed Sci 59:562–566

[NASS] USDA National Agricultural Statistics Service (2016) Arkansas Acreage Report 2016. https://www.nass.usda.gov/Statistics_by_State/Arkansas/Publications/Crop_Releases/Acreage/2016/aracreage16.pdf. Accessed: January 14, 2017

Norsworthy JK, Bond J, Scott RC (2013) Weed management practices and needs in Arkansas and Mississippi rice. Weed Technol 27:623–630

Ogle RE, Warren GF (1954) Fate and activity of herbicides in the soil. Weeds 3:255–258

Parker LW, Doxtader KG (1983) Kinetics of the microbial degradation of 2,4-D in soil: effects of temperature and moisture. J Environ Qual 12:553–558

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Riar DS, Norsworthy JK, Bararpour MT, Bell HD, Schrage BW (2012) Activation and length of residual herbicides under furrow and sprinkler irrigation. Pages 108–113 *in* Oosterhuis DM, ed. Summaries of Arkansas Cotton Research 2011. Arkansas Agricultural Experiment Station Research Series 602. Fayetteville, AR: Arkansas Agricultural Experiment Station

Ruiz-Santaella JP, Heredia A, Prado RD (2006) Basis of selectivity of cyhalofop-butyl in *Oryza sativa* L. Planta 223:191–199

Shaner DL, ed (2014) Herbicide Handbook. 10th edn. Lawrence, KS: Weed Science Society of America. Pp 11, 401–402

Smith HC, Ferrel JA, Webster TM, Fernandez JV, Dittmar PJ, Munoz PR, MacDonald GE (2016) Impact of irrigation volume of PRE herbicide activity. Weed Technol 30:793–800

Sondhia S, Khare RR (2014) Soil adsorption studies of a rice herbicide, cyhalofop-butyl, in two texturally different soils of India. Environ Monit Assess 186:5969–5976

Spader V, Lopes EC, dos Santos EG, Mendonca CG, Pelissari A (2012) Residual activity of ACCase inhibitor herbicides applied at pre-sowing of corn crop. Rev Bras Herb 11:42–48

Splittsoesser WE, Derscheid LA (1962) Effect of environment upon herbicides applied preemergence. Weeds 10:304–307

Steckel LE, Bond JA, Montgomery GB, Phillips TL, Nandula V (2017) Glyphosate-resistant barnyardgrass in Tennessee and Mississippi. Pages 182–183 *in* Proceedings of the Southern Weed Science Society 70th Annual Meeting, January 23–26, 2017. Birmingham, AL: Southern Weed Science Society

Talbert RE, Burgos NR (2007) History and management of herbicide-resistant barnyardgrass (*Echinochloa crus-galli*) in Arkansas rice. Weed Technol 21:324–331

Wauchope RD (1978) The pesticide content of surface water draining from agricultural fields – a review. J Environ Qual 7:459–478

Wauchope RD, Yeh S, Linders J, Kloskowski R, Tanaka K, Rubin K, Katayama A, Kordel W, Gerstl Z, Lane M, Unsworth JB (2002) Pesticide soil sorption parameters: theory, measurement, uses, limitations, and reliability. Pest Manag Sci 58:419–445

Youman C, Guice J, Rhodes A, Schultz A, Harden J (2016) Provisia™ rice production system efficacy and stewardship. Page 278 *in* Proceedings of the Southern Weed Science Society 69th Annual Meeting, February 7–11, 2016. San Juan, PR: Southern Weed Science Society

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