BioOne COMPLETE

Patterns of Diversity and Abundance of Carrion Insect Assemblages in the Natural Park "Hoces del Río Riaza" (Central Spain)

Authors: Baz, Arturo, Cifrián, Blanca, and Martín-Vega, Daniel

Source: Journal of Insect Science, 14(162) : 1-10

Published By: Entomological Society of America

URL: https://doi.org/10.1093/jisesa/ieu024

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at www.bioone.org/terms-of-use.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

Downloaded From: https://complete.bioone.org/journals/Journal-of-Insect-Science on 28 Mar 2024
Terms of Use: https://complete.bioone.org/terms-of-use

RESEARCH

Patterns of Diversity and Abundance of Carrion Insect Assemblages in the Natural Park "Hoces del Río Riaza" (Central Spain)

Arturo Baz,[1,2] Blanca Cifrián,[1] and Daniel Martín-Vega[1]

[1]Departamento de Ciencias de la Vida, Universidad de Alcalá, Ctra. Madrid-Barcelona km 33.6, 28871 Alcalá de Henares, Madrid, Spain
[2]Corresponding author, e-mail: arturo.baz@uah.es

Subject Editor: Robert Jetton

J. Insect Sci. 14(162): 2014; DOI: 10.1093/jisesa/ieu024

ABSTRACT. The patterns of diversity and abundance of the carrion insect species in the different habitats of the Natural Park "Hoces del Río Riaza" (central Spain) were studied with the use of carrion-baited traps. Representativeness of the inventories was assessed with the calculation of randomized species richness curves and nonparametric estimators. Coleoptera families, Silphidae and Dermestidae, and Diptera families, Calliphoridae and Muscidae, were dominant in every sampling habitat, but differences in the patterns of diversity and abundance were found. Lusitanian oakwood and riparian forest were the most diverse habitats with high abundance of saprophagous species, whereas more open (i.e., exposed to continuous sunlight during the day) habitats showed lower diversity values and a different species composition and distribution of species abundance, favoring thermophilous species and necrophagous species with high tolerance to different environmental conditions. Differences in the bioclimatical features of the sampled habitats are suggested to explain the composition and diversity of the carrion insect assemblages in different environments.

Key Words: Carrion insect, diversity, assemblage, Spain

Every natural environment contains three kinds of organisms whose interactions keep the ecosystem in working order: producers, consumers, and decomposers. The last group includes those organisms that feed on organic fecal matter as well as decaying plant and animal remains. Decomposing organic matter such as carrion is an ephemeral but valuable food source for different organisms (Barton et al. 2013), among which arthropods represent the most diverse and abundant group associated with this kind of resource (Braack 1987). Carrion supports high-diverse communities of coexisting insect species, which exploit the same resource (Woodcock et al. 2002); nevertheless, insects visiting carrion can exploit this resource in many different ways. In this sense, Leclercq (1978) and Braack (1987) distinguished four categories of carrion-attendant arthropods according to their kind of use of such resource: 1) necrophagous, i.e., those species that feed directly on carcasses and usually complete their life cycle on them, including both sarcophagous (those consuming flesh and soft tissues) and dermatophagous species (those consuming skin and dry tissues); 2) necrophilous, i.e., those species that are predators of necrophagous insects (mainly Diptera larvae), including also parasitoid species; 3) saprophagous, i.e., those feeding on decomposing organic matter but usually not completing their life cycle on a carcass; and 4) opportunistic or casual, i.e., those using the carcass as a refuge or those having a nonnecrophagous diet but occasionally obtaining nutrients from carrion as a food supplement. Moreover, several studies have shown habitat to affect the species composition and succession patterns of insects associated with carrion (Goff 1991, Anderson and VanLaerhoven 1996). Also, species abundances differ among habitats (Martín-Vega and Baz 2012, 2013), so each habitat can be typified by different dominant species.

A correct management of natural resources involves their study to obtain a better knowledge of their dynamics and composition. This article assesses the species richness and diversity patterns of carrion insects in a protected natural area in central Spain. The representativeness of the inventories, the species composition, and the observed diversity patterns within the different habitats of the study area are considered and discussed.

Materials and Methods

The aim of this study was to invent the sarcosaprophagous insect fauna of the Natural Park "Hoces del Río Riaza" (Segovia Province, central Spain), a protected area of 5,185 Ha around the Riaza river. The main vegetation of the park consists in riparian forests, although it can also be found in other types of forests. Concretely, there are five main representative habitat types in the park: 1) herbaceous and shrub vegetation, 2) juniper forest (*Juniperus thurifera L.*), 3) holm oakwood (*Quercus ilex L.* ssp. *ballota (Desf.)Samp*), 4) Lusitanian oakwood (*Quercus faginea Lamarck*), and 5) riparian forest (*Populus nigra L.*). Two sites for each habitat were selected, resulting in a total of 10 sampling sites (Fig. 1). Detailed information on the locations and bioclimatical features of the sampling sites are listed in Table 1. Climatic data were obtained from Ninyerola et al. (2005), and they represent the annual maximum, minimum, and mean temperature, as well as the total rainfall (Table 1). Moreover, a thermometer was installed next to each trap, registering minimum and maximum temperatures between surveys. Average minimum and maximum temperature and the mean temperature during the whole sampling period as registered by thermometers are listed in Table 1.

Sarcosaprophagous insects were collected using carrion-baited traps, modified from the design of Morón and Terrón (1984) as described by Baz et al. (2007). Traps were baited with squid, which has been shown to be very effective in attracting necrophagous insects (Newton and Peck 1975), semiburied in the ground, and protected from vertebrate scavengers with a perimeter of stones. One carrion-baited trap was installed at each site. Traps were operated 15 d each month from June to September 2007; thus, a total of 36 samples were obtained at the end of the survey as 4 samples were destroyed, probably by wild animals. Moreover, to determine whether some species were accidentally collected by the traps and not attracted to carrion, an identical but unbaited trap was installed next to each carrion-baited trap. The collected specimens were either preserved in 80% ethanol or oven dried and pinned, and deposited in the collection of the Department of Life Sciences of the University of Alcalá. Collected nonsarcosaprophagous insects (i.e., opportunistic or casual species) have not been considered

© The Author 2014. Published by Oxford University Press on behalf of the Entomological Society of America.
This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/by-nc/4.0/), which permits non-commercial re-use, distribution, and reproduction in any medium, provided the original work is properly cited. For commercial re-use, please contact journals.permissions@oup.com

Downloaded From: https://complete.bioone.org/journals/Journal-of-Insect-Science on 28 Mar 2024
Terms of Use: https://complete.bioone.org/terms-of-use

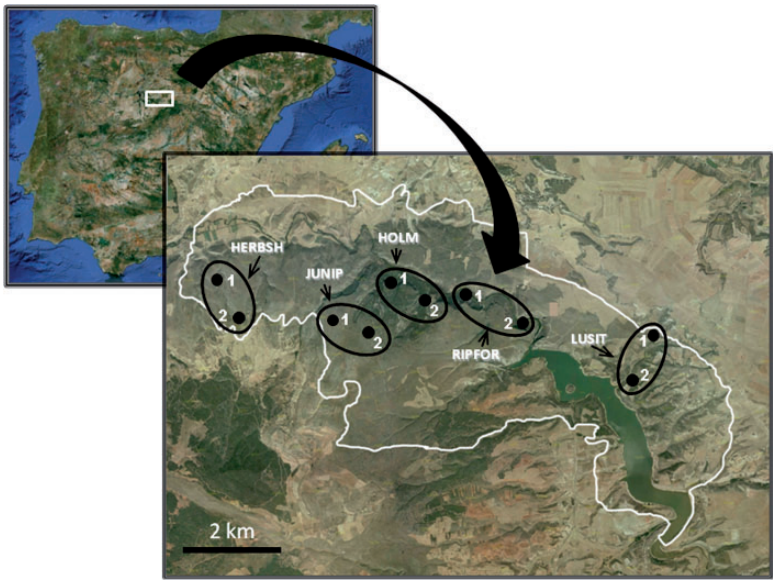


Fig. 1. Map showing the study area with an indication of the sampling sites. HERBSH, herbaceous and shrub vegetation; JUNIP, juniper forest; HOLM, holm oakwood; LUSIT, Lusitanian oakwood; RIPFOR, riparian forest.

