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Source: Florida Entomologist, 100(1) : 134-144

Published By: Florida Entomological Society

URL: <https://doi.org/10.1653/024.100.0119>

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Diversity and abundance of edaphic arthropods associated with conventional and organic sugarcane crops in Brazil

Luan Alberto Odorizzi dos Santos, Natalia Naranjo-Guevara, and Odair Aparecido Fernandes*

Abstract

Although studies have shown enhancement of insects, birds, and plants in organically managed agroecosystems, information on arthropod diversity and abundance in conventional and organic sugarcane farms is scarce. This research was conducted to analyze and compare the diversity and abundance of edaphic arthropods in organic and conventional sugarcane by using pitfall traps. The study was conducted during 2 growing seasons in Jaboticabal, São Paulo, Brazil. In total, 13,244 individuals belonging to 190 morphospecies were collected. In the conventional system, 4,964 specimens were collected, representing 122 morphospecies distributed in 15 orders and 50 families. In the organic system, 8,280 individuals were captured, representing 142 morphospecies in 13 orders and 45 families. Ants of the genera *Pheidole* Westwood, *Dorymyrmex* Mayr, *Camponotus* Mayr, and *Crematogaster* Lund (Hymenoptera: Formicidae) were predominant. Higher abundance and richness of arthropods (especially predators and omnivores) were found in the organic than the conventional system, which could be important in regulating key pests of sugarcane. Our results show that the organic management in sugarcane increased the abundance and diversity of arthropods.

Key Words: community; conservation biological control; environmental disturbance; functional group

Resumo

Embora estudos já tenham mostrado que há incremento de insetos, pássaros e plantas em agroecossistemas manejados organicamente, informação sobre diversidade e abundância de artrópodes em plantios orgânicos e convencionais de cana-de-açúcar é rara. Este trabalho foi realizado para analisar e comparar a diversidade e abundância de artrópodes edáficos em cana-de-açúcar convencional e orgânica utilizando armadilhas pitfall. O estudo foi conduzido durante duas safras de cana-de-açúcar em Jaboticabal, São Paulo, Brasil. Foram coletados 13244 indivíduos pertencentes a 190 morfo-espécies. No sistema convencional 4964 espécimens foram coletados e representaram 122 morfo-espécies, distribuídos em 15 ordens e 50 famílias. No sistema orgânico, 8280 indivíduos foram capturados, correspondendo a 142 morfoespécies, distribuídos em 13 ordens e 45 famílias. Formigas dos gêneros *Pheidole* Westwood, *Dorymyrmex* Mayr, *Camponotus* Mayr e *Crematogaster* Lund (Hymenoptera: Formicidae) foram predominantes. Maior abundância e riqueza de artrópodes (especialmente predadores e onívoros) foram encontradas no sistema orgânico em comparação ao sistema convencional e poderiam ser importantes para a regulação de pragas chaves da cana-de-açúcar. Os resultados mostram que o manejo orgânico em cana-de-açúcar aumentou a abundância e diversidade de artrópodes.

Palavras Chave: comunidade; controle biológico conservativo; distúrbio ambiental; grupo funcional

Conventional agriculture has often caused the simplification of agricultural landscapes, mainly due to the establishment of monocultures (Pogue & Schnell 2001). These simplified agricultural practices and overuse of insecticides can lead to a reduction in biodiversity (Butler et al. 2007), and thus the reduction of ecological services. On the other hand, with the increase of organic farming, conservative biological control is also expected to increase due to the reduction in pesticide use and land management, which in turn enhance survival, fecundity, efficiency, longevity, and maintenance of natural enemies of arthropod pests (Eilenberg et al. 2001; Landis et al. 2005).

Current agricultural system management can be characterized by frequent and intense disturbances, which are unfavorable for conservation of natural enemies (Letourneau 1998). Thus, development and maintenance of an ecological infrastructure to provide food resources,

shelter, and alternative prey and hosts are the basis of environmental management. Consequently, it is possible to expand natural biological control by preserving and increasing existing populations of beneficial arthropods in crops (Gurr et al. 2000; Landis et al. 2000; Wilkinson & Landis 2005).

Environmental problems associated with conventional sugarcane agriculture due to the use of fire prior to harvest (forbidden in certain Brazilian regions since 2014), and use of pesticides, are well documented (Nunes et al. 2006). However, few studies have characterized the soil-dwelling arthropods that could be affected by these disturbances in sugarcane agroecosystems (Castelo Branco et al. 2010; Pasqualin et al. 2012; Abreu et al. 2014). Consequently, the objective of this work was to analyze and compare the diversity and abundance of edaphic arthropods in conventional and organic sugarcane fields.

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Materials and Methods

CHARACTERIZATION OF THE AREA

The experiment was conducted in 2 sugarcane areas in Jaboticabal municipality, São Paulo, Brazil. The sugarcane variety RB5536 was used in each of 2 seasons. Each area was about 10 ha. In the conventional field (21.1978°S, 48.2897°W, altitude 589 m), agricultural practices included pre-harvest burning and herbicide use for weed control but no insecticide application. In the organic field (21.1858°S, 48.2450°W; altitude 623m), sugarcane has been harvested without burning for about 10 yr (green cane) and grown without use of any pesticide. In the 1st growing season (2011/2012, 7th ratoon), the experiment started when plants were in the 4th month of development, whereas in the 2nd growing season (2012/2013, 8th ratoon), the experiment started just after harvest.

COLLECTION METHOD

In each plot, 3 parallel transects distanced 10 m apart from each other were established. Five pitfall traps (700 mL plastic cups buried and adjusted to ground level) were installed every 10 m on each transect, with the 1st trap installed 20 m from the edge of the plot. The traps received 100 mL of solution (98 mL water + 2 mL detergent) to prevent captured arthropods from escaping.

Sample collections began 24 h after installation of traps to reduce the effect of disturbance caused by soil excavation and trap installation (Araújo et al. 2005). The traps remained for 48 h in the experimental areas. Collected arthropods were taken to the laboratory for sorting and identification. Fifteen monthly collections were conducted during the 2011/2012 (Dec 2011 to May 2012) and 2012/2013 (Oct 2012 to Mar 2013) growing seasons.

IDENTIFICATION OF COLLECTED ARTHROPODS

The arachnids were sent to Instituto Butantan (São Paulo, Brazil) for identification. The other arthropods collected were identified to lowest possible taxonomic level by using specialized literature (Loureiro & Queiroz 1990; Borror et al. 1992; Baccaro 2006; Suguituru et al. 2015). Unidentified species were differentiated into morphospecies (Oliver & Beattie 1996).

DATA ANALYSES

Data were analyzed using the software ANAFAU (Moraes & Haddad 2003), and the faunistic indices dominance, abundance, frequency, and constancy were obtained for each system. Moreover, the program performs residual analysis of discrepant data that can be classified into exclusive categories known as super-dominant, super-abundant, and super-frequent. The dominance was calculated by the equation $DL = (1/S) \times 100$, where DL = dominance limit and S = total number of species per sample. The abundance was calculated using the standard deviation and confidence interval of the arithmetic mean at 1 and 5% probabilities. For frequency, the equation $F = (n/N) \times 100$ was adopted, where F = frequency (%), n = number of specimens of each species collected, and N = total number of specimens of the collected species. For constancy, the equation $C = (P/N) \times 100$ was used, where C = constancy, P = number of samples containing the species, N = total number of samples collected (Silveira Neto et al. 1976).

