

Resistance of Inzen™ grain sorghum to multiple PRE- and POST-applied acetolactate synthase–inhibiting herbicides

Authors: Bowman, Hunter D., Barber, Tom, Norsworthy, Jason K., Roberts, Trenton L., Kelley, Jason, et al.

Source: Weed Technology, 35(1) : 57-64

Published By: Weed Science Society of America

URL: <https://doi.org/10.1017/wet.2020.69>

BioOne Complete ([complete.BioOne.org](https://complete.bioone.org)) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at www.bioone.org/terms-of-use.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

Research Article

Cite this article: Bowman HD, Barber T, Norsworthy JK, Roberts TL, Kelley J, Gbur EE, Steckel LE (2021) Resistance of Inzen™ grain sorghum to multiple PRE- and POST-applied acetolactate synthase-inhibiting herbicides. *Weed Technol.* **35**: 57–64. doi: [10.1017/wet.2020.69](https://doi.org/10.1017/wet.2020.69)

Received: 23 March 2020

Revised: 28 May 2020

Accepted: 18 June 2020

First published online: 30 June 2020

Nomenclature:

Nicosulfuron; pyriithiobac; grain sorghum, *Sorghum bicolor* L. Moench ssp. *bicolor*


Keywords:

Cross-resistance; injury; tolerance

Author for correspondence:

Hunter Dewayne Bowman, Department of Crop, Soil, and Environmental Sciences, 115 Plant Sciences Building, University of Arkansas, Fayetteville, AR 72701. Email: hdbowman16@gmail.com

Resistance of Inzen™ grain sorghum to multiple PRE- and POST-applied acetolactate synthase-inhibiting herbicides

Hunter D. Bowman¹ , Tom Barber², Jason K. Norsworthy³, Trenton L. Roberts⁴, Jason Kelley⁵ and Edward E. Gbur⁶

¹Former graduate student, Department of Crop, Soil, and Environmental Sciences, University of Arkansas, Fayetteville, AR, USA; ²Professor, Department of Crop, Soil, and Environmental Sciences, University of Arkansas, Lonoke, AR, USA; ³Professor and Elms Farming Chair of Weed Science, Department of Crop, Soil, and Environmental Sciences, University of Arkansas, Fayetteville, AR, USA; ⁴Associate Professor, Department of Crop, Soil, and Environmental Sciences, University of Arkansas, Fayetteville, AR, USA; ⁵Professor, Department of Crop, Soil, and Environmental Sciences, University of Arkansas, Lonoke, AR, USA and ⁶Professor, Agricultural Statistics Laboratory, University of Arkansas, Fayetteville, AR, USA

Abstract

A non-GMO trait called Inzen™ was recently commercialized in grain sorghum to combat weedy grasses, allowing the use of nicosulfuron POST in the crop. Inzen™ grain sorghum carries a double mutation in the acetolactate synthase (ALS) gene Val₅₆₀Ile and Trp₅₇₄Leu, which potentially results in cross-resistance to a wide assortment of ALS-inhibiting herbicides. To evaluate the scope of cross-resistance to Weed Science Society of America Group 2 herbicides in addition to nicosulfuron, tests were conducted in 2016 and 2017 at the Lon Mann Cotton Research Station near Marianna, AR, the Arkansas Agricultural Research and Extension Center in Fayetteville, AR, and in 2016 at the Pine Tree Research Station near Colt, AR. The tests included ALS-inhibiting herbicides from all five families: sulfonylureas, imidazolinones, pyrimidinylthiobenzoics, triazolinones, and triazolopyrimidines. Treatments were made PRE or POST to grain sorghum at a 1× rate for crops in which each herbicide is labeled. Grain sorghum planted in the PRE trial were Inzen™ and a conventional cultivar. Visible estimates of injury and sorghum heights were recorded at 2 and 4 wk after herbicide application, and yield data were collected at crop maturity. In the PRE trial, no visible injury, sorghum height reduction, or yield loss were observed in plots containing the Inzen™ cultivar. Applications made POST to the Inzen™ grain sorghum caused visible injury, sorghum height reduction, and yield loss of 20%, 13%, and 35%, respectively, only in plots where bispyribac-Na was applied. There was no impact on the crop from other POST-applied ALS-inhibiting herbicides. These results demonstrate that the Inzen™ trait confers cross-resistance to most ALS-inhibiting herbicides and could offer promising new alternatives for weed control and protection from carryover of residual ALS-inhibiting herbicides in grain sorghum.

Introduction

Grain sorghum is a popular crop to include in a crop rotation, because it permits the effective use of atrazine for control of many weeds, particularly broadleaf weeds (Owen and Powles 2010). However, producers still face weed control issues in grain sorghum because of the limited POST chemical options for weedy grass control once the crop has emerged (Smith and Scott 2015). More specifically, johnsongrass [*Sorghum halepense* (L.) Pers.], fall panicum [*Panicum dichotomiflorum* Michx.], barnyardgrass [*Echinochloa crus-galli* (L.) Beauv.], broadleaf signalgrass [*Urochloa platyphylla* Nash], and Texas panicum [*Panicum texanum* (Buckl.) R. Webster] can be the most troublesome grass species.

The ALS chemistry has not been available in grain sorghum until now because of the strong genetic similarities between johnsongrass and grain sorghum (Bowers et al. 2003). However, with the discovery of nicosulfuron-resistant weedy sorghum, researchers were able to cross-pollinate nicosulfuron resistance into grain sorghum, allowing for safe use of nicosulfuron in the crop (Anonymous 2016; Tuinstra and Al-Khatib 2008).