Table 1. Description of the localities sampled

Habitat	UTM coordinates	Elevation (m)	Temp (°C) mean	Temp (°C) max	Temp (°C) min.	Total rainfall	Temp *T* (°C) mean	Temp *T* (°C) max	Temp *T* (°C) min.
HERBSH-1	30T 446440, 4599328	996	12	18	5	521	21.77	38.77	4.78
HERBSH-2	30T 446756, 4598380	1,050	11	18	5	451	21.85	39.71	4
JUNIP-1	30T 449536, 4598040	1,081	11	17	5	517	22	40.7	3.3
JUNIP-2	30T 449212, 4598128	1,057	11	17	5	499	22.32	40.07	4.57
HOLM-1	30T 450328, 4599426	901	12	18	6	520	24.4	43	5.8
HOLM-2	30T 451196, 4599034	902	12	18	6	521	22.8	40.1	5.5
LUSIT-1	30T 456652, 4598028	973	12	18	5	454	22.69	40.38	5
LUSIT-2	30T 456171, 4597179	930	12	18	5	453	21.33	37.75	4.92
RIPFOR-1	30T 452051, 4598944	903	12	18	5	476	21.88	38	5.77
RIPFOR-2	30T 453316, 4598375	899	12	18	6	468	22.32	38.21	6.43

Average minimum and maximum temperature and the mean temperature registered by installed thermometers (temp *T*) are also included. The numbers 1 and 2 indicate the two sites selected for each habitat. HERBSH, herbaceous and shrub vegetation; JUNIP, juniper forest; HOLM, holm oakwood; LUSIT, Lusitanian oakwood; RIPFOR, riparian forest.

in this study, but their captures were discussed in a previous article (Baz et al. 2010).

Species richness was estimated in every sampling site to determine whether the observed species richness was representative enough of the total number of species. Species richness was estimated from the samples by nonparametric methods (Colwell and Coddington 1994). Six nonparametric estimators were used: abundance-based coverage estimator (ACE), incidence-based coverage estimator (ICE), jackknife 1, jackknife 2, Chao 1, and Chao 2. ACE is based on the proportion of abundant species (those with >10 collected individuals) and rare species (those with <10 collected individuals), whereas ICE is based on the proportion of frequent species (those collected in >10 sampling units) and infrequent species (those collected in <10 sampling units; Magurran 2004). Jackknife 1 estimator is based on the number of species that occur in only one sample (uniques), whereas jackknife 2 and Chao 2 estimators are based on the number of species that occur in only one sample (uniques), as well as the number that occur in exactly two samples (duplicates; Colwell and Coddington 1994). On the other hand, Chao 1 estimator is based on the number of observed species that are represented by only one individual (singletons) and the number that occur with exactly two individuals (doubletons; Colwell and Coddington 1994). Randomized accumulation curves of observed species richness using the Mao Tau estimator and different estimators were calculated for each sampling site using EstimateS software (Colwell 2005). One hundred randomizations were always used. Inventory completeness, defined as observed species richness in relation with estimated richness (Cardoso et al. 2009), was calculated using nonparametric estimations.

In addition to species richness, species diversity was also measured using Shannon diversity index and Simpson diversity index. Both indices combine information on richness and relative abundance in different ways, and they were also computed using EstimateS software (Colwell 2005). The distribution of the abundances of collected species was logarithmically depicted. A cluster analysis was performed to study the degree of association among habitats according to their species composition and the average number of collected individuals in each survey. Data were transformed using the log $(x + 1)$ prior to analysis, and a dendrogram was computed using the Squared Euclidean distance and the complete linkage method. Cluster analysis was performed using the software Statgraphics Plus 5.1 (Statistical Graphic Corp. 1994–2000 Princeton, New Jersey).

Downloaded From: https://complete.bioone.org/journals/Journal-of-Insect-Science on 28 Mar 2024
Terms of Use: https://complete.bioone.org/terms-of-use

Downloaded From: https://complete.bioone.org/journals/Journal-of-Insect-Science on 28 Mar 2024
Terms of Use: https://complete.bioone.org/terms-of-use

Table 2. List of carrion Coleoptera species collected and abundances by sampling site and habitat