Diversity was assessed using the species diversity indices of Shannon–Wiener (H') and Margalef (α). The equitability index (E) was calculated to evaluate the uniformity of the captures and how the individuals are distributed in the sample, using the ANAFAU software (Moraes & Haddad 2003). The similarity index assessed the number of species shared between the 2 cropping systems.

To estimate the total species richness for each system, the software EstimateS[®] 9.1 was used to generate species accumulation curves and to compare the conventional and organic system (Colwell 2006). Samples were randomized 100 times, without replacement, using the non-parametric estimator first order Jackknife (Jack 1), which uses the number of unique species or species occurring only once in a sample to produce richness estimates (Heltsh & Forrester 1983). Also, principal component analysis (STATISTICA, StatSoft, Inc., Tulsa, Oklahoma) was performed separately on the collection from each month. Thus, 15 samples (= months of evaluation) were used in the analysis for each area.

Results

ARTHROPOD RICHNESS AND ECOLOGICAL ANALYSIS

In total, 13,244 individuals belonging to 190 morphospecies were collected in sugarcane. The number of individuals collected during the 2 growing seasons in the conventional system was 4,964 (37.48%), and was represented by 122 morphospecies distributed in 15 orders and 50 families. In the organic system, 8,280 individuals (62.52%) were captured, corresponding to 142 morphospecies in 13 orders and 45 families (Table 1). The number of individuals collected in the organic crop was 66.8% higher, although the numbers of taxonomic orders and families observed were slightly lower than in the conventional crop.

The Shannon–Wiener index (H'), used to estimate the diversity of arthropods considering the uniformity of abundance of species, was 2.3 for the conventional system and 2.5 for the organic system (Table 1). The Margalef index (α), calculated by the number of species and the logarithm of the total number of individuals, was 14.1 for the conventional system and 17.1 for the organic system. The equitability or uniformity parameter, which varies from 0 to 1 (the closer to 1 the greater the equality of species abundance), was approximately 0.49 for both production systems, suggesting that the arthropod community sampled tends to coexist in both systems with some dominance of certain species. The similarity between the areas was 0.687.

FAUNISTIC ANALYSIS

Dominance

In total, 2 and 17 morphospecies in the conventional system and 4 and 30 morphospecies in the organic system were observed to be super-dominant and dominant, respectively (Table 2). The number of non-dominant species was about 7% lower for the conventional system (103 morphospecies) in comparison with the organic system (108 morphospecies) (Table 2).

Among the 43 super-dominant and dominant morphospecies, which represented 23.5% of the total number of morphospecies, 10 (23.3% of the super-dominant and dominant) were collected in both systems. The numbers of individuals of the dominant and non-dominant morphospecies were 1,577 and 367 (conventional system) and 1,825 and 345 (organic system), respectively. However, the number of individuals of the super-dominant morphospecies in the organic system (6,110) was twice that observed in the conventional system (3,020) (Table 2).

Ants (Hymenoptera: Formicidae) of the genera *Pheidole* Westwood and *Dorymyrmex* Mayr collected in the conventional system were super-dominant morphospecies whereas *Camponotus* Mayr and *Crematogaster* Lund were considered dominant. On the other hand, these 4 genera were super-dominant in the organic system. Super-dominant species are native species that behave as invaders in a disturbed environment (Silva-Mattos & Pivello 2009). Other ant genera

Table 1. Number of individuals captured, faunistic indices (dominance, abundance, frequency, constancy) and functional groups of arthropods captured using pitfall traps in conventional and organic sugarcane systems in Jaboticabal, São Paulo, Brazil, during the 2011/2012 and 2012/2013 growing seasons.

Taxon	Morphospecies	Conventional						Organic						Functional Group
		No. Individuals Captured	No. collections	Dominance	Abundance	Frequency	Constancy	No. Individuals Captured	No. collections	Dominance	Abundance	Frequency	Constancy	
Blattodea	Blattellidae sp.1	11	6	ND	C	F	Y	3	3	ND	R	LF	Z	Omnivore
	Blattellidae sp.2	.	.	.	.	.	.	44	11	D	VA	VF	W	Omnivore
Coleoptera	Carabidae	.	.	.	.	.	.	5	2	ND	R	LF	Z	Predator
	<i>Loxandrus</i> sp.1	.	.	.	.	.	.	4	2	ND	R	LF	Z	Predator
	<i>Loxandrus</i> sp.2	2	2	ND	R	LF	Z	.	.	.	.	.	.	Predator
	<i>Loxandrus</i> sp.3	.	.	.	.	.	.	2	2	ND	R	LF	Z	Predator
	<i>Pseudabarys</i> sp.1	.	.	.	.	.	.	1	1	ND	R	LF	Z	Predator
	<i>Pseudabarys</i> sp.2	1	1	ND	R	LF	Z	.	.	.	.	.	.	Predator
	<i>Pseudabarys</i> sp.3	.	.	.	.	.	.	1	1	ND	R	LF	Z	Predator
	<i>Carabidae</i> sp.1	.	.	.	.	.	.	1	1	ND	R	LF	Z	Predator
	<i>Carabidae</i> sp.2	.	.	.	.	.	.	1	1	ND	R	LF	Z	Predator
	<i>Carabidae</i> sp.3	.	.	.	.	.	.	1	1	ND	R	LF	Z	Predator
	<i>Carabidae</i> sp.4	.	.	.	.	.	.	1	1	ND	R	LF	Z	Predator
	<i>Carabidae</i> sp.5	2	2	ND	R	LF	Z	.	.	.	.	.	.	Predator
	<i>Megacephala</i> sp.1	2	2	ND	R	LF	Z	1	1	ND	R	LF	Z	Predator
	<i>Cincidelinae</i> sp.1	4	1	ND	R	LF	Z	20	7	D	C	F	Y	Predator
Cincidelinae	<i>Cincidelinae</i> sp.2	4	3	ND	R	LF	Z	.	.	.	.	.	.	Predator
Chrysomelidae	Bruchinae sp.1	111	2	D	VA	VF	Z	.	.	.	.	.	.	Detritivore
	<i>Metamasius hemipterus</i>	1	1	ND	R	LF	Z	4	2	ND	R	LF	Z	Herbivore
Curculionidae	<i>Curculionidae</i> sp.1	1	1	ND	R	LF	Z	.	.	.	.	.	.	Herbivore
	<i>Conoderus scalaris</i>	4	3	ND	R	LF	Z	.	.	.	.	.	.	Herbivore
Elateridae	<i>Conoderus</i> sp.1	.	.	.	.	.	.	2	2	ND	R	LF	Z	Herbivore
	<i>Conoderus</i> sp.2	.	.	.	.	.	.	1	1	ND	R	LF	Z	Herbivore
Passalidae	<i>Passalidae</i> sp.1	1	1	ND	R	LF	Z	.	.	.	.	.	.	Herbivore
	<i>Rhizophagidae</i> sp.1	261	3	D	VA	VF	Z	.	.	.	.	.	.	Herbivore
Scarabaeidae	<i>Ataenius</i> sp.1	1	1	ND	R	LF	Z	.	.	.	.	.	.	Detritivore
	<i>Ataenius</i> sp.2	1	1	ND	R	LF	Z	.	.	.	.	.	.	Detritivore
	<i>Canthon</i> sp.1	11	6	ND	C	F	Y	.	.	.	.	.	.	Detritivore
	<i>Canthon</i> sp.2	.	.	.	.	.	.	3	1	ND	R	LF	Z	Detritivore
	<i>Canthon</i> sp.3	.	.	.	.	.	.	1	1	ND	R	LF	Z	Detritivore
	<i>Canthon</i> sp.4	.	.	.	.	.	.	1	1	ND	R	LF	Z	Detritivore
	<i>Canthon</i> sp.5	1	1	ND	R	LF	Z	.	.	.	.	.	.	Detritivore
	<i>Cyclocephala</i> sp.1	.	.	.	.	.	.	22	7	D	A	VF	Y	Herbivore
	<i>Cyclocephala</i> sp.2	.	.	.	.	.	.	2	2	ND	R	LF	Z	Herbivore
	<i>Cyclocephala</i> sp.3	10	5	ND	C	F	Y	.	.	.	.	.	.	Herbivore

SD = super-dominant; D = dominant; ND = non-dominant; SA = super-abundant; VA = very abundant; A = abundant; C = common; D = dispersed; R = rare; SF = super-frequent; VF = very frequent; F = frequent; LS = less frequent; W = constant; Y = accessory; Z = accidental.