Johnsongrass can reproduce through seed as a summer annual or through rhizomes as a perennial. Johnsongrass can produce up to 28,000 seeds and up to 90 m of rhizomes in one season of grain sorghum growth (Horowitz 1973). Not only does johnsongrass reduce grain sorghum yield by competing with the crop, but it also produces allelopathic chemicals that inhibit grain sorghum growth (Mueller et al. 1993). Johnsongrass alone can reduce corn (*Zea mays* L.) yields 74% to 100% (Bendixen 1986). Control of the troublesome weed has been achieved in other crops such as soybean (*Glycine max* L.) by using selective herbicides such as dinitroanilines and acetyl-coenzyme A carboxylase-inhibiting herbicides (Langemeier and Witt 1986;

© Weed Science Society of America, 2020. This is an Open Access article, distributed under the terms of the Creative Commons Attribution licence (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted re-use, distribution, and reproduction in any medium, provided the original work is properly cited.



McWhorter 1977; Riley and Shaw 1988). The evolution of resistance, however, has reduced these options (Heap 2020). The introduction of glyphosate-resistant corn, cotton, and soybeans provided a more effective option for control with glyphosate. Prior to the introduction of glyphosate-resistant corn, nicosulfuron was the primary option for POST control of grasses in corn.

With the Inzen™ technology introduced to the market, questions have been proposed as to whether or not the herbicide-resistant trait will confer resistance to other Weed Science Society of America (WSSA) Group 2 ALS-inhibiting herbicides. ALS-inhibiting herbicides, as defined by the WSSA (2017), primarily starve the plant of branched-chain amino acids, specifically leucine, isoleucine, and valine. The five families composing herbicides that inhibit ALS function include sulfonylureas, imidazolinones, pyrimidinylthio-benzoics, triazolinones, and triazolopyrimidines.

Cross-resistance across a suite of ALS-inhibiting herbicide families was documented by Tranel and Wright (2002). However, their research found that ALS resistance generally is grouped into one of three categories: sulfonylurea resistant, imidazolinone resistant, or a broad cross-resistance. Resistance to ALS herbicides was first discovered only 5 yr after the introduction of the first sulfonylurea herbicide chlorsulfuron. The mechanism of resistance to chlorsulfuron was reduced sensitivity of the target ALS enzyme to inhibition by the herbicide (Mallory-Smith et al. 1990). Currently, there are nine confirmed ALS enzyme mutations that confer resistance to ALS-inhibiting herbicides: Ala₁₂₂, Pro₁₉₇, Ala₂₀₅, Asp₃₇₆, Arg₃₇₇, Trp₅₇₄, Ser₆₅₃, Val₅₆₀, and Gly₆₅₄. The Ala₁₂₂ mutation is the most common one documented to confer cross-resistance (Tranel et al. 2016). PCR screening of Inzen™ sorghum revealed a double mutation of Val₅₆₀Ile and Trp₅₇₄Leu (Tuinstra and Al-Khatib 2008). The Trp₅₇₄Leu point mutation has been found in Palmer amaranth (*Amaranthus palmeri* S. Watson), which resulted in ALS cross-resistance to all five families in the species (Molin et al. 2016). The Val₅₆₀Ile point mutation has also been documented conferring ALS cross-resistance to all five ALS-inhibiting herbicide families in johnsongrass (Werle et al. 2016). Another mechanism of ALS-inhibiting herbicide resistance is increased herbicide metabolism resulting in detoxification of the herbicide. However, enhanced metabolism only results in cross-resistance to ALS herbicides less than 10% of the time (Hall et al. 1994). Therefore, the objective of this research was to evaluate the cross-resistance of Inzen™ grain sorghum to PRE and POST applications of various WSSA Group 2 ALS-inhibiting herbicides.

Materials and Methods

Research was conducted at the Lon Mann Cotton Research Station (LMCRS) near Marianna, AR (34.44°N, 90.45°W), and the Arkansas Agricultural Research and Extension Center (AAREC) in Fayetteville, AR (36.05°N, 94.55°W), in 2016 and 2017, and the Pine Tree Research Station (PTRS) near Colt, AR, (35.07°N, 90.10°W), in 2016 to evaluate the scope of cross-resistance of Inzen™ sorghum herbicides to WSSA Group 2. The soil texture at LMCRS was a Calloway silt loam (fine-silty, mixed, active, thermic Aquic Fraglossudalfs) with 2% sand, 82.3% silt, and 15.6% clay, a pH of 5.5, and 2.2% organic matter (OM). The AAREC soil texture was a Captina silt loam (fine-silty, siliceous, active, mesic Typic Fragiudults) with 22% sand, 64% silt, and 14% clay, a pH of 5.8, and 1.8% OM. The PTRS soil texture was a Calloway silt loam (fine-silty, mixed, active, thermic Aquic Fraglossudalfs) with 10.6% sand, 68.6% silt, and 20.8% clay, a pH of 7.5, and 1.3% OM.

Inzen™ (Pioneer, Johnston, IA) and a Dekalb (Monsanto Co., St. Louis, MO) conventional grain sorghum cultivar ('DKS 53-67') were planted in the PRE experiment at all locations (Figures 1–3). Cultivars were planted at 217,000 seeds ha⁻¹ to a 2.5- to 3-cm depth in both years. All plots consisted of two rows, 8.5 m long. Plots to evaluate ALS resistance PRE were arranged as a split-plot design where the whole-plot factor was ALS herbicide applied (22 herbicides), and the sub-plot factor was seed technology planted (Inzen™ or conventional). The POST experiment was planted to Inzen™ sorghum only and treated POST with multiple ALS-inhibiting herbicides. The experimental design was a randomized complete block with the fixed effect being herbicide treatment and random effects consisting of site-year and replication.

PRE herbicide applications were made immediately following planting, and POST herbicide applications were made when sorghum reached the V4 growth stage using an air-pressurized tractor-mounted spray boom at LMCRS and a CO₂-pressurized backpack sprayer at PTRS and AAREC. Both sprayers were equipped with TeeJet® Air Induction XR 110015 nozzles (TeeJet® Technologies, Glendale Heights, IL). All treatments were applied with a ground speed of 4.8 km h⁻¹. Herbicides were applied in a spray volume of 112.2 L ha⁻¹ at LMCRS and 140 L ha⁻¹ at PTRS and AAREC. Herbicide treatments and corresponding rates are listed in Table 1.