Coleoptera species	HERBSH-1	HERBSH-2	**Total HERBSH**	JUNIP-1	JUNIP-2	**Total JUNIP**	HOLM-1	HOLM-2	**Total HOLM**	LUSIT-1	LUSIT-2	**Total LUSIT**	RIPFOR-1	RIPFOR-2	**Total RIPFOR**	**Total Captures**
Silphidae																
T. rugosus (L. 1758)	662	688	**1,350**	1,268	792	**2,060**	1,387	1,183	**2,570**	135	37	**172**	584	597	**1,181**	**7,333**
T. ruficornis (Küster 1851)	359	236	**595**	368	248	**616**	237	275	**512**	16	19	**35**	64	63	**127**	**1,885**
Thanatophilus sinuatus (F. 1775)	3	9	**12**	1	1	**2**	6	19	**25**	11	0	**11**	45	24	**69**	**119**
Nicrophorus interruptus Stephens 1830	1	0	**1**	0	2	**2**	6	17	**23**	1	1	**2**	13	21	**34**	**62**
Nicrophorus vestigator Herschel 1807	3	8	**11**	3	5	**8**	7	3	**10**	9	4	**13**	22	10	**32**	**74**
Nicrophorus humator (Gleditsch 1767)	0	7	**7**	3	2	**5**	5	15	**20**	0	1	**1**	3	2	**5**	**38**
Nicrophorus vespillo (L. 1758)	0	1	**1**	1	0	**1**	0	6	**6**	0	0	**0**	3	1	**4**	**12**
Necrodes littoralis (L. 1758)	0	0	**0**	0	1	**1**	0	0	**0**	0	0	**0**	0	0	**0**	**1**
Silpha tristis Illiger 1798	0	0	**0**	0	0	**0**	0	0	**0**	0	0	**0**	9	0	**9**	**9**
Silpha puncticollis Lucas 1846	0	0	**0**	0	0	**0**	0	0	**0**	0	0	**0**	5	0	**5**	**5**
Dermestidae																
D. frischi Kugelann 1792	212	664	**876**	379	451	**830**	105	139	**244**	104	78	**182**	74	46	**120**	**2,252**
D. undulatus Brahm 1790	7	50	**57**	17	15	**32**	55	82	**137**	84	316	**400**	165	164	**329**	**955**
Dermestes mustelinus Erichson 1846	0	3	**3**	0	3	**3**	0	0	**0**	0	3	**3**	9	0	**9**	**18**
Dermestes hispanicus Kalik 1952	2	2	**4**	0	1	**1**	0	0	**0**	0	2	**2**	0	1	**1**	**8**
Dermestes olivieri Lepesme 1939	1	2	**3**	0	0	**0**	0	0	**0**	0	0	**0**	0	0	**0**	**3**
Nitidulidae																
Nitidula flavomaculata Rossi 1790	0	10	**10**	7	7	**14**	0	0	**0**	0	0	**0**	2	0	**2**	**26**
Nitidula carnaria (Schaller 1783)	28	31	**59**	10	20	**30**	60	54	**114**	38	19	**57**	37	23	**60**	**310**
Omosita discoidea (F. 1775)	0	0	**0**	0	0	**0**	0	0	**0**	0	1	**1**	1	1	**2**	**3**
Soronia oblonga Brisout de Barneville 1863	0	0	**0**	0	0	**0**	0	0	**0**	0	0	**0**	1	0	**1**	**1**
Catopidae																
Catops coracinus Kellner 1846	0	0	**0**	4	1	**5**	4	18	**22**	41	5	**46**	15	186	**201**	**274**
Catops fuliginosus Erichson 1837	0	0	**0**	0	0	**0**	5	0	**5**	0	2	**2**	0	0	**0**	**7**
Catops nitidicollis Kraatz 1856	0	0	**0**	0	0	**0**	2	0	**2**	0	0	**0**	0	0	**0**	**2**
Ptomaphagus subvillosus (Goeze 1777)	0	2	**2**	1	0	**1**	13	1	**14**	9	13	**22**	37	3	**40**	**79**
Ptomaphagus tenuicornis (Rosenhauer 1856)	0	0	**0**	0	0	**0**	0	0	**0**	1	1	**2**	2	25	**27**	**29**
Ptinidae																
Ptinus (*Cyphoderes*) *hirticornis* (Kiesenwetter 1867)	0	1	**1**	0	0	**0**	2	1	**3**	1	1	**2**	0	0	**0**	**6**
Ptinus sp (♀ no identificable)	0	0	**0**	0	0	**0**	0	0	**0**	0	1	**1**	0	0	**0**	**1**
Histeridae																
Saprinus acuminatus (F. 1798)	3	5	**8**	0	0	**0**	0	0	**0**	0	0	**0**	0	0	**0**	**8**
Saprinus caerulescens (Hoffmann 1803)	0	2	**2**	7	5	**12**	1	0	**1**	1	3	**4**	7	0	**7**	**26**
Saprinus detersus (Illiger 1807)	27	50	**77**	41	15	**56**	24	49	**73**	8	8	**16**	47	24	**71**	**293**
Saprinus figuratus Marseul 1855	0	0	**0**	0	0	**0**	0	0	**0**	1	1	**2**	0	0	**0**	**2**
S. furvus Erichson 1834	40	76	**116**	33	21	**54**	8	8	**16**	6	1	**7**	0	1	**1**	**194**
Saprinus georgicus Marseul 1862	11	10	**21**	12	5	**17**	11	13	**24**	5	0	**5**	3	4	**7**	**74**
Saprinus inmundus (Gyllenhal 1827)	0	2	**2**	0	0	**0**	0	0	**0**	0	0	**0**	4	0	**4**	**6**
Saprinus lautus Erichson 1839	0	0	**0**	2	0	**2**	2	2	**4**	1	0	**1**	1	0	**1**	**8**
Saprinus maculatus (Rossi 1792)	1	0	**1**	2	0	**2**	0	0	**0**	0	0	**0**	0	0	**0**	**3**
Saprinus melas Küster 1849	9	21	**30**	5	2	**7**	1	2	**3**	3	5	**8**	0	1	**1**	**49**
Saprinus politus (Brahm 1790)	10	4	**14**	11	0	**11**	5	3	**8**	1	2	**3**	15	3	**18**	**54**
Saprinus subnitescens Bickhardt 1909	25	18	**43**	8	18	**26**	19	17	**36**	10	9	**19**	31	19	**50**	**174**
Saprinus tenuistrius Marseul 1855	6	1	**7**	0	1	**1**	1	0	**1**	0	0	**0**	0	0	**0**	**9**
Margarinotus brunneus (F. 1775)	0	1	**1**	0	0	**0**	1	1	**2**	0	0	**0**	3	3	**6**	**9**
Margarinotus ignobilis (Marseul 1854)	0	0	**0**	0	0	**0**	0	0	**0**	0	0	**0**	1	1	**2**	**2**
Atholus duodecimstriatus (Schrank 1781)	0	0	**0**	0	0	**0**	0	0	**0**	0	0	**0**	3	1	**4**	**4**

(*continued*)

Table 2. Continued

Coleoptera species	HERBSH-1	HERBSH-2	**Total HERBSH**	JUNIP-1	JUNIP-2	**Total JUNIP**	HOLM-1	HOLM-2	**Total HOLM**	LUSIT-1	LUSIT-2	**Total LUSIT**	RIPFOR-1	RIPFOR-2	**Total RIPFOR**	**Total Captures**
Chalcionellus decemstriatus (Rossi 1792)	0	0	**0**	0	0	**0**	0	0	**0**	0	0	**0**	1	0	**1**	**1**
Onthophilus striatus (Forster 1771)	0	0	**0**	0	0	**0**	0	0	**0**	0	0	**0**	1	1	**2**	**2**
Staphylinidae																
Creophillus maxillosus (L. 1758)	16	4	**20**	7	4	**11**	6	9	**15**	7	4	**11**	64	20	**84**	**141**
Ontholestes tessellatus (Geoffroy 1785)	0	0	**0**	1	2	**3**	2	3	**5**	0	0	**0**	37	16	**53**	**61**
Platydracus meridionalis (Rosenhauer 1847)	1	6	**7**	0	7	**7**	0	0	**0**	15	19	**34**	2	5	**7**	**55**
Philonthus intermedius (Lacordaire 1835)	0	0	**0**	0	0	**0**	3	2	**5**	0	0	**0**	100	1	**101**	**106**
Philonthus politus (L. 1758)	0	0	**0**	0	0	**0**	0	1	**1**	0	0	**0**	1	0	**1**	**2**
Quedius latinus Gridelli 1938	0	0	**0**	0	0	**0**	0	0	**0**	0	0	**0**	3	7	**10**	**10**
Scarabaeidae																
Scarabaeus laticollis L. 1767	3	3	**6**	37	1	**38**	0	0	**0**	0	0	**0**	0	0	**0**	**44**
Onthophagus (Palaeonthophagus) lemur (F. 1781)	0	1	**1**	1	0	**1**	1	3	**4**	3	2	**5**	4	2	**6**	**17**
Onthophagus (Palaeonthophagus) coenobita (Herbst 1783)	0	0	**0**	0	0	**0**	0	0	**0**	0	0	**0**	5	5	**10**	**10**
Onthophagus (Palaeonthophagus) similis (Scriba 1790)	0	0	**0**	0	0	**0**	0	1	**1**	0	0	**0**	3	0	**3**	**4**
Onthophagus (Trichonthophagus) maki (Illiger 1803)	0	1	**1**	1	0	**1**	0	0	**0**	0	0	**0**	1	0	**1**	**3**
Onthophagus (Furconthophagus) furcatus (F. 1781)	0	1	**1**	0	0	**0**	0	0	**0**	0	0	**0**	0	0	**0**	**1**
Trogiidae																
Trox perlatus Goeze 1777	0	1	**1**	0	2	**2**	0	0	**0**	0	0	**0**	0	0	**0**	**3**
Trox cricetulus Ádám 1994	0	0	**0**	3	1	**4**	0	0	**0**	0	1	**1**	1	0	**1**	**6**
Carabidae																
Calathus granatensis Vuillefroy 1866	0	0	**0**	2	0	**2**	2	30	**32**	0	0	**0**	0	0	**0**	**34**
Calathus ambiguus (Paykull 1790)	0	0	**0**	0	0	**0**	0	1	**1**	0	0	**0**	0	0	**0**	**1**
Calathus fuscipes (Goeze 1777)	0	0	**0**	0	0	**0**	0	1	**1**	0	0	**0**	0	0	**0**	**1**
Calathus cinctus Motschulsky 1850	0	0	**0**	0	0	**0**	0	1	**1**	0	0	**0**	0	0	**0**	**1**
Ocys harpaloides (Audinet-Serville 1821)	0	0	**0**	0	0	**0**	0	0	**0**	0	0	**0**	0	1	**1**	**1**
Trechus obtusus Erichson 1837	0	0	**0**	0	0	**0**	0	0	**0**	0	0	**0**	1	0	**1**	**1**