Table 1. (Continued) Number of individuals captured, faunistic indices (dominance, abundance, frequency, constancy) and functional groups of arthropods captured using pitfall traps in conventional and organic sugarcane systems in Jaboticabal, São Paulo, Brazil, during the 2011/2012 and 2012/2013 growing seasons.

Taxon	Morphospecies	Conventional						Organic						Functional Group
		No. Individuals Captured	No. collections	Dominance	Abundance	Frequency	Constancy	No. Individuals Captured	No. collections	Dominance	Abundance	Frequency	Constancy	
Staphylinidae	Staphylinidae sp.1	3	2	ND	R	LF	Z	1	1	ND	R	LF	Z	Predator
	Staphylinidae sp.2	165	2	D	VA	VF	Z	1	1	ND	R	LF	Z	Predator
	Staphylinidae sp.3	.	.	.	.	.	.	1	1	ND	R	LF	Z	Predator
	Staphylinidae sp.4	16	6	ND	C	F	Y	81	6	D	VA	VF	Y	Predator
	Staphylinidae sp.5	1	1	ND	R	LF	Z	1	1	ND	R	LF	Z	Predator
	Staphylinidae sp.6	.	.	.	.	.	.	3	2	ND	R	LF	Z	Predator
	Staphylinidae sp.7	5	3	ND	R	LF	Z	.	.	.	.	.	.	Predator
Dermaptera Forficulidae	<i>Doru</i> sp.	1	1	ND	R	LF	Z	.	.	.	.	.	.	Predator
	Forficulidae sp.1	15	5	ND	C	F	Y	115	3	D	VA	VF	Z	Predator
	Forficulidae sp.2	.	.	.	.	.	.	1	1	ND	R	LF	Z	Predator
	<i>Labidura</i> sp.	15	5	ND	C	F	Y	1	1	ND	R	LF	Z	Predator
Diptera Agromyzidae Asilidae Culicidae Dolichopodidae Drosophilidae	Agromyzidae sp.1	.	.	.	.	.	.	24	3	D	A	VF	Z	Herbivore
	Asilidae sp.	5	2	ND	R	LF	Z	.	.	.	.	.	.	Predator
	Culicidae sp.1	.	.	.	.	.	.	2	2	ND	R	LF	Z	Omnivore
	Culicidae sp.2	7	1	ND	D	LF	Z	.	.	.	.	.	.	Omnivore
	<i>Condylostylus</i> sp.	2	1	ND	R	LF	Z	.	.	.	.	.	.	Predator
	Dolichopodidae sp.1	2	1	ND	R	LF	Z	7	4	ND	D	LF	Y	Predator
	Drosophilidae sp.1	1	1	ND	R	LF	Z	1	1	ND	R	LF	Z	Detritivore
	Drosophilidae sp.2	.	.	.	.	.	.	21	6	D	C	F	Y	Omnivore
	Drosophilidae sp.3	8	2	ND	D	LF	Z	1	1	ND	R	LF	Z	Omnivore
	Drosophilidae sp.4	.	.	.	.	.	.	1	1	ND	R	LF	Z	Omnivore
Muscidae	Drosophilidae sp.5	.	.	.	.	.	.	1	1	ND	R	LF	Z	Omnivore
	Muscidae sp.1	5	1	ND	R	LF	Z	2	1	ND	R	LF	Z	Omnivore
	Muscidae sp.2	.	.	.	.	.	.	1	1	ND	R	LF	Z	Omnivore
	Mycetophilidae sp.1	.	.	.	.	.	.	2	2	ND	R	LF	Z	Omnivore
Mycetophilidae Phoridae	Mycetophilidae sp.2	.	.	.	.	.	.	2	1	ND	R	LF	Z	Omnivore
	Phoridae sp.1	11	6	ND	C	F	Y	60	11	D	VA	VF	W	Detritivore
	Phoridae sp.2	12	5	ND	C	F	Y	53	12	D	VA	VF	W	Detritivore
	Phoridae sp.3	7	4	D	D	LF	Y	40	7	D	VA	VF	Y	Detritivore
	Phoridae sp.4	8	3	ND	D	LF	Z	8	2	ND	C	F	Z	Detritivore
	Phoridae sp.5	.	.	.	.	.	.	15	6	D	C	F	Y	Detritivore
	Phoridae sp.6	.	.	.	.	.	.	1	1	ND	R	LF	Z	Detritivore
	Phoridae sp.7	.	.	.	.	.	.	1	1	ND	R	LF	Z	Detritivore
	Phoridae sp.8	3	3	ND	R	LF	Z	1	1	ND	R	LF	Z	Detritivore

SD = super-dominant; D = dominant; ND = non-dominant; SA = super-abundant; VA = very abundant; A = abundant; C = common; D = dispersed; R = rare; SF = super-frequent; VF = very frequent; F = frequent; LS = less frequent; W = constant; Y = accessory; Z = accidental.