Atrazine (Aatrex; Syngenta Crop Protection, LLC, Greensboro, NC) plus S-metolachlor (Dual; Syngenta Crop Protection, LLC, Greensboro, NC) were applied at planting to help maintain weed-free plots. Any escapes from the PRE application were controlled by a POST application of the same mixture. Further escapes were hand-weeded. Recommendations from soil sample results analyzed at the soil testing and research laboratory in Marianna, AR, were followed for fertility management. Pest management decisions were based on University of Arkansas Extension recommendations (Espinoza 2015; McLeod and Greene 2015). Traditional furrow irrigation was used to provide soil moisture for all tests, except in Fayetteville where overhead irrigation was used.

Visible ratings for crop injury and sorghum heights were recorded at 2 and 4 wk after planting (WAP) and POST application (WAA). Visible ratings of herbicide application were on a 0 to 100% scale, with 0 being no injury and 100% equaling plant death (Frans et al. 1986). Five random plants in each plot were measured to estimate an average of sorghum height and then divided by the average sorghum height of nontreated plots so as to obtain relative sorghum heights. Because of an issue of sterile seed in 2016, yield data were only collected in 2017. Both rows of Inzen™ and conventional sorghum were harvested using a small-plot combine, and grain moisture was adjusted to 14%. Harvested plots were recorded as kg ha⁻¹ and converted to relative yield by dividing each plot by the average yield of the nontreated plots.

All data collected were subjected to ANOVA using JMP (JMP Pro13, SAS Institute Inc., Cary, NC), with significant means separated using Fisher's protected LSD ($\alpha = 0.05$). As the nontreated plot in each replication was used to convert sorghum height and yield to a percentage of the nontreated, data from these plots were not included in the analysis. All treatments containing Inzen™ technology were excluded from the statistical analysis of visible crop injury, because no injury was observed. Therefore, in the PRE experiment visible injury to the conventional hybrid was analyzed as a randomized complete block. All data between site-years were analyzed together, with locations considered random.

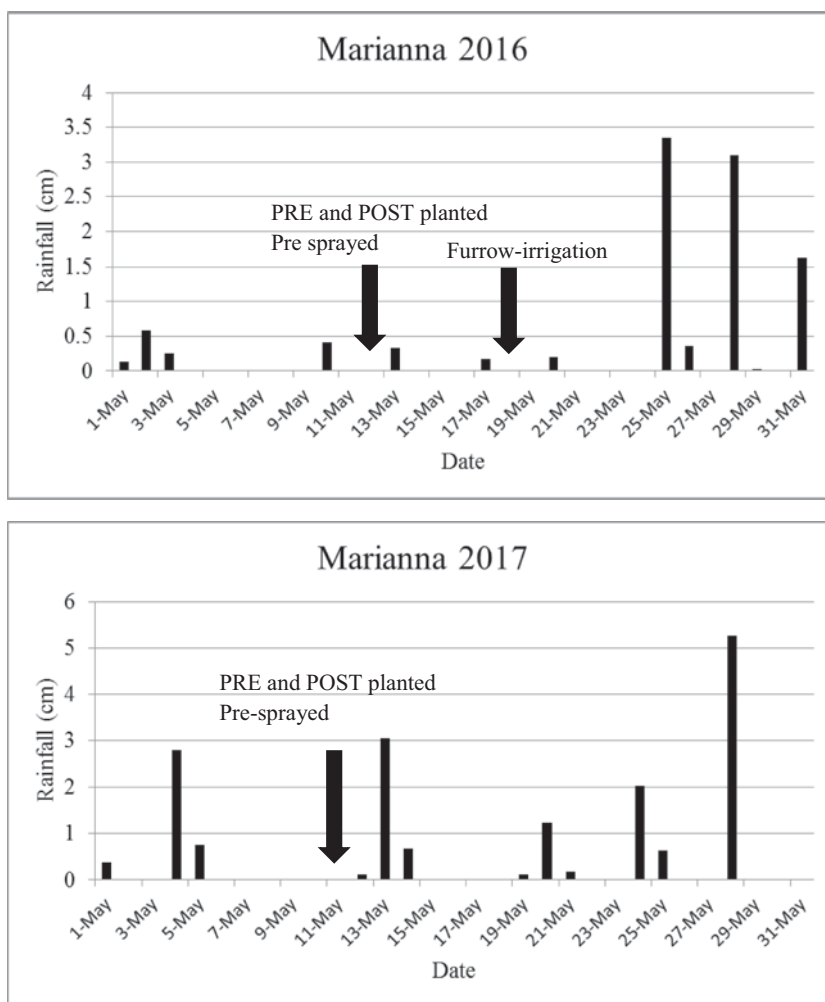


Figure 1. Planting/application timing and rainfall at Marianna, AR (LMCRS) in 2016 and 2017.

Results and Discussion

Sensitivity to PRE Applications of ALS-Inhibiting Herbicides

All experiments received adequate rainfall within 7 d of application, with the exception of LMCRS in 2016 (Figures 1–3). Soil moisture is important for activation of soil-applied herbicides (Curran 2001). Because herbicides applied to the soil are absorbed by the hypocotyl of plants during germination, the soil must receive adequate rainfall or irrigation within 7 d of application for herbicide activity. To combat the lack of rainfall, an irrigation application was applied on May 18, 2016 at LMCRS. Activation of herbicides by irrigation did not result in a difference of visible injury 2 WAP ($P = 0.1725$) or 4 WAP, ($P = 0.3930$) between locations.