Downloaded From: https://complete.bioone.org/journals/Journal-of-Insect-Science on 28 Mar 2024
Terms of Use: https://complete.bioone.org/terms-of-use

Downloaded From: https://complete.bioone.org/journals/Journal-of-Insect-Science on 28 Mar 2024
Terms of Use: https://complete.bioone.org/terms-of-use

Table 3. List of carrion Diptera species collected and abundances by sampling site and habitat

Diptera species	HERBSH-1	HERBSH-2	**Total HERBSH**	JUNIP-1	JUNIP-2	**Total JUNIP**	HOLM-1	HOLM-2	**Total HOLM**	LUSIT-1	LUSIT-2	**Total LUSIT**	RIPFOR-1	RIPFOR-2	**Total RIPFOR**	**Total captures**
Sarcophagidae																
Sarcophaga lehmanni (Müeller 1922)	10	5	**15**	12	16	**28**	8	10	**18**	22	9	**31**	51	13	**64**	**156**
Sarcophaga portschinskyi (Rohdendorf 1937)	0	4	**4**	6	7	**13**	3	4	**7**	17	25	**42**	0	0	**0**	**66**
Sarcophaga argyrostoma Robineau-Desvoidy 1830	1	2	**3**	1	9	**10**	3	4	**7**	8	0	**8**	0	0	**0**	**28**
Sarcophaga africa (Wiedemann 1824)	0	0	**0**	0	0	**0**	0	2	**2**	0	0	**0**	1	0	**1**	**3**
Sarcophaga belgiana Lehrer 1976	0	0	**0**	1	1	**2**	0	0	**0**	1	0	**1**	0	0	**0**	**3**
Sarcophaga variegata Scopoli 1763	0	0	**0**	0	0	**0**	0	0	**0**	0	0	**0**	3	0	**3**	**3**
Sarcophaga subvicina Rohdendorf 1937	0	0	**0**	0	0	**0**	0	0	**0**	0	0	**0**	1	0	**1**	**1**
Sarcophaga teretirostris Pandellé 1896	0	0	**0**	0	0	**0**	0	0	**0**	0	0	**0**	1	0	**1**	**1**
Sarcophaga cultellata (Pandelle 1896)	0	1	**1**	0	0	**0**	0	0	**0**	0	0	**0**	0	0	**0**	**1**
Ravinia pernix (Harris 1780)	1	0	**1**	3	3	**6**	0	1	**1**	0	0	**0**	0	0	**0**	**8**
Sarcophila latifrons (Fallén 1817)	5	5	**10**	36	20	**56**	0	10	**10**	27	0	**27**	10	0	**10**	**113**
Calliphoridae																
Calliphora vomitoria (L. 1758)	1	0	**1**	0	0	**0**	15	14	**29**	1	0	**1**	9	62	**71**	**102**
Calliphora vicina Robineau-Desvoidy 1830	6	28	**34**	16	28	**44**	37	53	**90**	96	27	**123**	35	101	**136**	**427**
C. albiceps (Wiedemann 1819)	15	107	**122**	266	187	**453**	77	82	**159**	185	19	**204**	555	214	**769**	**1,707**
Lucilia sericata (Meigen 1826)	25	16	**41**	11	14	**25**	17	8	**25**	20	10	**30**	19	25	**44**	**165**
Lucilia caesar (L. 1758)	0	0	**0**	0	0	**0**	14	12	**26**	1	1	**2**	15	21	**36**	**64**
Lucilia richardsi (Collin 1926)	0	0	**0**	0	1	**1**	0	0	**0**	1	0	**1**	0	0	**0**	**2**
Pollenia rudis (F. 1794)	4	32	**36**	6	25	**31**	52	50	**102**	55	5	**60**	445	63	**508**	**737**
Pollenia labialis Robineau-Desvoidy 1863	0	4	**4**	3	32	**35**	3	4	**7**	7	0	**7**	69	16	**85**	**138**
Pollenia fulvipalpis Macquart 1835	0	0	**0**	0	10	**10**	0	0	**0**	7	6	**13**	0	0	**0**	**23**
Pollenia bicolor Robineau-Desvoidy 1830	0	0	**0**	0	3	**3**	1	0	**1**	0	1	**1**	0	0	**0**	**5**
Pollenia ponti Rognes 1991	0	0	**0**	1	0	**1**	0	0	**0**	1	1	**2**	0	0	**0**	**3**
Stomorhina lunata (F. 1805)	0	0	**0**	0	0	**0**	0	0	**0**	1	0	**1**	0	0	**0**	**1**
Muscidae																
Muscina levida (Harris ×)	2	7	**9**	24	35	**59**	353	325	**678**	153	67	**220**	250	50	**300**	**1,266**
Muscina prolapsa (Harris 1780)	0	2	**2**	6	35	**41**	31	40	**71**	82	97	**179**	61	160	**221**	**514**
Muscina stabulans (Fallén 1817)	0	1	**1**	0	7	**7**	2	2	**4**	1	1	**2**	7	3	**10**	**24**
Hydrotaea aenescens (Wiedemann 1830)	0	0	**0**	0	0	**0**	0	0	**0**	1	0	**1**	1	0	**1**	**2**
H. armipes (Fallén 1825)	0	19	**19**	145	24	**169**	582	228	**810**	259	19	**278**	178	299	**477**	**1,753**
Hydrotaea capensis (Wiedemann 1818)	0	1	**1**	3	1	**4**	0	0	**0**	0	0	**0**	18	0	**18**	**23**
Hydrotaea cyrtoneurina (Zetterstedt 1845)	0	0	**0**	0	0	**0**	0	0	**0**	10	0	**10**	18	36	**54**	**64**
Hydrotaea dentipes (F. 1805)	0	0	**0**	1	0	**1**	9	8	**17**	0	0	**0**	29	129	**158**	**176**
H. ignava (Harris 1780)	7	9	**16**	40	8	**48**	194	209	**403**	59	7	**66**	380	481	**861**	**1,394**
M. domestica L. 1758	24	109	**133**	180	59	**239**	0	1	**1**	5	1	**6**	90	3	**93**	**472**
Helina reversio (Harris 1780)	0	1	**1**	3	19	**22**	0	0	**0**	6	1	**7**	0	0	**0**	**30**
Helina pubescens (Stein 1893)	0	0	**0**	0	1	**1**	0	0	**0**	0	0	**0**	0	0	**0**	**1**
Helina impuncta (Fallén 1825)	0	0	**0**	0	0	**0**	0	0	**0**	0	2	**2**	0	0	**0**	**2**
Helina depuncta (Fallén 1825)	0	1	**1**	0	0	**0**	0	0	**0**	1	0	**1**	0	4	**4**	**6**
Polietes lardarius (F. 1751)	0	0	**0**	1	0	**1**	0	0	**0**	0	0	**0**	0	0	**0**	**1**
Phaonia mediterranea Hennig 1963	0	0	**0**	0	0	**0**	1	0	**1**	1	0	**1**	0	0	**0**	**2**
Phaonia subventa (Harris 1780)	0	0	**0**	0	0	**0**	2	1	**3**	1	2	**3**	1	0	**1**	**7**
Phaonia pallida (F. 1787)	0	0	**0**	0	0	**0**	0	1	**1**	0	0	**0**	2	1	**3**	**4**
Eudasyphora cyanella (Meigen 1826)	0	0	**0**	0	0	**0**	0	0	**0**	0	0	**0**	0	1	**1**	**1**
Tachinidae																
Tachina magnicornis (Zetterstedt 1844)	5	2	**7**	14	4	**18**	1	2	**3**	5	0	**5**	8	3	**11**	**44**
Peleteria rubescens (Robineau-Desvoidy 1830)	0	13	**13**	6	23	**29**	0	0	**0**	1	0	**1**	3	0	**3**	**46**
Peleteria varia (F. 1794)	0	0	**0**	1	3	**4**	2	9	**11**	13	0	**13**	24	7	**31**	**59**
Heleomyzidae																
Suillia affinis (Meigen 1830)	0	17	**17**	10	40	**50**	85	36	**121**	81	78	**159**	2	0	**2**	**349**
Suillia tuberiperda (Rondani 1867)	0	0	**0**	0	0	**0**	0	0	**0**	0	0	**0**	2	1	**3**	**3**
Suillia flagripes (Czerny 1904)	0	1	**1**	5	4	**9**	43	2	**45**	11	18	**29**	12	5	**17**	**101**