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Taxon	Morphospecies	Conventional						Organic						Functional Group
		No. Individuals Captured	No. collections	Dominance	Abundance	Frequency	Constancy	No. Individuals Captured	No. collections	Dominance	Abundance	Frequency	Constancy	
Philoilidae Psychodidae Sciaridae	Philoilidae sp.1	.	.	.	.	.	.	6	4	ND	D	LF	Y	Omnivore
	Psychodidae sp.1	.	.	.	.	.	.	1	1	ND	R	LF	Z	Omnivore
	Sciaridae sp.1	.	.	.	.	.	.	3	2	ND	R	LF	Z	Omnivore
	Sciaridae sp.2	1	1	ND	R	LF	Z	6	3	ND	D	LF	Z	Omnivore
	Sciaridae sp.3	3	3	ND	R	LF	Z	20	6	D	C	F	Y	Omnivore
	Sciaridae sp.4	.	.	.	.	.	.	1	1	ND	R	LF	Z	Omnivore
	Sciaridae sp.5	8	3	ND	D	LF	Z	43	4	D	VA	VF	Y	Omnivore
	Sciaridae sp.6	3	2	ND	R	LF	Z	32	3	D	VA	VF	Z	Omnivore
	Sciaridae sp.7	.	.	.	.	.	.	1	1	ND	R	LF	Z	Omnivore
	Sciaridae sp.8	.	.	.	.	.	.	1	1	ND	R	LF	Z	Omnivore
	Sciaridae sp.9	2	1	ND	R	LF	Z	1	1	ND	R	LF	Z	Omnivore
	Sciaridae sp.10	2	1	ND	R	LF	Z	1	1	ND	R	LF	Z	Omnivore
Sphaeroceridae	Sciaridae sp.11	1	1	ND	R	LF	Z	1	1	ND	R	LF	Z	Omnivore
	Sciaridae sp.12	1	1	ND	R	LF	Z	.	.	.	.	.	.	Omnivore
	Sphaeroceridae sp.1	.	.	.	.	.	.	23	3	D	A	VF	Z	Omnivore
	Sphaeroceridae sp.2	.	.	.	.	.	.	7	2	ND	D	LF	Z	Omnivore
	Sphaeroceridae sp.3	.	.	.	.	.	.	3	1	ND	R	LF	Z	Omnivore
	Sphaeroceridae sp.4	.	.	.	.	.	.	1	1	ND	R	LF	Z	Omnivore
	Sphaeroceridae sp.5	1	1	ND	R	LF	Z	1	1	ND	R	LF	Z	Omnivore
	Tachinidae sp.1	11	4	ND	C	F	Y	6	1	ND	D	LF	Z	Omnivore
	Ulidiidae sp.1	1	1	ND	R	LF	Z	.	.	.	.	.	.	Omnivore
	Diptera sp.1	.	.	.	.	.	.	2	1	ND	R	LF	Z	Omnivore
	Diptera sp.2	.	.	.	.	.	.	1	1	ND	R	LF	Z	Omnivore
	Diptera sp.3	3	2	ND	R	LF	Z	1	1	ND	R	LF	Z	Omnivore
Tachinidae Ulidiidae	Diptera sp.4	.	.	.	.	.	.	9	1	ND	C	F	Z	Omnivore
	Diptera sp.5	.	.	.	.	.	.	1	1	ND	R	LF	Z	Omnivore
	Diptera sp.6	.	.	.	.	.	.	1	1	ND	R	LF	Z	Omnivore
	Diptera sp.7	.	.	.	.	.	.	1	1	ND	R	LF	Z	Omnivore
	Diptera sp.8	1	1	ND	R	LF	Z	2	1	ND	R	LF	Z	Omnivore
	Aphididae sp.1	24	2	D	C	F	Z	11	3	ND	C	F	Z	Herbivore
	Aetallionidae sp.1	.	.	.	.	.	.	4	3	ND	R	LF	Z	Herbivore
	<i>Mahanarva fimbriolata</i>	2	2	ND	R	LF	Z	11	5	ND	C	F	Y	Herbivore
Cercopidae Coreidae Cydnidae	Coreidae sp.	1	1	ND	R	LF	Z	.	.	.	.	.	.	Herbivore
	<i>Scaptocoris castanea</i>	2	2	ND	R	LF	Z	90	11	D	VA	VF	W	Herbivore
	<i>Cyrtomenus mirabilis</i>	3	2	ND	R	LF	Z	.	.	.	.	.	.	Herbivore

SD = super-dominant; D = dominant; ND = non-dominant; SA = super-abundant; VA = very abundant; A = abundant; C = common; D = dispersed; R = rare; SF = super-frequent; VF = very frequent; F = frequent; LS = less frequent; W = constant; Y = accessory; Z = accidental.

Table 1. (Continued) Number of individuals captured, faunistic indices (dominance, abundance, frequency, constancy) and functional groups of arthropods captured using pitfall traps in conventional and organic sugarcane systems in Jaboticabal, São Paulo, Brazil, during the 2011/2012 and 2012/2013 growing seasons.

Taxon	Morphospecies	Conventional						Organic						Functional Group
		No. Individuals Captured	No. collections	Dominance	Abundance	Frequency	Constancy	No. Individuals Captured	No. collections	Dominance	Abundance	Frequency	Constancy	
Reduviidae	<i>Rasahus</i> sp.	1	1	ND	R	LF	Z	.	.	.	.	.	.	Predator
	Reduviidae sp.	1	1	ND	R	LF	Z	.	.	.	.	.	.	Predator
Hymenoptera Apidae Formicidae	Hemiptera species (nymph)	.	.	.	.	.	.	25	3	D	VA	VF	Z	Herbivore
	<i>Apis mellifera</i>	1	1	ND	R	LF	Z	.	.	.	.	.	.	Pollinator
	<i>Acanthognathus</i> sp.	1	1	ND	R	LF	Z	1	1	ND	R	LF	Z	Predator
	<i>Acromyrmex</i> sp.	21	8	D	C	F	W	10	5	ND	C	F	Y	Herbivore
	<i>Anochetus</i> sp.	2	2	ND	R	LF	Z	.	.	.	.	.	.	Predator
	<i>Atta</i> sp.1	.	.	.	.	.	.	2	2	ND	R	LF	Z	Herbivore
	<i>Atta</i> spp.	33	9	D	VA	VF	W	22	8	D	A	VF	W	Herbivore
	<i>Brachymyrmex</i> spp.	253	14	D	VA	VF	W	498	15	D	VA	VF	W	Omnivore
	<i>Camponotus</i> spp.	184	14	D	VA	VF	W	622	15	SD	SA	SF	W	Omnivore
	<i>Crematogaster</i> spp.	145	8	D	VA	VF	W	1,749	15	SD	SA	SF	W	Predator
	<i>Dolichoderus</i> spp.	63	2	D	VA	VF	Z	8	2	ND	C	F	Z	Predator
	<i>Dorymyrmex</i> spp.	988	15	SD	SA	SF	W	1,026	15	SD	SA	SF	W	Predator
	<i>Ectatomma</i> spp.	66	12	D	VA	VF	W	62	10	D	VA	VF	W	Predator
	<i>Gnamptogenys</i> spp.	25	7	D	A	VF	Y	17	5	D	C	F	Y	Predator
	<i>Hypoanera</i> sp.	8	1	ND	D	LF	Z	9	2	ND	C	F	Z	Predator
	<i>Odontomachus</i> sp.	26	6	D	A	VF	Y	8	5	ND	C	F	Y	Predator
	<i>Pachycondyla</i> sp.	1	1	ND	R	LF	Z	4	2	ND	R	LF	Z	Predator
	<i>Paratrechina</i> sp.	.	.	.	.	.	.	3	2	ND	R	LF	Z	Predator
Vespidae	<i>Pheidole</i> spp.	2,032	15	SD	SA	SF	W	2,716	15	SD	SA	SF	W	Omnivore
	<i>Pseudomyrmex</i> sp.	1	1	ND	R	LF	Z	3	3	ND	R	LF	Z	Predator
	<i>Solenopsis</i> sp.	.	.	.	.	.	.	1	1	ND	R	LF	Z	Omnivore
	<i>Tapinona</i> sp.	98	2	D	VA	VF	Z	7	1	ND	D	LF	Z	Predator
	<i>Trachymyrmex</i> sp.	1	1	ND	R	LF	Z	.	.	.	.	.	.	Herbivore
	<i>Wasmania</i> sp.	.	.	.	.	.	.	10	1	ND	C	F	Z	Predator
	Formicidae sp.1	3	1	ND	R	LF	Z	2	2	ND	R	LF	Z	Predator
	Vespidae sp.1	2	2	ND	R	LF	Z	1	1	ND	R	LF	Z	Predator
	Termitidae sp.1	3	3	ND	R	LF	Z	26	5	D	VA	VF	Y	Detritivore
Lepidoptera Hesperiidae	Hesperiidae sp.1	5	2	ND	R	LF	Z	4	2	ND	R	LF	Z	Herbivore
	Hesperiidae sp.2	1	1	ND	R	LF	Z	1	1	ND	R	LF	Z	Herbivore
	Noctuidae sp.1	1	1	ND	R	LF	Z	2	1	ND	R	LF	Z	Herbivore

SD = super-dominant; D = dominant; ND = non-dominant; SA = super-abundant; VA = very abundant; A = abundant; C = common; D = dispersed; R = rare; SF = super-frequent; VF = very frequent; F = frequent; LS = less frequent; W = constant; Y = accessory; Z = accidental.