For the response variable visible injury at 2 WAP, a main effect of herbicide was found ($P = 0.0309$) (Table 2). At 2 WAP, flumetsulam-methyl and bispyribac-Na caused the least injury (6%) to grain sorghum, and thiencazone-methyl caused the most injury (96%). Bispyribac-Na is used for POST control of weeds (Anonymous 2012) and does not have residual activity. Inzen™ grain sorghum at 2 WAP had a high degree of resistance to all herbicides applied based on no more than 1% injury observed (data not shown). Nicosulfuron, which is labeled in Inzen™ grain sorghum, caused 49% injury to conventional sorghum (Table 3). These results were consistent with previous studies of injury to

conventional grain sorghum applied with nicosulfuron, which showed levels ranging from 19% to 67% depending upon rate applied (Matocha and Jones 2015).

By 4 WAP, a main effect of herbicide was found ($P < 0.0001$) for the response variable visible injury (Table 2). By 4 WAP, Inzen™ grain sorghum exhibited no signs of injury from any of the herbicides (data not shown). The conventional grain sorghum seemed to recover from some of the injury by 4 WAP, similar to published results from other researchers (Matocha and Jones 2015). The conventional technology was injured only 1% by bispyribac-Na, whereas 96% injury was caused by thiencazone-methyl (Table 3). Nicosulfuron at 4 WAP resulted in 44% injury, which was similar to injury seen from trifloxysulfuron, diclosulam, and propoxycarbazine. Both rimsulfuron and imazapic caused 93% injury, which was not different from thiencazone-methyl or pyriithiobac. Observation of injury with imazethapyr (75%) and pyriithiobac (83%) is essential, because there is the potential of herbicide carryover in a crop rotation with rice or cotton (Table 3); however, no injury was observed on Inzen™ grain sorghum. These data prove that Inzen™ grain sorghum can be implemented as a safe option in a rotation that follows ALS-inhibiting herbicide applications and permits potential for weed control.

A two-way interaction between herbicide and technology was observed for sorghum height 2 WAP ($P = 0.0071$) (Table 2).

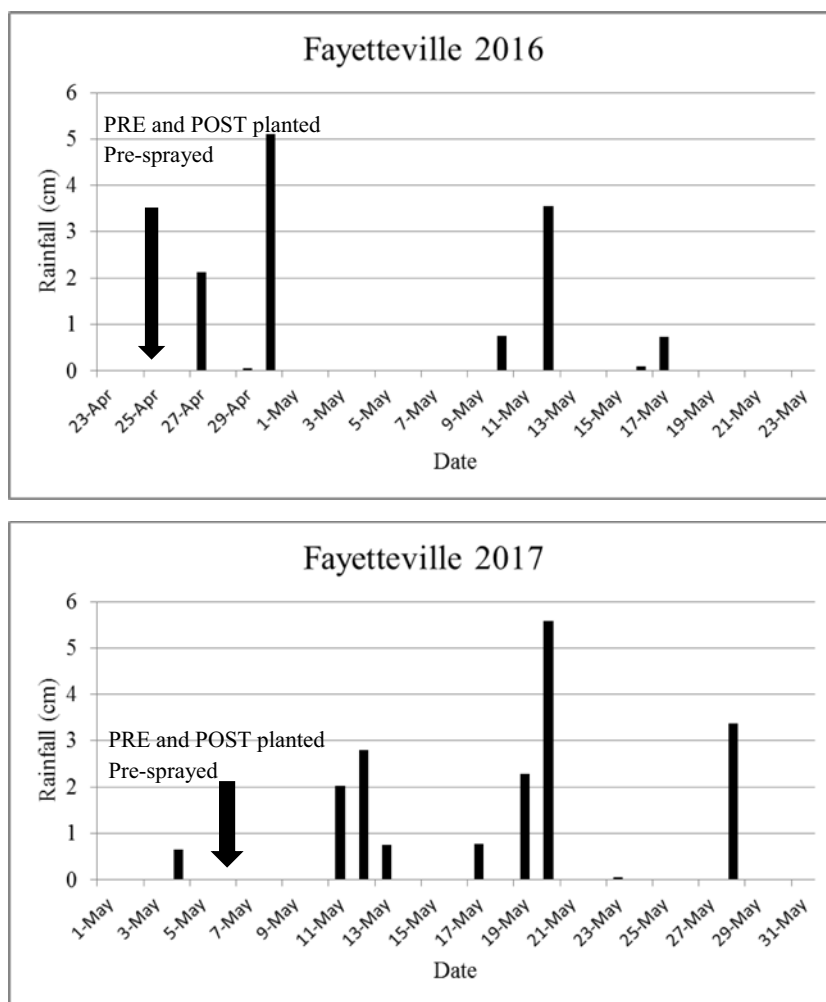


Figure 2. Planting/application timing and rainfall at Fayetteville, AR (AAREC) in 2016 and 2017.

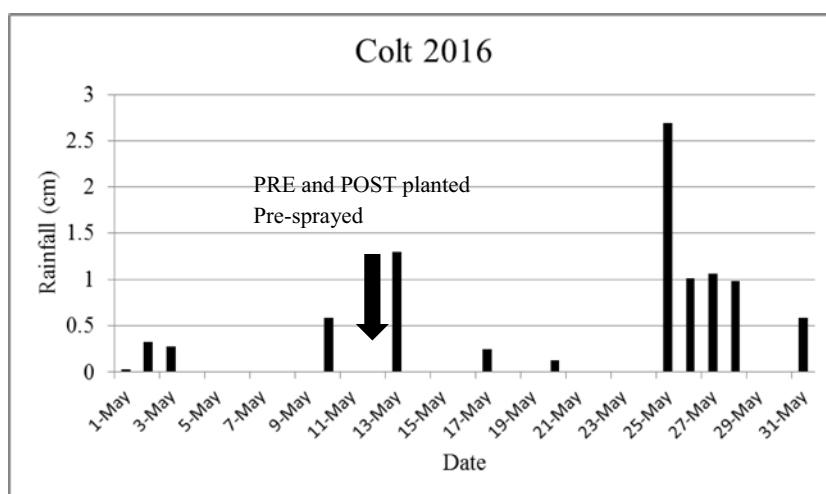


Figure 3. Planting/application timing and rainfall at Colt, AR (PTRS) in 2016.

