

# Allometric and Non-Allometric Patterns in Sexual Dimorphism Discrimination of Wing Shape in Ophion intricatus: Might Two Male Morphotypes Coexist?

Authors: Benítez, Hugo A., Bravi, Raffaella, Parra, Luis E., Sanzana, Maria-Jose, and Sepúlveda-Zúñiga, Einer

Source: Journal of Insect Science, 13(143): 1-10

Published By: Entomological Society of America

URL: https://doi.org/10.1673/031.013.14301

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at <u>www.bioone.org/terms-of-use</u>.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.



# Allometric and non-allometric patterns in sexual dimorphism discrimination of wing shape in *Ophion intricatus*: Might two male morphotypes coexist?

Hugo A. Benítez<sup>1,2a</sup>\*, Raffaella Bravi<sup>3b</sup>, Luis E. Parra<sup>4c</sup>, Maria-Jose Sanzana<sup>4d</sup>, Einer Sepúlveda-Zúñiga<sup>4e</sup>

<sup>1</sup>Faculty of Life Sciences, University of Manchester, Michael Smith Building, Oxford Road, Manchester M13 9PT, UK
 <sup>2</sup>Instituto de Alta Investigación, Universidad de Tarapacá, Casilla 7-D Arica, Chile
 <sup>3</sup>Environmental Biology Department, University Roma Tre, V.le G. Marconi 446, 00146, Rome, Italy
 <sup>4</sup>Departamento de Zoología, Facultad de Ciencias Naturales y Oceanográficas, Universidad de Concepción, Concepción, Casilla: 160-C - 4070386, Chile

## Abstract

Bees and wasps could exhibit shape and size sexual dimorphism, and most of their morphological variation could depend on phenotypic responses due to environmental pressure during ontogenetic development. More complex measurement techniques related to size and shape rather than simply to mass and length should be required to analyze such a complex sexual dimorphism. In this study, differences related to wing shape and size of males and females of Ophion intricatus Brullé (Hymenoptera: Ichneumonidae) were evaluated using geometric morphometrics. Right and left wings of specimens were used, and a photographic matrix was constructed in which 18 morphological landmarks based on shape and vein patterns of the wings were digitalized. A multivariate analysis of wing shape showed significant differences between sexes and sites. The geometric variation demonstrated that the points at the intersection of radial and cubital-anal veins might be key characters to differentiate between sexes. This study also showed the presence of two clearly different male morphotypes coexisting in the same study site. However, it should be noted that the results of this study showed that the variation in wing shape is an analytical character in the determination of sexual differences in the family Ichneumonidae. These differences raise the question of whether sexual dimorphism of wing shape may be modulated by natural selection.

| Keywords: allometry, geometric morphometrics, sexual selection, sexual shape dimorphism, wasp   |
|---|
| Abbreviations: PCA, principal component analysis  |
| Correspondence: a hugo.benitez@postgrad.manchester.ac.uk, b raffaella.bravi@uniroma3.it, c luparra@udec.cl, d mariajosanza-                 |
| na@udec.cl, e <u>einersepulveda@udec.cl</u> , *Corresponding author   |
| Editor: Paul Ode was editor of this paper.  |
| Received: 5 April 2012 Accepted: 11 June 2013 Published: 4 December 2013  |
| <b>Copyright:</b> This is an open access paper. We use the Creative Commons Attribution 3.0 license that permits unrestricted use, provided |
| that the paper is properly attributed.  |
| ISSN: 1536-2442   Vol. 13, Number 145   |
| Cite this paper as:   |

Benítez HA, Bravi R, Parra LE, Sanzana M-J, Sepúlveda-Zúñiga E. 2013. Allometric and non-allometric patterns in sexual dimorphism discrimination of wing shape in *Ophion intricatus*: Might two male morphotypes coexist? *Journal of Insect Science* 13:143. Available online: <u>http://www.insectscience.org/13.143</u>