Table 1. (Continued) Number of individuals captured, faunistic indices (dominance, abundance, frequency, constancy) and functional groups of arthropods captured using pitfall traps in conventional and organic sugarcane systems in Jaboticabal, São Paulo, Brazil, during the 2011/2012 and 2012/2013 growing seasons.

Taxon	Morphospecies	Conventional						Organic						Functional Group
		No. Individuals Captured	No. collections	Dominance	Abundance	Frequency	Constancy	No. Individuals Captured	No. collections	Dominance	Abundance	Frequency	Constancy	
Pieridae	Pieridae sp. 1	2	1	ND	R	LF	Z	.	.	.	.	.	.	Herbivore
Lepidoptera	Lepidoptera sp. 1.	.	.	.	.	.	.	7	4	ND	D	LF	Y	Herbivore
	Lepidoptera sp. 2.	4	3	ND	R	LF	Z	80	2	D	VA	VF	Z	Herbivore
	Lepidoptera sp. 3 (immature)	2	2	ND	R	LF	Z	.	.	.	.	.	.	Herbivore
Neuroptera														
Chrysopidae	<i>Chrysoperla externa</i>	1	1	ND	R	LF	Z	.	.	.	.	.	.	Herbivore
Hemeroptera	Hemeroptera sp. 1	2	2	ND	R	LF	Z	.	.	.	.	.	.	Omnivore
Orthoptera														
Acrididae	Acrididae sp. 1	11	6	ND	C	F	Y	107	11	D	VA	VF	W	Herbivore
Gryllidae	Acrididae sp. 2	12	3	ND	C	F	Z	.	.	.	.	.	.	Herbivore
	<i>Gryllus assimilis</i>	71	13	D	VA	VF	W	3	2	ND	R	LF	Z	Herbivore
Thysanoptera														
Thysanoptera	Thysanoptera sp. 1	2	1	ND	R	LF	Z	.	.	.	.	.	.	Herbivore
Araneae														
Araneidae	Araneidae sp. 1	1	1	ND	R	LF	Z	.	.	.	.	.	.	Predator
Corinnidae	<i>Corinna</i> sp.	14	7	ND	C	F	Y	81	9	D	VA	VF	W	Predator
	Corinnidae sp. 1	.	.	.	.	.	.	58	5	D	VA	VF	Y	Predator
Ctenidae	Castianeirinae sp. 1	3	1	ND	R	LF	Z	3	3	ND	R	LF	Z	Predator
	Ctenidae sp. 1	.	.	.	.	.	.	1	1	ND	R	LF	Z	Predator
Gnaphosidae	Gnaphosidae sp. 1	1	1	ND	R	LF	Z	.	.	.	.	.	.	Predator
	Gnaphosidae sp. 2	2	1	ND	R	LF	Z	.	.	.	.	.	.	Predator
Hahniidae	Hahniidae sp. 1	1	1	ND	R	LF	Z	.	.	.	.	.	.	Predator
	Hahniidae sp. 2	1	1	ND	R	LF	Z	.	.	.	.	.	.	Predator
Linyphiidae	Hahniidae sp. 3	1	1	ND	R	LF	Z	.	.	.	.	.	.	Predator
	<i>Leptophantes</i> sp.	1	1	ND	R	LF	Z	4	3	ND	R	LF	Z	Predator
Lycosidae	<i>Meioneta</i> sp. 1	1	1	ND	R	LF	Z	3	3	ND	R	LF	Z	Predator
	<i>Meioneta</i> sp. 2	1	1	ND	R	LF	Z	2	2	ND	R	LF	Z	Predator
Miturgidae	Linyphiidae sp. 1	.	.	.	.	.	.	1	1	ND	R	LF	Z	Predator
	Lycosidae sp. 1	2	1	ND	R	LF	Z	1	1	ND	R	LF	Z	Predator
Oonopidae	Lycosidae sp. 2	6	5	ND	R	LF	Y	11	7	ND	C	F	Y	Predator
	<i>Terminius insularis</i>	2	2	ND	R	LF	Z	2	1	ND	R	LF	Z	Predator
Onopidae	Miturgidae sp. 1	1	1	ND	R	LF	Z	23	8	D	A	VF	W	Predator
	Onopidae sp. 1	.	.	.	.	.	.	7	4	ND	D	LF	Y	Predator
								2	1	ND	R	LF	Z	Predator

SD = super-dominant; D = dominant; ND = non-dominant; SA = super-abundant; VA = very abundant; A = abundant; C = common; D = dispersed; R = rare; SF = super-frequent; VF = very frequent; F = frequent; LS = less frequent; W = constant; Y = accessory; Z = accidental.

Table 1. (Continued) Number of individuals captured, faunistic indices (dominance, abundance, frequency, constancy) and functional groups of arthropods captured using pitfall traps in conventional and organic sugarcane systems in Jaboatão, São Paulo, Brazil, during the 2011/2012 and 2012/2013 growing seasons.

Taxon	Morphospecies	Conventional						Organic						Functional Group
		No. Individuals Captured	No. collections	Dominance	Abundance	Frequency	Constancy	No. Individuals Captured	No. collections	Dominance	Abundance	Frequency	Constancy	
Philodromidae	<i>Berlandiella</i> sp.1	.	.	.	.	.	.	1	1	ND	R	LF	Z	Predator
	<i>Berlandiella</i> sp.2	.	.	.	.	.	.	1	1	ND	R	LF	Z	Predator
	Philodromidae sp.1	.	.	.	.	.	.	1	1	ND	R	LF	Z	Predator
Salticidae	Salticidae sp.1	1	1	ND	R	LF	Z	.	.	.	.	.	.	Predator
	Salticidae sp.2	1	1	ND	R	LF	Z	.	.	.	.	.	.	Predator
	Salticidae sp.3	1	1	ND	R	LF	Z	.	.	.	.	.	.	Predator
	Salticidae sp.4	.	.	.	.	.	.	1	1	ND	R	LF	Z	Predator
	Salticidae sp.5	.	.	.	.	.	.	2	1	ND	R	LF	Z	Predator
	Salticidae sp.6	4	1	ND	R	LF	Z	8	4	ND	C	F	Y	Predator
Scytodidae	Scytodes sp.	3	2	ND	R	LF	Z	3	3	ND	R	LF	Z	Predator
	<i>Scytodes ytu</i>	1	1	ND	R	LF	Z	.	.	.	.	.	.	Predator
	Scytodidae sp.1	2	2	ND	R	LF	Z	.	.	.	.	.	.	Predator
Tetragnathidae	Tetragnathidae sp.1	3	2	ND	R	LF	Z	.	.	.	.	.	.	Predator
	<i>Coleosoma</i> sp.1	3	2	ND	R	LF	Z	5	4	ND	R	LF	Y	Predator
	<i>Coleosoma</i> sp.2	3	3	ND	R	LF	Z	.	.	.	.	.	.	Predator
	<i>Dipoena</i> sp.1	7	2	ND	D	LF	Y	5	5	ND	R	LF	Y	Predator
	<i>Dipoena</i> sp.2	5	4	ND	R	LF	Z	29	8	D	VA	VF	W	Predator
	<i>Dipoena</i> sp.3	1	1	ND	R	LF	Z	.	.	.	.	.	.	Predator
Titanocidae	Theridiidae sp.1	.	.	.	.	.	.	2	2	ND	R	LF	Z	Predator
	<i>Goeldia</i> sp.1	4	2	ND	R	LF	Z	9	4	ND	C	F	Y	Predator
	<i>Goeldia</i> sp.2	.	.	.	.	.	.	7	2	ND	D	LF	Z	Predator
	Opiliones sp.1	1	1	ND	R	LF	Z	20	2	D	C	F	Z	Predator
Diplopoda	Diplopoda sp.1	24	11	D	C	F	W	74	10	D	VA	VF	W	Detritivore
Chilopode	Chilopode sp.1	15	8	ND	C	F	W	.	.	.	.	.	.	Detritivore
	Chilopode sp.2	.	.	.	.	.	.	13	3	ND	C	F	Z	Detritivore
Ecological Indices		Conventional						Organic						
Shannon–Wiener (H')		2.343						2.455						
Margalef (α)		14.101						17.063						
Equitability		0.488						0.487						
Similarity		0.687						0.687						