Table 4. Number of observed species and number of species calculated from each estimator for each sampling habitat

Habitat	Observed species	ACE	ICE	Chao 1	Chao 2	Jackknife 1	Jackknife 2
HERBSH	60	82.4	82.38	87.2	82.14	80.13	89.11
		72.81%	72.83%	68.80%	73.04%	74.88%	67.33%
JUNIP	66	76.34	78.05	74.25	75.71	80.57	86.17
		86.45%	84.56%	88.88%	87.17%	81.92%	76.59%
HOLM	63	73.47	74.38	79.5	72.89	76.6	82.3
		85.75%	84.70%	79.24%	86.43%	82.24%	76.55%
LUSIT	66	79.41	77.84	74.27	78.25	84.38	91.13
		83.11%	78.74%	88.86%	84.34%	78.21%	72.42%
RIPFOR	80	96.29	100.64	102.67	96.1	101	110
		83.08%	79.50%	77.92%	83.25%	79.21%	72.72%
Total	112	125.3	127.06	125.6	121.72	131.44	133.82
		89.38%	87.77%	89.17%	92.01%	85.21%	83.69%

HERBSH, herbaceous and shrub vegetation; JUNIP, juniper forest; HOLM, holm oakwood; LUSIT, Lusitanian oakwood; RIPFOR, riparian forest.

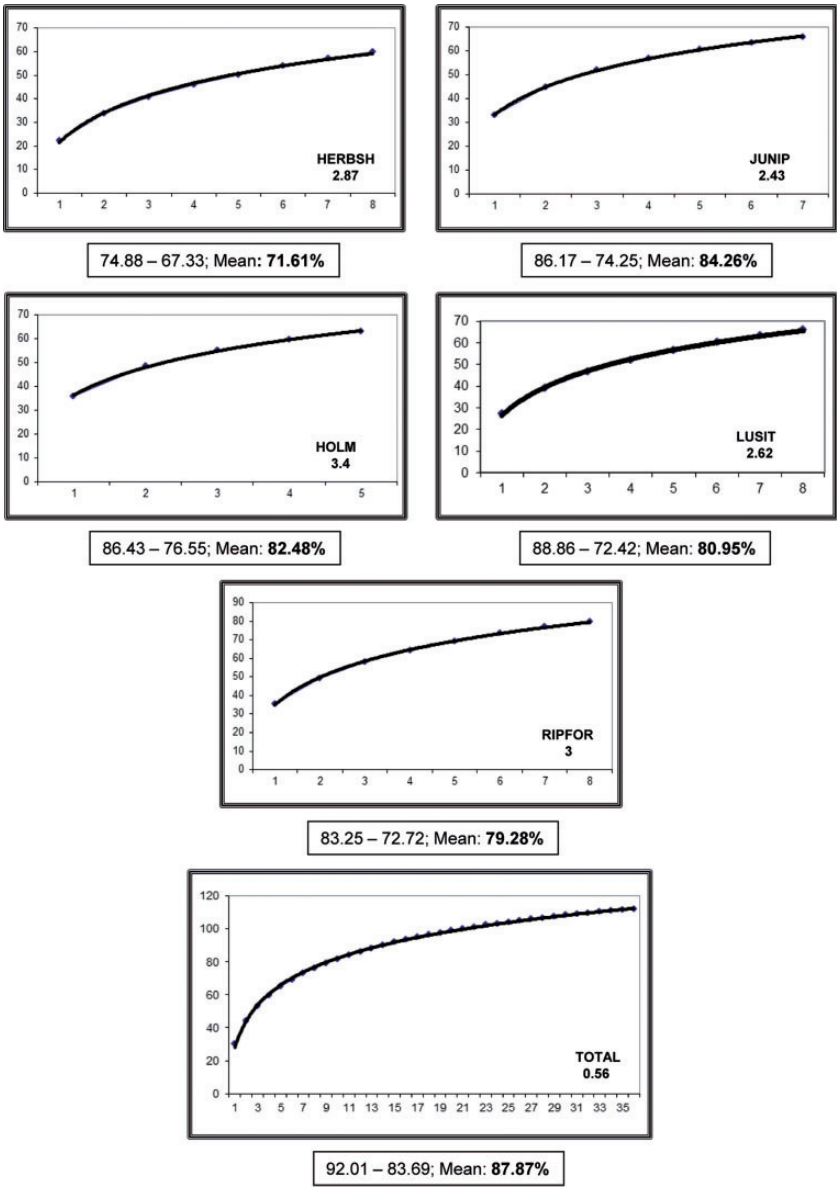


Fig. 2. Randomized accumulation curves for observed species richness in each sampling habitat and for the total captures. The rate of appearance of new inventoried species is included below the abbreviation for each habitat. Maximum, minimum, and mean values of inventory completeness according to the different estimators are also included. HERBSH, herbaceous and shrub vegetation; JUNIP, juniper forest; HOLM, holm oakwood; LUSIT, Lusitanian oakwood; RIPFOR, riparian forest.

Downloaded From: https://complete.bioone.org/journals/Journal-of-Insect-Science on 28 Mar 2024
Terms of Use: https://complete.bioone.org/terms-of-use

Results

In total, 25,033 individuals belonging to 112 carrion insect species from orders Coleoptera and Diptera were collected in carrion-baited traps throughout the sampling period. A complete list of these Coleoptera and Diptera species and numbers collected in each sampling site are presented in Tables 2 and 3, respectively. Among Coleoptera, the most abundant species were the silphids *Thanatophilus rugosus* (7,333 individuals) and *Thanatophilus ruficornis* (1,885 individuals), and the dermestid *Dermestes frischi* (2,252 individuals; Table 2). Among Diptera, the most abundant species were the muscids *Hydrotaea armipes* (1,753 individuals) and *Hydrotaea ignava* (1,394 individuals), and the calliphorid *Chrysomya albiceps* (1,707 individuals; Table 3). As mentioned earlier, opportunistic or casual insect species has not been included in this study, but details on their captures can be found in Baz et al. (2010). Unbaited traps virtually captured no insects, and in any case, only accidental species were collected in these traps.