A reduction in sorghum height was found in all plots containing the conventional sorghum 2 WAP, with the exception of imazosulfuron, imazamox, bispyribac-Na, penoxsulam-methyl, and flumetsulam-methyl. There was no reduction in Inzen™ grain sorghum height

at 2 WAP for any of the herbicides applied. Comparing Inzen™ and conventional grain sorghum within each herbicide, a sorghum height reduction in the conventional variety was found following all herbicides, except imazosulfuron, imazamox, bispyribac-Na,

Table 1. Herbicides and rates applied for PRE and POST acetolactate synthase (ALS)-inhibiting herbicide sensitivity experiments in 2016 and 2017.^{a-c}

Herbicide	Trade name	ALS herbicide family	Rate	Manufacturer	Address
			g ai or ae ha ⁻¹		
Rimsulfuron	Resolve	Sulfonylurea	17.5	DuPont	Wilmington, DE
Primisulfuron	Beacon	Sulfonylurea	40.0	Syngenta	Greensboro, NC
Nicosulfuron	Accent	Sulfonylurea	35.2	DuPont	Wilmington, DE
Trifloxysulfuron	Envoke	Sulfonylurea	7.9	Syngenta	Greensboro, NC
Chlorsulfuron + metsulfuron	Finesse	Sulfonylurea	21.9 + 4.4	DuPont	Wilmington, DE
Chlorimuron	Classic	Sulfonylurea	8.8	DuPont	Wilmington, DE
Imazosulfuron	League	Sulfonylurea	336	Valent	Walnut Creek, CA
Imazapic	Cadre	Imidazolinone	70.1	BASF	Research Triangle Park, NC
Imazethapyr	Newpath	Imidazolinone	70.1	BASF	Research Triangle Park, NC
Imazamox	Beyond	Imidazolinone	43.8	BASF	Research Triangle Park, NC
Imazaquin	Scepter	Imidazolinone	17.2	BASF	Research Triangle Park, NC
Pyrithiobac	Staple	Pyrimidinylthiobenzoic acid	58.9	DuPont	Wilmington, DE
Bispyribac-Na	Regiment	Pyrimidinylthiobenzoic acid	35.3	Valent	Walnut Creek, CA
Diclosulam	Strongarm	Triazolopyrimidine	26.5	Dow	Indianapolis, IN
Cloransulam-methyl	First Rate	Triazolopyrimidine	17.7	Dow	Indianapolis, IN
Penoxsulam-methyl	Grasp	Triazolopyrimidine	40.3	Dow	Indianapolis, IN
Flumetsulam-methyl	Python	Triazolopyrimidine	7.2	Dow	Indianapolis, IN
Thiencarbazone-methyl	Varro	Sulfonylaminocarbonyl triazolinone	28.0	Bayer	Research Triangle Park, NC
Propoxycarbazone	Olympus	Sulfonylaminocarbonyl triazolinone	44.2	Bayer	Research Triangle Park, NC
Flucarbazone	Everest	Sulfonylaminocarbonyl triazolinone	15.3	Arysta	Cary, NC

^aSensitivity experiment conducted near Colt, AR, in 2016.^bSensitivity experiment conducted near Marianna, AR, in 2016 and 2017.^cSensitivity experiment conducted in Fayetteville, AR, in 2016 and 2017.**Table 2.** ANOVA for PRE acetolactate synthase-inhibiting herbicides to grain sorghum from 2016 and 2017.^{a-d}

Variable	Source	DF	F ratio	P value ^e
Visible injury 2 WAP (%)	Herbicide	20	2.3597	0.0309*
Sorghum heights 2 WAP (cm)	Herbicide	20	2.3597	0.0309*
	Hybrid	1	4.1527	0.2904
	Herbicide × hybrid	20	3.1191	0.0071*
Visible injury 4 WAP (%)	Herbicide	20	63.8662	<0.0001*
Sorghum heights 4 WAP (cm)	Herbicide	20	4.7759	0.0005*
	Hybrid	1	5.2362	0.2623
	Herbicide × hybrid	20	3.5162	0.0035*
Relative yield (%)	Herbicide	20	45.1931	<0.0001*
	Hybrid	1	871.9586	<0.0001*
	Herbicide × hybrid	20	60.2547	<0.0001*

^aSensitivity experiment conducted near Colt, AR, in 2016.^bSensitivity experiment conducted near Marianna, AR, in 2016 and 2017.^cSensitivity experiment conducted in Fayetteville, AR, in 2016 and 2017.^dAbbreviations: DF, degrees of freedom; WAP, weeks after planting.^e*Denotes significance.

penoxsulam-methyl, and flumetsulam-methyl (Table 4). Sorghum height was influenced by the interaction of herbicide applied and technology planted 4 WAP ($P = 0.0035$) (Table 2). Plots containing the conventional grain sorghum, 4 WAP, had a reduction in sorghum heights ranging from 1% to 68%. The greatest sorghum height reductions in the conventional sorghum were observed when pyriithiobac, imazapic, thiencarbazone-methyl, and rimsulfuron were applied (Table 4). No height reductions were seen in plots containing Inzen™ grain sorghum.