#### Introduction

Bees and wasps, as most insects, exhibit sexual size and shape dimorphism (e.g., Hurlbutt 1987; Mackauer 1996; O'Neill 2001; Pretorius 2005; Mitrovski-Bogdanović et al. 2009; Zikic et al. 2009). Most of the morphological variation in wasps could be due to phenotypic responses (plasticity) that act during ontogenetic development as a consequence of environmental pressures (Shreeves and Field 2008). Females have some adaptive advantages over males, such as greater fecundity and parental care, because females are typically larger than males (Forrest 1987; Andersson 1994; Moller and Zamora-Muñoz 1997; Reeve and Farbain 1999). However, in some species, males are longer but have less mass than females (e.g., Cepeda-Pizarro et al. 1996). Those considerations suggest that inferring and quantifying sexual dimorphism requires more complex measurement techniques related to shape and size rather than simply to mass and length (Gidaszewski et al. 2009; Benítez et al. 2010). Another important factor to be evaluated within sexual dimorphism is allometry, defined as the association between size and shape, or the covariation of parts due to variation in size. All types of allometry can contribute to the sexual differentiation at the respective levels of variation (Gidaszewski et al. 2009). Allometry is classified according to the cause of variation in size that gives

rise to the allometric relationship, which is usually ontogenetic growth or evolutionary change; therefore, there is either ontogenetic or evolutionary allometry, respectively. If individuals of the same species are examined at the same ontogenetic stage, both of these sources of "dynamic" variation are kept constant, and the remaining individual variation would be "static," corresponding to static allometry (Klingenberg 1996).

Ophionines are solitary koinobiont endoparasitoids of the larvae of other holometabolous insects. Many ophionines oviposit into relatively mature larvae in their 3<sup>rd</sup> or 4<sup>th</sup> (Vickery 1929) or 4<sup>th</sup> or 5<sup>th</sup> instar (Moutia and Courtois 1952; Yu and Horstmann 1997), though some species attack young larvae (Price 1975). Generally a single egg is deposited free in the haemocoel. No species of ophionine are known to develop gregariously (Gauld and Bolton 1988; Yu and Horstmann 1997). The genus Ophion Fabricius (Hymenoptera: Ichneumonidae) includes about 200 species (Gauld 1978) distributed worldwide, and most of them inhabit temperate climates (Townes 1971). Most Ophion species are parasitoids, laying their eggs in Lasiocampidae, Noctuidae, and Notodontidae pupae or larvae.

In the present study, static allometry is analysed on the species *Ophion intricatus* Brullé (Hymenoptera: Ichneumonidae). Geometric

morphometrics techniques were used, with the aim to investigate sexual dimorphism of wings and to characterize the wings as a trait of sexual differentiation, as well as to show that there is a differentiation pattern in males of the same species coexisting in the same habitat. This is the first step for further studies on behavior at group level in the area.

#### **Materials and Methods**

#### Data acquisition

A total of 136 specimens of O. intricatus (50 female and 86 males) were used in this study, and they were collected in the Botanical Park "Pedro del Rio Zañartu" - Bio Bio Region, Chile (36° 47' S, 73° 09' W). Wasps were collected with UV light traps (GE Power Pro 950, www.ge.com) using an 800watt electric generator. The light sources were placed over a white sheet to increase the luminosity. Traps were installed for 4 hr in different sampling points. Following the key of Gauld (1978), specimens were carefully checked for their species status, and the process was based on taxo-morphological characters. Once collected, specimens were processed and mounted. Right and left wings were removed from the body and then mounted in fixed molds (compressed between two microscope slides) for subsequent image capturing.

#### **Morphometric analysis**

Right and left wings of wasps were photographed with a Sony 10 DSC-H7 camera (<u>www.sony.com</u>) with directed fiber optics lighting. Then, photographic matrixes were constructed using TpsUtil 1.57 program (Rohlf 2008). Eighteen morphological landmarks (Type 1 landmarks) were digitized for left and right wings in all specimens, selected according to external anatomy and vein pattern of the wings (Figure 1, Table 1) (Zelditch et al. 2012). For this purpose, TpsDig 2.12 software (Rohlf 2008b) was used. The x-y coordinates of the biologically homologous landmarks were then aligned and superimposed with the method of minimum least squares based on the generalized procrustes analysis (Dryden and Mardia 1998; Rohlf and Slice 1990).