The sampling habitat with the highest number of inventoried species was the riparian forest (80 species), while the rest of sampling habitats showed 60–66 inventoried species (Table 4). The species accumulation curves for each sampling habitat were not quite reaching the asymptote by the end of the sampling process; in the curve for the total of inventoried species in the study area, the asymptotic tendency was clearer (Fig. 2). All the nonparametric estimators showed inventory completeness >70% in every sampling habitat, with the exception of the herbaceous and shrub vegetation where the inventory completeness was 68.8 and 67.33% by Chao 1 and jackknife 2 estimators, respectively (Table 4). Inventory completeness was >80% for the total of inventoried species in the study area, according to all the nonparametric estimators (Table 4).

Both Shannon and Simpson diversity indices showed Lusitanian oakwood and riparian forest as the most diverse habitats; the rest of sampling habitats showed lower but similar diversity values (Table 5). The logarithmical distribution of species abundance (Figs. 3–5) in two of the less diverse habitats, herbaceous and shrub vegetation and juniper forest, showed that only six species were represented by the collection of more than 100 specimens. The silphid beetles *T. rugosus* and *T. ruficornis*, the dermestid beetle *D. frischi*, the calliphorid fly *C. albiceps,* and the muscid fly *Musca domestica* were the most abundant species in both habitats, together with the histerid beetle *Saprinus furvus* in herbaceous and shrub vegetation and the muscid fly *H. armipes* in juniper forest (Fig. 3). On the other hand, holm oakwood, Lusitanian oakwood, and riparian forest showed 11, 9, and 14 species represented by the collection of more than 100 specimens, respectively (Figs. 4 and 5). The silphid *T. rugosus* was also the most abundant

Table 5. Number of observed species and Shannon and Simpson diversity indices values for each sampling habitat

Habitat	Observed species	Shannon	Simpson
HERBSH	60	2.18	4.91
JUNIP	66	2.29	4.97
HOLM	63	2.35	5.23
LUSIT	66	3.03	14.05
RIPFOR	80	3.01	12.36

HERBSH, herbaceous and shrub vegetation; JUNIP, juniper forest; HOLM, holm oakwood; LUSIT, Lusitanian oakwood; RIPFOR, riparian forest.

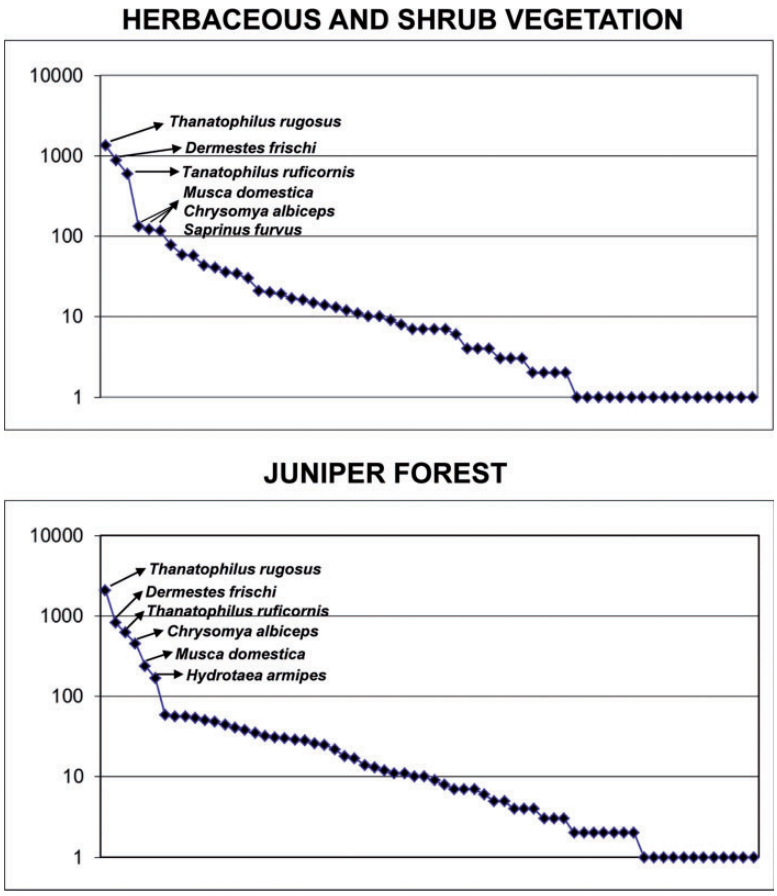


Fig. 3. Distribution of the abundances of the inventoried species in herbaceous and shrub vegetation and juniper forest habitats. Species names of those represented by the collection of >100 specimens are shown.

Downloaded From: https://complete.bioone.org/journals/Journal-of-Insect-Science on 28 Mar 2024
Terms of Use: https://complete.bioone.org/terms-of-use

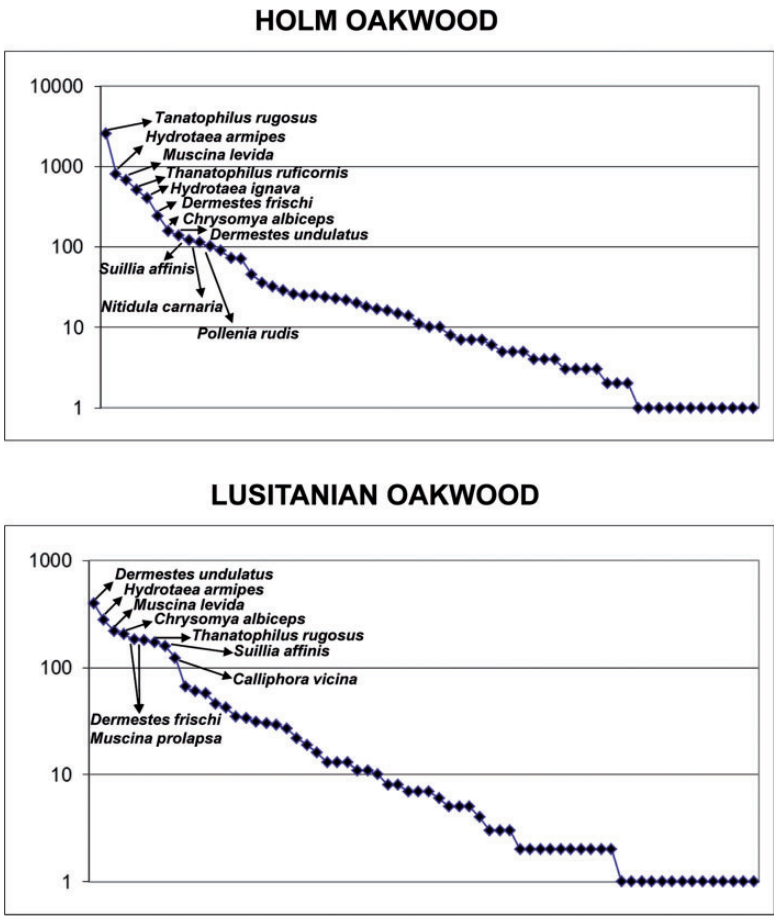


Fig. 4. Distribution of the abundances of the inventoried species in holm oakwood and Lusitanian oakwood habitats. Species names of those represented by the collection of >100 specimens are shown.