No reduction in relative yield was found in Inzen™ plots. Differences in relative yield were found in plots of the conventional grain sorghum ($P < 0.0001$) with all herbicides except trifloxysulfuron, imazosulfuron, propoxycarbazone, flucarbazone, penoxsulam-methyl, and flumetsulam-methyl (Tables 2 and 5). When comparing technologies, a difference between conventional and Inzen™ grain sorghum was found with 13 of the herbicides

Table 3. Visible injury (%) from PRE applications of acetolactate synthase-inhibiting herbicides to conventional grain sorghum in 2016 and 2017.^{a-d}

Herbicide	Rate	Injury ^e	
		2 WAP	4 WAP
	g ai or ae ha ⁻¹	%	
Rimsulfuron	17.5	91	93
Primisulfuron	40.0	60	58
Nicosulfuron	35.2	49	44
Trifloxysulfuron	7.9	43	36
Chlorsulfuron + metsulfuron	21.9 + 4.4	37	29
Chlorimuron	8.8	32	22
Imazosulfuron	336	12	1
Imazapic	70.1	87	93
Imazethapyr	70.1	67	75
Imazamox	43.8	25	20
Imazaquin	17.2	21	13
Pyrithiobac	58.9	79	83
Bispyribac-Na	35.3	6	1
Diclosulam	26.5	48	42
Cloransulam-methyl	17.7	21	12
Penoxsulam-methyl	40.3	12	3
Flumetsulam-methyl	7.2	6	1
Thiencarbazone-methyl	28.0	96	99
Propoxycarbazone	44.2	46	38
Flucarbazone	15.3	20	10
LSD (0.05) ^f		10	12

^aSensitivity experiment conducted near Colt, AR, in 2016.^bSensitivity experiment conducted near Marianna, AR, in 2016 and 2017.^cSensitivity experiment conducted in Fayetteville, AR, in 2016 and 2017.^dAbbreviations: WAP, weeks after planting.^eVisual injury ratings were conducted on a scale of 0 to 100%, with 0% being no injury and 100% denoting total plant death.^fMeans within a column can be compared using Fisher's protected LSD ($\alpha = 0.05$).

tested (Table 5). These results for Inzen™ grain sorghum are consistent with those of Werle et al. (2016), where the double point mutation of Val₅₆₀Ile and Trp₅₇₄Leu provided broad cross-resistance to ALS-inhibiting herbicides.

Table 4. Grain sorghum heights at 2 and 4 wk following PRE acetolactate synthase-inhibiting herbicide applications in Fayetteville, AR, and near Marianna, AR, in 2017.

		Sorghum height cm				
		2 WAP ^{a,b}		4 WAP ^{c,d}		
Herbicide	Rate	Inzen™	Conventional	Inzen™	Conventional	
	g ai or ae ha ⁻¹	% of nontreated				
Rimsulfuron	17.5	87	1	98	1	
Primisulfuron	40.0	106	51	109	53	
Nicosulfuron	35.2	104	44	108	94	
Trifloxysulfuron	7.9	102	61	111	68	
Chlorsulfuron + metsulfuron	21.9 + 4.4	96	49	100	37	
Chlorimuron	8.8	102	47	102	51	
Imazosulfuron	336	103	94	109	95	
Imazapic	70.1	91	25	95	16	
Imazethapyr	70.1	100	36	104	27	
Imazamox	43.8	102	80	107	85	
Imazaquin	17.2	108	73	111	65	
Pyriathiobac	58.9	92	26	99	16	
Bispyribac-Na	35.3	106	95	104	81	
Diclosulam	26.5	102	59	102	59	
Cloransulam-methyl	17.7	102	52	105	85	
Penoxsulam-methyl	40.3	100	89	103	89	
Flumetsulam-methyl	7.2	102	98	104	98	
Thiencarbazone-methyl	28.0	103	4	108	4	
Propoxycarbazone	44.2	97	54	100	90	
Flucarbazone	15.3	104	64	108	85	
LSD (0.05) ^f		24	24	20	16	20

^aAverage sorghum height of Inzen™ grain sorghum in nontreated plots was 32 cm at 2 WAP and 40 cm in nontreated conventional plots.

^bWhole-plot LSD for herbicide at 2 WAP was 24 and sub-plot LSD for seed technology planted was 24.

^cAverage sorghum height of Inzen™ grain sorghum in nontreated plots was 45 cm at 4 WAP and 54 cm in nontreated conventional plots.

^dWhole-plot LSD for herbicide at 4 WAP was 16 and sub-plot LSD for seed technology planted was 20.

Table 5. Relative yield from 2017 of Inzen™ and conventional grain sorghum following PRE acetolactate synthase-inhibiting herbicide applications in Fayetteville, AR, and near Marianna, AR.

Herbicide	Rate	Grain yield ^a	
		Inzen TM _b	Conventional ^c
		% of nontreated	
	g ai or ae ha ⁻¹		
Rimsulfuron	17.5	109	1
Primisulfuron	40.0	104	45
Nicosulfuron	35.2	104	58
Trifloxysulfuron	7.9	98	69
Chlorsulfuron + metsulfuron	21.9 + 4.4	104	39
Chlorimuron	8.8	107	43
Imazosulfuron	336	97	81
Imazapic	70.1	106	0
Imazethapyr	70.1	96	12
Imazamox	43.8	101	52
Imazaquin	17.2	109	42
Pyriithiobac	58.9	93	40
Bispyribac-Na	35.3	95	50
Diclosulam	26.5	104	63
Cloransulam-methyl	17.7	100	71
Penoxsulam-methyl	40.3	95	93
Flumetsulam-methyl	7.2	105	100
Thiencarbazone-methyl	28.0	99	1
Propoxycarbazone	44.2	115	84
Flucarbazone	15.3	118	98
LSD (0.05) ^d		12	8
			12

^aWhole-plot LSD for herbicide was 8, and sub-plot LSD for seed technology planted was 12.

^bNontreated plots containing the Inzen™ technology yielded 8,480 kg ha⁻¹.

^cNontreated plots containing the conventional technology yielded 7,828 kg ha⁻¹.

^dMeans within the LSD are not significantly different according to Fisher's protected LSD ($\alpha = 0.05$).