SD = super-dominant; D = dominant; ND = non-dominant; SA = super-abundant; VA = very abundant; A = abundant; C = common; D = dispersed; R = rare; SF = super-frequent; VF = very frequent; F = frequent; LS = less frequent; W = constant; Y = accessory; Z = accidental.

Table 2. Distribution of morphospecies in relation to faunistic indices of dominance, abundance, frequency, and constancy captured in pitfall traps in conventional and organic sugarcane systems in Brazil (2011/12 and 2012/13 growing seasons).

Faunistic index	Classification	No. of morphospecies (%)		Total no. of individuals (%)	
		Conventional	Organic	Conventional	Organic
Dominance	SD	2 (1.6)	4 (2.8)	3,020 (60.84)	6,110 (73.8)
	D	17 (14)	30 (21.2)	1,577 (31.76)	1,825 (22.04)
	ND	103 (84.4)	108 (76)	367 (7.4)	345 (4.16)
Abundance	SA	2 (1.65)	4 (2.8)	3,020 (60.84)	6,110 (73.8)
	VA	11 (9.1)	19 (13.4)	1,450 (29.21)	1,598 (19.3)
	A	2 (1.6)	5 (3.5)	51 (1.03)	114 (1.38)
	C	16 (13.2)	19 (13.4)	218 (4.39)	238 (2.87)
	D	7 (5.7)	9 (6.3)	53 (1.07)	60 (0.72)
	R	84 (68.8)	86 (60.6)	172 (3.46)	160 (1.93)
Frequency	SF	2 (1.6)	4 (2.8)	3,020 (60.84)	6,110 (73.8)
	VF	13 (10.6)	24 (16.9)	1,501 (30.24)	1,712 (20.68)
	F	16 (13.2)	19 (13.4)	218 (4.4)	238 (2.87)
	LF	91 (74.6)	95 (66.9)	225 (4.52)	220 (2.65)
Constancy	W	11 (9)	16 (11.3)	3,832 (77.2)	7,253 (87.6)
	Y	16 (13.2)	23 (16.2)	193 (3.89)	457 (5.52)
	Z	95 (77.8)	103 (72.5)	939 (18.91)	570 (6.88)

SD = super-dominant; D = dominant; ND = non-dominant; SA = super-abundant; VA = very abundant; A = abundant; C = common; D = dispersed; R = rare; SF = super frequent; VF = very frequent; F = frequent; LS = less frequent; W = constant; Y = accessory; Z = accidental.

were important and classified as dominant. For example, leaf-cutting ants of the genus *Atta* F. are important pests in sugarcane crops and were found in both sugarcane fields. Moreover, other beneficial ant genera (*Brachymyrmex* Mayr, *Ectatomma* Smith, and *Gnamptogenys* Roger) were found in both organic and conventional systems. Also, we found dominant morphospecies of spiders (4 morphospecies in total) only in the organic system.

Abundance

Similarly to the observed dominance, morphospecies of the genera *Pheidole* and *Dorymyrmex* were super-abundant in both the conventional and organic systems, but morphospecies of the genera *Camponotus* and *Crematogaster* were super-abundant only in the organic system. These 2 latter genera, on the other hand, were classified as very abundant in the conventional system. The super-abundant and very abundant species are native species that behave as invaders in a disturbed environment (Silva-Mattos & Pivello 2009). The conventional system presented a greater number of rare morphospecies (84) compared with common (16), very abundant (11), occasional (7), and abundant (2) morphospecies. A similar trend was observed for the organic system, in which a greater number of rare morphospecies (86) were observed compared with common (19), very abundant (19), and occasional morphospecies (9), as well as abundant (5) morphospecies. Therefore, 69.4% and 61.1% of the morphospecies were rare in the conventional and organic system, respectively. Twenty-eight morphospecies (14.7% of the total morphospecies) considered to be rare occurred on both systems.

Frequency

The super-frequent morphospecies (i.e., species that occurred on all sampling dates) were the same as the super-abundant and the super-dominant morphospecies. Ants of the genera *Camponotus*, *Crematogaster*, *Dorymyrmex*, and *Pheidole* were the super-frequent morphospecies in the organic system, whereas only the last 2 genera were super-frequent in the conventional system (Tables 1 and 2).

Most morphospecies were considered uncommon, i.e., their occurrence was below 34% of the samplings, in both systems. In this category, 91 (47.9%) and 95 (51%) morphospecies occurred at low frequencies in the conventional and organic systems, respectively.

Constancy

In this category, most morphospecies occurred accidentally. In the conventional system, 95 morphospecies were considered accidental, 16 accessories, and 11 constant, whereas in the organic system, there were 103 accidental, 23 accessories, and 16 constant morphospecies (Table 1). The most constant morphospecies were the ants with 30% and 40% of total morphospecies in the organic and the conventional system, respectively.

We also analyzed the specimens classified among the high faunistic values (super-dominant, dominant, super-abundant, very abundant, abundant, super-frequent, very frequent, frequent, and constant), and we found 22 (8 predators, 5 omnivores, 5 detritivores, and 4 herbivores) and 10 (5 predators, 3 omnivores, and 2 herbivores) species for organic and conventional systems, respectively. Among the morphospecies collected, 7 were common in both systems (3 omnivores [*Brachymyrmex* spp., *Camponotus* spp., *Pheidole* spp.], 3 predators [*Crematogaster* spp., *Dorymyrmex* spp., *Ectatomma* spp.], and 1 herbivore [*Atta* spp.]), 3 were found exclusively in the conventional system (2 predators and 1 herbivore), and 15 were found exclusively in the organic system (5 predators, 5 detritivores, 2 omnivores, and 3 herbivores).

Estimates of species richness were similar and close to the expected number of species as shown in Fig. 1. The species curves for both organic and conventional sugarcane systems tended to stabilize (plateau) when 15 samples were taken on a monthly basis. Thus, the number of monthly collections and traps adopted in the study was adequate for assessing species diversity in sugarcane agroecosystems.