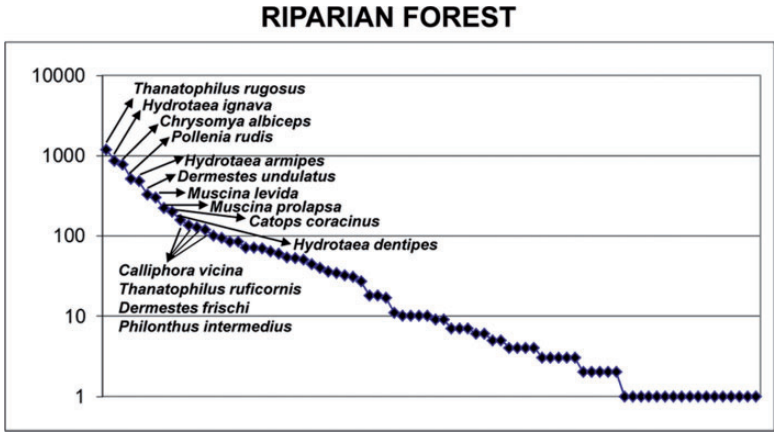


Fig. 5. Distribution of the abundances of the inventoried species in riparian forest habitat. Species names of those represented by the collection of >100 specimens are shown.

species in holm oakwood and riparian forest, whereas in Lusitanian oakwood, the most abundant species was the dermestid beetle *Dermestes undulatus* (Figs. 4 and 5). Among others, the muscid flies *H. ignava*, *H. armipes*, and *Muscina levida*, the blow fly *C. albiceps,* and the dermestid *D. frischi* were also abundant in these habitats (Figs. 4 and 5).

Finally, the cluster analysis showed two groups of habitats clearly separated (Fig. 6): one corresponding to more open habitats (herbaceous and shrub vegetation and juniper forest) and the other corresponding to more wooded areas (holm oakwood, Lusitanian oakwood, and riparian forest). Interestingly, the two habitats included in the first group were also the two least diverse habitats (Table 5).

Discussion

Faunistic inventories are generally incomplete due to the fact that the number of inventoried species continues increasing with sampling effort (Gotelli and Colwell 2001, Jiménez-Valverde and Hortal 2003). In accordance with it, in this study, none of the randomized

Downloaded From: https://complete.bioone.org/journals/Journal-of-Insect-Science on 28 Mar 2024
Terms of Use: https://complete.bioone.org/terms-of-use

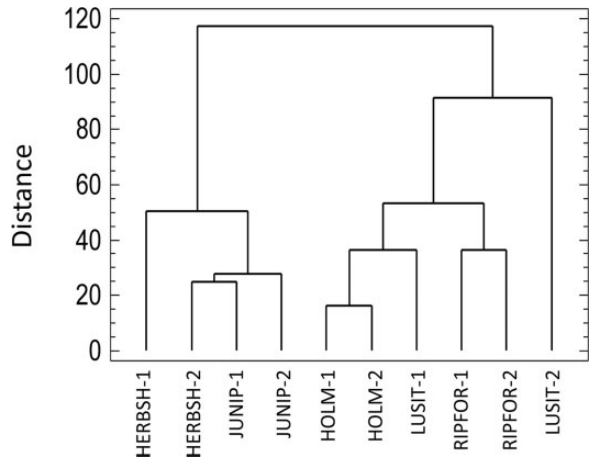


Fig. 6. Dendrogram from cluster analysis for the sampling sites according to their species composition and the average number of collected individuals in each survey. The numbers 1 and 2 indicate the two sites selected for each habitat. HERBSH, herbaceous and shrub vegetation; JUNIP, juniper forest; HOLM, holm oakwood; LUSIT, Lusitanian oakwood; RIPFOR, riparian forest.

accumulation curves reached the asymptote by the end of the sampling process (Fig. 2). However, estimations of species richness become stable when inventories cover at least 70% of the total number of species (Jiménez-Valverde and Hortal 2003). Thus, nonparametric estimators showed that the inventory completeness of this study was representative enough of the species richness in the study area (Table 4).

Riparian forest was the sampling habitat with the greatest species richness (Table 5), whereas both Shannon and Simpson indices showed not only riparian forest but also Lusitanian oakwood as the most diverse habitats (Table 5). Diversity is actually a set of concepts (Peet 1974), and it is not a synonym of number of species. For example, both juniper forest and Lusitanian oakwood showed the same number of inventoried species, but juniper forest was clearly a less diverse habitat in accordance with Shannon and Simpson indices values (Table 5). The explanation lies in the combination of the number of species and their relative abundance. In juniper forest, fewer species dominate in terms of number of individuals than in Lusitanian oakwood, where the number of collected specimens appears to be more equally distributed among the inventoried species (Figs. 3 and 4). Among the most abundant species, the silphids *T. rugosus* and *T. ruficornis* and the dermestids *D. undulatus* and *D. frischi* clearly stood out (Figs. 3–5). Those species are well represented in every type of natural habitats in central Spain, dominating the necrophagous beetles assemblages (Martín-Vega and Baz 2012). On the other hand, the blow fly *C. albiceps* and Muscidae species from genera *Muscina* and *Hydrotaea* have also been found among the dominant species in the sarcosaprophagous Diptera assemblages in summer months in the different natural habitats of central Spain (Martínez-Sánchez et al. 2000, Martín-Vega and Baz 2013), in accordance with the present results (Figs. 3–5). Carrion sustains a high diverse insect community exploiting the same resource at the same time, although calliphorid flies and silphid and dermestid beetles usually monopolize the different carcass tissues along the insect succession (Braack 1987, Matuszewski et al. 2010).

The cluster analysis divided the sampled habitats into two well-differentiated groups (Fig. 6), which corresponded to the least diverse habitats (herbaceous and shrub vegetation and juniper forest), and, on the other hand, the most diverse (holm oakwood, Lusitanian oakwood, and riparian forest; Table 5). The fact that these two groups also coincide with the more open (i.e., less wooded) and the more forest habitats allow speculating on the reasons for the different diversity values. Thus, in deciduous and sclerophyllous forests, the ground usually accumulates great quantities of fallen leaves, which may favor the proliferation of saprophagous species. Furthermore, forest habitats may also contain a higher diversity of small vertebrates than more open, homogeneous habitats (Tews et al. 2004), thus providing carrion insects with higher numbers of animal carcasses. It may explain the high abundance of *Muscina* and *Suillia* species in wooded habitats (Figs. 4 and 5; Table 3). Despite muscid flies of genus *Muscina* breed frequently on carrion, they are also common on fungus and other types of decaying organic matter (Gregor et al. 2002). Also, heleomyzid flies of genus *Suillia* can be collected abundantly in association with carrion (e.g., Martín-Vega and Baz 2013), but they are mainly considered as saprophagous (Séguy 1934). Both *Muscina* and *Suillia* species are among the dominant species, which typify the sarcosaprophagous fly assemblages of wooded habitats in central Spain (Martín-Vega and Baz 2013). Moreover, the accumulated fallen leaves in wooded habitats may favor the presence of invertebrates, which can be parasitized by some fly species that are also attracted to carrion. This is the case of *Pollenia* species (Figs. 4 and 5; Table 3), which are primarily considered as parasites or predators of earthworms, but they have also been collected in association with carrion (Baz et al. 2007, Martín-Vega and Baz 2013). Also, *Muscina* species can act as parasites of larvae of other insects (Gregor et al. 2002). On the other hand, the traps installed in more open habitats appeared to be subjected to higher temperatures (Table 1) as they were more exposed to continuous sunlight during the day. Such situation may have favored the dominance of the silphids *T. rugosus* and *T. ruficornis*, and the dermestid *D. frischi* (Fig. 3), which are tolerant to different environmental conditions (Martín-Vega and Baz 2012). Particularly, *D. frischi* appears to be more dominant in open habitats (Fig. 3); it may be due to its food preferences. As dermestid species breed on carcasses in advanced stages of decomposition, they are usually more abundant in those habitats with low humidity and high temperatures, where carrion desiccates quickly (Sánchez Piñero 1997, Martín-Vega and Baz 2012). In this sense, open habitats may have also favored the dominance of thermophilous species like *C. albiceps* and *M. domestica*, which show a preference for warm conditions in its spatial distribution (Baz et al. 2007, Martín-Vega and Baz 2013); moreover, *C. albiceps* have shown to be more abundant on carrion under sunny conditions than shaded conditions (Prado e Castro et al. 2011).