Table 6. ANOVA of Inzen™ grain sorghum from POST-applied acetolactate synthase-inhibiting herbicides in 2016 and 2017.^{a-d}

Variable	Source	DF	F ratio	P value ^e
Sorghum heights 2 WAA (cm)	Herbicide	20	72.6881	<0.0001*
Sorghum heights 4 WAA (cm)	Herbicide	20	5.4095	<0.0001*
Relative yield (%)	Herbicide	20	0.8829	0.6118

^aSensitivity experiment conducted near Colt, AR, in 2016.

^bSensitivity experiment conducted near Marianna, AR, in 2016 and 2017.

^cSensitivity experiment conducted in Fayetteville, AR, in 2016 and 2017.

^dAbbreviations: DF, degrees of freedom; WAA, weeks after application.

^e*Denotes significance.

POST Inzen™ Grain Sorghum Evaluation

No visible injury, sorghum height reduction, or yield loss occurred in the Inzen™ sorghum treated with any of the PRE-applied ALS-inhibiting herbicides evaluated. However, bispyribac-Na POST caused a 12% sorghum height reduction and 35% yield loss in Inzen™ sorghum, but this yield loss did not represent a statistically significant impact on the crop (Table 6). These results are consistent with the Regiment® (bispyribac-Na) label, which states that typical ALS injury may be observed as early as 3 to 7 d after application. Injury from bispyribac results in temporary yellowing and a reduction in sorghum height (Anonymous 2012). By 4 WAA, no visible injury was observed in plots treated with bispyribac-Na.

Grain sorghum heights were reduced 28% by bispyribac-Na 2 WAA, and 12% by 4 WAA. All other plots contained plants with similar heights to the nontreated (Table 7).

Table 7. Relative sorghum heights of Inzen™ grain sorghum to POST-applied acetolactate synthase-inhibiting herbicides in 2017 at Fayetteville, AR, and near Marianna, AR.^a

Herbicide	Rate g ai or ae ha ⁻¹	Sorghum heights	
		2 WAA ^b	4 WAA ^c
		—% of nontreated—	
Rimsulfuron	17.5	103	99
Primisulfuron	40.0	103	99
Nicosulfuron	35.2	102	101
Trifloxysulfuron	7.9	102	99
Chlorsulfuron + metsulfuron	21.9 + 4.4	103	100
Chlorimuron	8.8	103	100
Imazosulfuron	336	103	99
Imazapic	70.1	103	99
Imazethapyr	70.1	104	101
Imazamox	43.8	103	100
Imazaquin	17.2	103	101
Pyrithiobac	58.9	103	100
Bispyribac-Na	35.3	72	88
Diclosulam	26.5	103	100
Cloransulam-methyl	17.7	103	101
Penoxsulam-methyl	40.3	102	100
Flumetsulam-methyl	7.2	102	100
Thiencarbazone-methyl	28.0	102	101
Propoxycarbazone	44.2	103	99
Flucarbazone	15.3	103	101
LSD (0.05) ^d		2	3

^aAbbreviations: WAA, weeks after application.^bAverage sorghum height of nontreated plots was 82 cm at 2 WAA.^cAverage sorghum height of nontreated plots was 93 cm at 4 WAA.^dMeans within the LSD are not significantly different according to Fisher's protected LSD ($\alpha = 0.05$).

Grain yield was not affected by ALS-inhibiting herbicide when applied to Inzen™ grain sorghum ($P = 0.6118$) (Table 6). Cross-resistance among ALS-inhibiting herbicides to Inzen™ sorghum is consistent with previous research that documented resistance to both sulfonylurea and imidazolinone herbicides when the double gene mutation of Val₅₆₀Ile and Trp₅₇₄Leu was present (Werle et al 2017).

Lack of visible injury, and reduction of sorghum height or yield with Inzen™ grain sorghum can be attributed to the ALS double gene mutation of Val₅₆₀Ile and Trp₅₇₄Leu. Trp₅₇₄ is the second most documented ALS gene mutation and has been identified in 41 weed species. The specific substitution of Trp₅₇₄Leu is the most documented and accounts for 38 of the 41 Trp₅₇₄ substitutions. In all 38 documented cases of Trp₅₇₄Leu, resistance to two or more of the five ALS families was confirmed (Heap 2020). Palmer amaranth, barnyardgrass, and johnsongrass account for 3 of these 38 cases and are among the top five most problematic weeds in many Arkansas row crops (Hernandez et al. 2015; Molin et al. 2016; Panozzo et al. 2013; Singh et al. 2019).

Practical Implications

As many weed species can evolve resistance to a specific site of action, it is important to note that within WSSA Group 2, resistance to a specific ALS-inhibiting herbicide does not necessarily constitute resistance to all ALS-inhibiting herbicides. The broad cross-resistance to multiple ALS-inhibiting herbicides seen in Inzen™ could potentially allow for these products to be used within the crop for weed control, along with reducing current plant-back intervals for grain sorghum to WSSA Group 2 herbicides.

These results may prove beneficial when developing a herbicide program for grain sorghum, as there is potential to incorporate

other herbicides into the program, depending on weed species present. By enabling use of the ALS site of action, Inzen™ will allow more options for herbicide diversification, further delaying the development of weed resistance. However, Werle et al. (2016) confirmed cross-resistance to nicosulfuron and imazethapyr in populations of johnsongrass present in Kansas, Missouri, and Nebraska. The possibility of ALS-resistant johnsongrass spreading further emphasizes the necessity of proper stewardship of the Inzen™ technology.

Acknowledgments. Funding for this research was provided by the Arkansas Corn and Grain Sorghum Board. No conflicts of interest have been declared.