A comparison of the edaphic arthropods by using principal component analysis indicated that there was a difference between the sugarcane systems. Eleven samples (73.3%) out of 15 taken from the organic sugarcane field presented negative values, whereas 13 samples

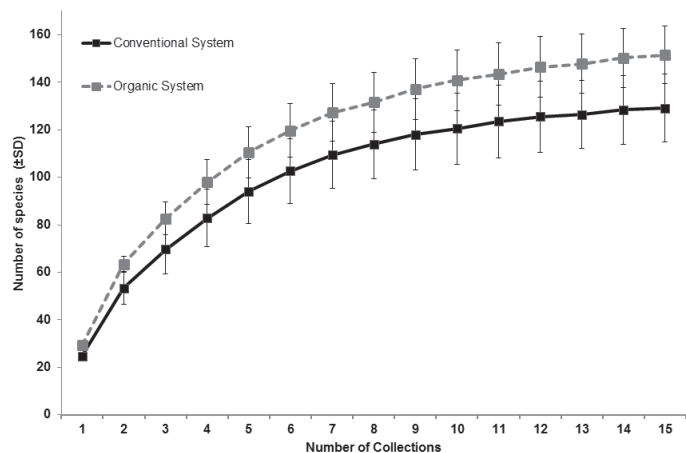


Fig. 1. Curve estimating species richness of edaphic arthropods in conventional and organic sugarcane fields in Jaboticabal, São Paulo, Brazil. Error bars represent the standard deviation.

(86.7%) out of 15 from the conventional field presented positive values (Fig. 2). Therefore, the communities were different both quantitatively and qualitatively. Ants generally were the most abundant species found on both systems, but they were more abundant in the organic than the conventional system.

Discussion

Twice as many soil-dwelling arthropods were captured in the organic cropping system, relative to conventional sugarcane production. Also, greater numbers of arthropod predators and omnivores and smaller numbers of herbivores were captured in the organic field. This is shown by the high dominance and frequency of coleopteran and hymenopteran predators in the organic system. These predators are re-

ported as important natural control agents of several pests that occur in different stages of sugarcane development (Mendonça & Marques 2005; Costa et al. 2007; Silva et al. 2009).

Our results also suggest that compared with the conventional system, the organic system could provide greater availability and abundance of resources such as pollen, nectar, and alternative sources of food and shelter, which favor the abundance and diversity of species (Landis et al. 2000). Root (1973) noted that systems providing appropriate conditions (food and shelter) tend to have greater abundance of arthropod predators and omnivores, and therefore greater potential for biological control of herbivores.

Among the predators, ant and spider species occurred frequently in both systems. Several studies have shown ants in the genera *Crematogaster*, *Dorymyrmex*, *Ectatomma*, and *Pheidole* are effective control agents of pests in sugarcane (Rossi & Fowler 2002, 2004; Araújo et al. 2004, 2005; Pereira et al. 2004; Philpott et al. 2008; Schatz et al. 2008), but their population dynamics remain to be better understood. For spiders in the sugarcane agroecosystem, there is practically no published information about their diversity, although this work suggests that they are especially abundant elements in organic sugarcane production, and their contribution should be assessed.

Pheidole spp. and *Dorymyrmex* spp. ants were the only super-dominant species in the conventional system. Thus, even with the disruption caused by the use of fire in conventional harvesting, populations of these species were not affected, as also observed by Araújo et al. (2004). Ant species generally present rapid colony restructuring, wide foraging area, and social organization. These features may have contributed to the high abundance, frequency, and constancy in the conventional area (Rossi & Fowler 2002; Araújo et al. 2004, 2005). The fact that most super-dominant species were detected in the organic system (*Pheidole* spp., *Dorymyrmex* spp., *Crematogaster* spp., and *Camponotus* spp.) may be related to the nesting strategy of these species. According to Longino (2003), *Crematogaster* and *Dorymyrmex* ants have shallow nests and therefore may be more affected by the fire, so the reestablishment of colonies requires longer period of time.

However, it is not only the direct effects of fire that can lead to reduced biodiversity in the conventional system. Herbicides are used to control weeds whose elimination can indirectly affect the population of arthropods. Many herbivorous insects feed on the weeds that occur in crops (Chiverton & Sotherton 1991). Arthropod predators and parasitoids also can utilize these weeds to supplement their diet by feeding on pollen and nectar. Thus, herbicides can affect biodiversity, either by acting directly (on herbivores) or indirectly (on predators and omnivores).

In spite of the higher species richness in the organic than the conventional system, both the faunistic and diversity indices were generally similar. We hypothesize that this finding may be related to surrounding sugarcane areas, which may have been used as a shelter or refuge for some arthropods, especially beneficial arthropods, during harvest in the 2 systems. Although this needs to be further studied in this agroecosystem, harvesting the sugarcane at different times may facilitate the movement of arthropods between different areas, allowing the reoccupation of the disturbed environment more quickly.

The study of arthropod biodiversity may allow us to identify important naturally occurring beneficials in the agroecosystem. Despite having used only pitfall trapping, this study provided comparative information on biodiversity in the sugarcane agroecosystem under 2 management systems. This new information may assist future studies on biological control, or even risk assessment of genetically modified sugarcane, where it is essential to know the diversity of arthropods. Overall, our results indicated that the organic management of sugarcane improves the abundance and diversity of arthropods (especially predators and omnivores) relative to conventional management.

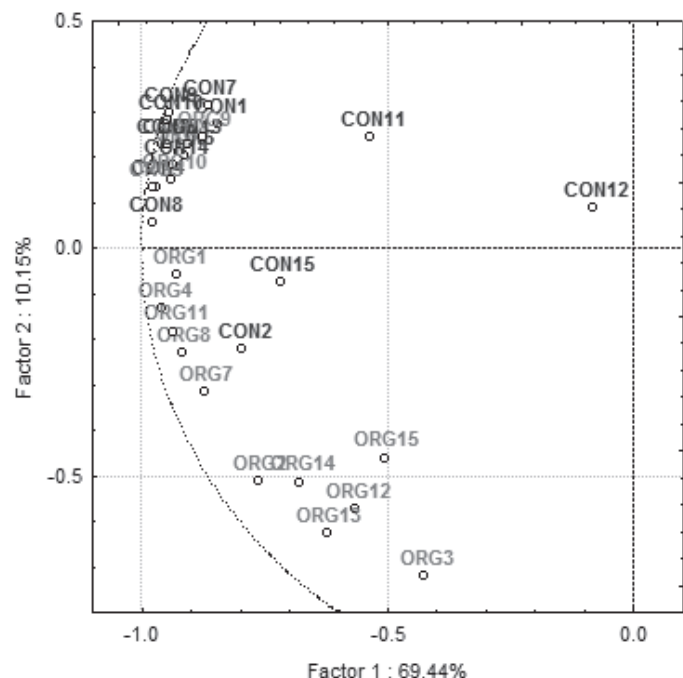


Fig. 2. Principal component analysis of the arthropod communities in organic (ORG) and conventional (CON) sugarcane fields.

Acknowledgments

The authors are grateful to Antonio Brescovit and João Lucas Chavari, Instituto Butantan, São Paulo, Brazil, who kindly identified the spiders. Scholarship to the first author was provided by CAPES, Brazil.