In conclusion, the different characteristics of the studied habitats and their climatic conditions may be determinant explaining the composition and diversity of their typical carrion insect assemblages. From an applied point of view, differences in habitat preferences and species composition of carrion insects may provide forensic investigators with useful information indicating possible postmortem movements of a corpse (Amendt et al. 2011). Further studies on the diversity and composition of carrion insect species in different habitats and environments within a same region need to be done.

Acknowledgments

We are grateful to Txomin Gilgado, Jacinto Gamo, Jaime Potti, Jorge Vicente, and Gonzalo Baz for their company and their help during the sampling process. We are also grateful to the staff of the "House of the Park Hoces del Río Riaza" for their kindness and good disposal, and to the ground keepers of the Natural Park and especially to Luis Mira for sharing with us their knowledge about the study area. Finally, we acknowledge Francisco Sánchez-Aguado (Paco), Director-Conservator of the Natural Park (and nevertheless, friend), for his infinite patience, besides other regards. Two anonymous reviewers provided valuable comments and suggestions, which improved this manuscript. This work was supported by the Consejería de Medio Ambiente de la Junta de Castilla y León (Project SG-25/2007) within the framework of the development of catalogues programmed by the Natural Park "Hoces del Río Riaza," intended for improving the knowledge about its natural resources. The authors are members of

Downloaded From: https://complete.bioone.org/journals/Journal-of-Insect-Science on 28 Mar 2024
Terms of Use: https://complete.bioone.org/terms-of-use

the University Institute of Research in Police Sciences (IUICP) of the University of Alcalá.

References Cited

Amendt, J., C. S. Richards, C. P. Campobasso, R. Zehner, and M.J.R. Hall. 2011. Forensic entomology: applications and limitations. Forensic Sci. Med. Pathol. 7: 379–392.

Anderson, G., and S. L. VanLaerhoven. 1996. Initial studies on insect succession on carrion in southwestern British Columbia. J. Forensic Sci. 41: 617–625.

Barton, P. S., S. A. Cunningham, D. B. Lindenmayer, and A. D. Manning. 2013. The role of carrion in maintaining biodiversity and ecological processes in terrestrial ecosystems. Oecologia 171: 761–772.

Baz, A., B. Cifrián, D. Martín-Vega, and M. Baena. 2010. Phytophagous insects captured in carrion-baited traps. Bull. Insectol. 63: 21–30.

Baz, A., B. Cifrián, L. M. Díaz-Aranda, and D. Martín-Vega. 2007. The distribution of adult blow-flies (Diptera: Calliphoridae) along an altitudinal gradient in central Spain. Ann. Soc. Entomol. France 43: 289–296.

Braack, L.E.O. 1987. Community dynamics of carrion-attendant arthropods in tropical African woodland. Oecologia 72: 402–409.

Cardoso, P., S. S. Henriques, C. Gaspar, L. C. Crespo, R. Carvalho, J. B. Schmidt, P. Sousa, and T. Szűts. 2009. Species richness and composition assessment of spiders in a Mediterranean scrubland. J. Insect Conserv. 13: 45–55.

Colwell, R. K. 2005. Estimates: statistical estimation of species richness and shared species from samples, version 7.5. (http://viceroy.eeb.ucom.edu/estimates) (accessed 10 December 2012).

Colwell, R. K., and J. A. Coddington. 1994. Estimating terrestrial biodiversity through extrapolation. Philos. Trans. R. Soc. Lond. B 345: 101–118.

Goff, M. L. 1991. Comparison of insect species associated with decomposing remains recovered inside dwellings and outdoors on the island of Oahu, Hawaii. J. Forensic Sci. 36: 748–753.

Gotelli, N. J., and R. K. Colwell. 2001. Quantifying biodiversity: procedures and pitfalls in the measurement and comparison of species richness. Ecol. Lett. 4: 379–391.

Gregor, F., R. Rozkošný, M. Barták, and J. Vaňhara. 2002. The Muscidae (Diptera) of central Europe. Masaryk University, Brno, Czech Republic.

Jiménez-Valverde, A., and J. Hortal. 2003. Las curvas de acumulación de especies y la necesidad de evaluar la calidad de los inventarios biológicos. Rev. Ibérica Aracnol. 8: 151–161.

Leclercq, M. 1978. Entomologie et médecine légale: datation de la mort. Masson, Paris, France.

Magurran, A. E. 2004. Measuring biological diversity. Wiley-Blackwell, London, United Kingdom.

Martín-Vega, D., and A. Baz. 2012. Spatiotemporal distribution of necrophagous beetles (Coleoptera: Dermestidae, Silphidae) assemblages in natural habitats of central Spain. Ann. Entomol. Soc. Am. 105: 44–53.

Martín-Vega, D., and A. Baz. 2013. Sarcosaprophagous Diptera assemblages in natural habitats in central Spain: spatial and seasonal changes in composition. Med. Vet. Entomol. 27: 64–76.

Martínez-Sánchez, A., S. Rojo, and M. A. Marcos-García. 2000. Annual and spatial activity of dung flies and carrion in a Mediterranean holmoak pasture ecosystem. Med. Vet. Entomol. 14: 56–63.

Matuszewski, S., D. Bajerlein, S. Konwerski, and K. Szpila. 2010. Insect succession and carrion decomposition in selected forests of central Europe. Part 1: pattern and rate of decomposition. Forensic Sci. Int. 194: 85–93.

Morón, M. A., and R. A. Terrón. 1984. Distribución altitudinal y estacional de los insectos necrófilos en la Sierra Norte de Hidalgo, México. Acta Zool. Mexicana 3: 1–47.

Newton, A., and S. B. Peck. 1975. Baited pitfall traps for beetles. Coleopt. Bull. 29: 45–46.

Ninyerola, M., X. Pons, and J. M. Roure. 2005. Atlas climático digital de la Península Ibérica. Metodología y aplicaciones en bioclimatología y geobotánica. Universidad Autónoma de Barcelona, Bellaterra, Spain.

Peet, R. K. 1974. The measurement of species diversity. Annu. Rev. Ecol. Syst. 5: 285–307.

Prado e Castro, C., J. P. Sousa, M. I. Arnaldos, J. Gaspar, and M. D. García. 2011. Blowflies (Diptera: Calliphoridae) activity in sun exposed and shaded carrion in Portugal. Ann. Soc. Entomol. France 47: 128–139.

Sánchez Piñero, F. 1997. Analysis of spatial and seasonal variability of carrion beetle (Coleoptera) assemblages in two arid zones of Spain. Environ. Entomol. 26: 805–814.

Séguy, E. 1934. Diptères (Brachycères) (Muscidae Acalypterae et Scatophagidae). Faune France 28: 1–832.

Tews, J., U. Brose, V. Grimm, K. Tielbörger, M. C. Wichmann, M. Schwager, and F. Jeltsch. 2004. Animal species diversity driven by habitat heterogeneity/diversity: the importance of keystone structures. J. Biogeogr. 31: 79–92.

Woodcock, B. A., A. D. Watt, and S. R. Leather. 2002. Aggregation, habitat quality and coexistence: a case study on carrion fly communities in slug cadavers. J. Anim. Ecol. 71: 131–140.

Received 9 January 2013; accepted 4 March 2013.

Downloaded From: https://complete.bioone.org/journals/Journal-of-Insect-Science on 28 Mar 2024
Terms of Use: https://complete.bioone.org/terms-of-use