References

- Anonymous (2012) Regiment® Herbicide. Valent BioSciences LLC. <https://www.valent.com>. Accessed: March 30, 2019
- Anonymous (2016) Zest™ WDG. Corteva AgriScience. <https://www.corteva.us>. Accessed: March 28, 2019
- Bendixen LE (1986) Corn (*Zea mays*) yield in relationship to johnsongrass (*Sorghum halepense*) population. Weed Sci 34:449–451
- Bowers JE, Abbey C, Anderson S, Chang C, Draye X, Hoppe AH, Jessup R, Lemke C, Lennington J, Li Z, Lin Y, Liu S, Luo L, Marler BS, Ming R, Mitchell SE, Oiang D, Reischmann K, Schulze SR, Skinner DN, Wang Y, Kresovich S, Schertz KF, Paterson AH (2003) A high-density genetic recombination map of sequence-tagged sites for sorghum, as a framework for comparative structural and evolutionary genomics of tropical grains and grasses. Genetics 165:367–386
- Curran WS (2001) Persistence of herbicides in soil. Penn State Extension. Agronomy Facts 36:1–4. <http://extension.psu.edu/pests/weeds/control/persistence-of-herbicides-in-soil>. Accessed: May 28, 2019
- Espinoza L (2015) Fertilization and Liming. Arkansas Grain Sorghum Production Handbook. MP297:21–24. Little Rock, AR: University of Arkansas Cooperative Extension Service
- Frans RR, Talbert MD, Crowley H (1986) Experimental design and techniques for measuring and analyzing plant responses to weed control practices. Page 29 in Camper ND, ed, Research Methods in Weed Science. Champaign, IL: Southern Weed Science Society
- Hall LM, Holtum JAM, Powles SB (1994) Mechanisms responsible for cross resistance and multiple resistance. Pages 243–262 in Powles SB and Holtum JAM, eds, Herbicide Resistance in Plants: Biology and Biochemistry. Boca Raton, FL: CRC Press
- Heap I (2020) International Herbicide-Resistant Weed Database. www.weedscience.org. Accessed: February 13, 2020
- Hernandez MJ, Leon R, Fischer AJ, Gebauer M, Galdames R, Flgurea R (2015) Target-site resistance to nicosulfuron in johnsongrass (*Sorghum halepense*) from Chilean corn fields. Weed Sci 63:631–640
- Horowitz M (1973) Spatial growth of *Sorghum halepense*. Weed Res 13:200–208
- Langemeier MA, Witt WW (1986) Johnsongrass (*Sorghum halepense*) control in reduced-tillage systems. Weed Sci 25:264–267
- Mallory-Smith CA, Thill DC, Dial MJ (1990) Identification of sulfonylurea herbicide-resistant prickly lettuce (*Lactuca serriola*). Weed Technol 4:163–168
- Masson JA, Webster EP, Williams BJ (2001) Flood depth, application timing, and imazethapyr activity in imidazolinone-tolerant rice (*Oryza sativa*). Weed Technol 15:315–319
- Matocha MA, Jones CA (2015) Effect of carrier volume on grain sorghum response to simulated drift of nicosulfuron. Weed Technol 29:684–688
- McLeod P, Greene J (2015) Major insect pests of grain sorghum in Arkansas and their management. Pages 25–36 in Espinoza L, Kelley J, eds, Arkansas Grain Sorghum Production Handbook. MP297. Little Rock, AR: University of Arkansas Cooperative Extension Service
- McWhorter CG (1977) Johnsongrass control in soybeans with soil-incorporated dinitroaniline herbicides. Weed Sci 25:264–267
- Molin WT, Nandula VK, Wright AA, Bond JA (2016) Transfer and expression of ALS inhibitor resistance from Palmer amaranth (*Amaranthus palmeri*) to an *A. spinosus* × *A. palmeri* hybrid. Weed Sci 64:240–247

- Mueller JP, Lewis WM, Green JT, Burns JC (1993) Yield and quality of silage corn as altered by johnsongrass infestation. *Agron J* 85:49–52
- Owen MJ, Powles SB (2010) Glyphosate-resistant rigid ryegrass (*Lolium rigidum*) population in the western Australian grain belt. *Weed Technol* 24:44–49
- Panozzo S, Scarabel L, Tranel PJ, Sattin M (2013) Target-site resistance to ALS inhibitors in the polyploid species *Echinochloa crus-galli*. *Pesticide Biochem Physiol* 105:93–101
- Riley DG, Shaw DR (1988) Influence of imazethapyr on the control of pitted morningglory (*Ipomoea lacunosa*) and johnsongrass (*Sorghum halepense*) with chlorimuron, imazaquin, and imazethapyr. *Weed Sci* 36: 663–666
- Singh SV, Salas-Perez RA, Bagavathiannan MV, Lawton-Rauh A, Roma-Burgos N (2019) Target-site mutation accumulation among ALS inhibitor-resistant Palmer amaranth. *Pest Manag Sci* 75:1131–1139
- Smith K, Scott B (2015) Weed control in grain sorghum. Pages 47–49 in Espinoza L, Kelley J, eds, *Arkansas Grain Sorghum Production Handbook*. MP297. Little Rock, AR: University of Arkansas Cooperative Extension Service
- Tranel PJ, Wright TR (2002) Resistance of weeds to ALS-inhibiting herbicides: what have we learned? *Weed Sci* 50:700–712
- Tranel PJ, Wright TR, Heap IM (2016) Mutations in herbicide-resistant weeds to ALS inhibitors. <http://www.weedscience.org/Mutations/MutationDisplayALL.aspx>. Accessed: October 6, 2017
- Tuinstra MR, Al-Khatib K, inventors; assignee (2008) Acetolactate synthase herbicide resistant sorghum. International patent application WO 2008073800. Patentlaan, NL World Intellectual Property Organization
- Werle R, Begcy K, Yerka MK, Mower JP, Dweikat I, Jhala AJ, Lindquist JL (2017) Independent evolution of acetolactate synthase-inhibiting herbicide resistance in weedy sorghum populations across common geographic regions. *Weed Sci* 65:164–176
- Werle R, Jhala A, Yerka MK, Dille JA, Lindquist JL (2016) Distribution of herbicide-resistant shattercane and johnsongrass populations in sorghum production areas of Nebraska and northern Kansas. *Agron J* 108:321–328