References Cited

- Abreu RRL, Lima SS, Oliveira NCR, Leite LFC. 2014. Fauna edáfica sob diferentes níveis de palhada em cultivo de cana-de-açúcar. *Pesquisa Agropecuária Tropical* 44: 409–416.
- Araújo MS, Della Lucia TMC, Picanço MC. 2004. Impacto da queima da palhada da cana-de-açúcar no ritmo diário de forrageamento de *Atta bisphaerica* Forel (Hymenoptera: Formicidae). *Revista Brasileira de Zoologia* 2: 33–38.
- Araújo RA, Araújo MS, Gönring AHR, Guedes RNC. 2005. Impacto da queimada controlada da palhada da cana-de-açúcar sobre a comunidade de insetos locais. *Neotropical Entomology* 34: 649–658.
- Baccaro FB. 2006. Chave para as principais subfamílias e gêneros de formigas (Hymenoptera: Formicidae). Instituto Nacional de Pesquisas da Amazônia, Programa de Pesquisa em Biodiversidade, Faculdades Cathedral, Barra do Garças, Mato Grosso, Brazil.
- Borror DJ, Triplehorn CA, Johnson NF. 1992. *An Introduction to the Study of Insects*, 6th Edition. Saunders College Publishing, Philadelphia, Pennsylvania.
- Butler SJ, Vickery JA, Norris K. 2007. Farmland biodiversity and the footprint of agriculture. *Science* 315: 381–384.
- Castelo Branco RT, Portela GLF, Barbosa OAA, Silva PRR, Pádua LEM. 2010. Análise faunística de insetos associados à cultura da cana-de-açúcar, em área de transição floresta amazônica- cerrado (mata de cocal), no município de União-Piauí, Brasil. *Semina* 31: 113–1120.
- Chiverton PA, Sotherton NW. 1991. The effects of beneficial arthropods of the exclusion of herbicides from cereal crop edges. *Journal of Applied Ecology* 28: 1027–1039.
- Colwell RK. 2006. EstimateS: Statistical Estimation of Species Richness and Shared Species from samples. Version 8.0 User's Guide and Application, <http://viceroy.eeb.uconn.edu/estimates> (last accessed 9 Dec 2016).
- Costa NP, Oliveira HD, Brito CH, Silva AB. 2007. Influência do nim na biologia do predador *Euborellia annulipes* e estudo de parâmetros para a sua criação massal. *Revista de Biologia e Ciências da Terra* 7: 1–10.
- Eilenberg J, Hajek A, Lomer C. 2001. Suggestions for unifying the terminology in biological control. *Biocontrol* 46: 387–400.
- Gurr GM, Wratten SD, Barbosa P. 2000. Success in conservation biological control of arthropods, pp. 105–132 *In* Gurr G, Whitten S [eds.], *Biological Control: Measures of Success*. Kluwer Academic Press, Dordrecht, Netherlands.
- Heltshe JF, Forrester NE. 1983. Estimating species richness using the Jackknife procedure. *Biometrics* 39: 1–11.
- Landis DA, Wratten SD, Gurr GM. 2000. Habitat management to conserve natural enemies of arthropod pests in agriculture. *Annual Review of Entomology* 45: 175–201.
- Landis DA, Menalled FD, Costamagna AC, Wilkinson TK. 2005. Manipulating plant resources to enhance beneficial arthropods in agricultural landscapes. *Weed Science* 53: 902–908.
- Letourneau DK. 1998. Conserving biology, lessons for conserving natural enemies, pp. 9–38 *In* Barbosa P [ed.], *Conservation Biological Control*. Academic Press; San Diego, California.
- Longino JT. 2003. The *Crematogaster* (Hymenoptera, Formicidae, Myrmicinae) of Costa Rica. *Zootaxa* 151: 1–150.
- Loureiro MC, Queiroz RMVB. 1990. Insetos de Viçosa—Formicidae. Universidade Federal de Viçosa, Viçosa, Minas Gerais, Brazil.
- Mendonça AF, Marques EJ. 2005. Cigarrinha da folha *Mahanarva posticata* (Stål) (Hemiptera: Cercopidae), pp. 295–301 *In* Mendonça AF [ed.], *Cigarrinhas da Cana-de-Açúcar*. Insecta. Maceió, Alagoas, Brazil.
- Moraes RCB, Haddad ML. 2003. Software para análise faunística-ANAFU, p.195 *In* Simpósio de Controle Biológico, 8. São Pedro. Resumos. Sociedade Entomológica do Brasil, Piracicaba, Brazil.
- Nunes L, Silva I, Pitê M, Rego F, Leather S, Serrano A. 2006. Carabid (Coleoptera) community change following prescribed burning and the potential use of carabids as indicator species to evaluate the effects of fire management in Mediterranean regions. *Silva Lusitana* 14: 85–100.
- Oliver I, Beattie AJ. 1996. Invertebrate morphospecies as surrogates for species: a case study. *Conservation Biology* 10: 99–109.
- Pasqualim LA, Dionísio JA, Zawadneak MAC, Marçal CT. Macrofauna edáfica em lavouras de cana-de-açúcar e mata no noroeste do Paraná, Brasil. *Semina* 33: 7–18.
- Pereira JA, Bento A, Cabanas JE, Torres LM, Herz A, Hassan SA. 2004. Ants as predators of the egg parasitoid *Trichogramma cacoeciae* (Hymenoptera: Trichogrammatidae) applied for biological control of the olive moth, *Prays oleae* (Lepidoptera: Plutellidae) in Portugal. *Biocontrol Science and Technology* 14: 653–664.
- Philpott SM, Perfecto I, Vandermeer J. 2008. Behavioral diversity of predatory arboreal ants in coffee agroecosystems. *Environmental Entomology* 37: 181–191.
- Pogue DW, Schnell GD. 2001. Effects of agriculture on habitat complexity in a prairie-forest ecotone in the southern Great Plains of North America. *Agriculture, Ecosystems & Environment* 87: 287–298.
- Root RB. 1973. Organization of plant–arthropod association in simple and diverse habitats: the fauna of collards (*Brassica oleraceae*). *Ecological Monographs* 43: 95–124.
- Rossi MN, Fowler HG. 2002. Manipulation of fire ant density, *Solenopsis* spp., for short-term reduction of *Diatraea saccharalis* larval densities in Brazil. *Scientia Agrícola* 59: 389–392.
- Rossi MN, Fowler HG. 2004. Predaceous ant fauna in new sugarcane fields in the state of São Paulo, Brazil. *Brazilian Archives of Biology and Technology* 47: 805–811.
- Schatz B, Kjellberg F, Nyawa S, Hossaert-McKey M. 2008. Fig wasps: a staple food for ants on *Ficus*. *Biotropica* 40: 190–195.
- Silva AB, Batista JL, Brito CH. 2009. Capacidade predatória de *Euborellia annulipes* (Lucas, 1847) sobre *Spodoptera frugiperda* (Smith, 1797). *Acta Scientiarum* 31: 7–11.
- Silva-Mattos DM, Pivello VR. 2009. O impacto das plantas invasoras nos recursos naturais de ambientes terrestres—alguns casos brasileiros. *Ciência e Cultura* 61: 27–30.
- Silveira Neto S, Nakano O, Barbin D, Nova NAV. 1976. *Manual de Ecologia de Insetos*. Agronômica Ceres, São Paulo, Brazil.
- Suguituru SS, Morini MSC, Feitosa RM, Silva RR [eds.]. 2015. *Formigas do Alto Tietê*. Canal 6 Editora, Bauru, São Paulo, Brazil.
- Wilkinson TK, Landis DA. 2005. Habitat diversification in biological control: the role of plant resources, pp. 305–325 *In* Wäckers FL, van Rijn PCJ, Bruin J [eds.], *Plant-Provided Food for Carnivorous Insects. A Protective Mutualism and its Applications*. Cambridge University Press, Cambridge, United Kingdom.