Once the Cartesian x-y coordinates for all points were obtained, shape information was extracted from the coordinate data by using a full Procrustes fit (Rohlf and Slice 1990; Dryden and Mardia 1998). Due to the matching symmetry of the wings, the dataset contained two separate landmark configurations, one for the left and one for the right side. The analysis of shape included the reflection of all configurations from one body side to its mirror image (for details see Klingenberg and McIntyre 1998; Klingenberg et al. 2002). After reflection, all configurations can be superimposed in the Procrustes fit, a procedure that removes in formation on location orientation and rotation and standardizes each specimen to unit centroid size, which is the square root of the

**Table I.** Anatomical description of landmarks in the wing of *Ophion intricatus*. A = Anal; Cu = Cubital; M = Medium; R = Radial; Rs = Sector radial; <math>M+Cu = Medium cubital; m = Medium transversal; cu = Cubital transversal; m+cu = Medium cubital transversal.

| Landmark | Anatomical structure |  |  |  |  |
|----------|----------------------|--|--|--|--|
| #1       | Proximal Base M+Cu1  |  |  |  |  |
| #2       | Distal Base 1A       |  |  |  |  |
| #3       | Distal Base M+Cu1    |  |  |  |  |
| #4       | Distal Base A        |  |  |  |  |
| #5       | Proximal Base Cu1b   |  |  |  |  |
| #6       | Proximal Base 1m-cu  |  |  |  |  |
| #7       | Prior Base M         |  |  |  |  |
| #8       | Prior Base of stigma |  |  |  |  |
| #9       | Distal Base Cu1b     |  |  |  |  |
| #10      | Proximal Base M      |  |  |  |  |
| #11      | Prior Base Rs        |  |  |  |  |
| #12      | Proximal Base M      |  |  |  |  |
| #13      | Distal Base Rs       |  |  |  |  |
| #14      | Distal Base Cu2b     |  |  |  |  |
| #15      | Distal Base Cula     |  |  |  |  |
| #16      | Distal Base M        |  |  |  |  |
| #17      | Prior Base Rs        |  |  |  |  |
| #18      | Proximal Base R1     |  |  |  |  |

summed squared Euclidean distances from each landmark to the specimen centroid and provides an estimate of the size of the studied structure (Dryden and Mardia 1998). Centroid size was used as a measure of whole body size, while the information about the shape variation was extracted from the Procrustes superimposition.

After performing the Procrustes fit, the covariance matrix generated from the Procrustes coordinates, and which included the measures of the association between Procrustes coordinates (the x-y coordinates of each landmark after the Procrustes superimposition), were extracted. Shape variation in the entire dataset was then analyzed with principal component analysis (PCA) based on decomposition of covariance matrix.

Additionally, to assess shape and size variation between sexes, Procrustes ANOVA was performed, and the results are reported as sums of squares and mean squares that are dimensionless (Klingenberg and McIntyre 1998; Klingenberg et al. 2002).

Static allometry was evaluated by a multivariate regression of shape, pooled within sex (Procrustes coordinates), on centroid size. Then, the predicted values were used as shape variables, accounting for the allometric component of shape variation, and the residuals were used as the non-allometric component of shape variation. A plot of the regression scores (the projection of shape data onto the direction of the regression vector in the shape tangent space) was performed to better visualize allometric relarelationships (Drake and Klingenberg 2008; Klingenberg et al. 2012). All morphometric and statistical analyses were performed using the free morphometric software Morpho J 1.05f (Klingenberg 2011).

#### Results

PCA for allometric and non-allometric components of within-sex variation, showed how the first principal components accumulated shape variation in very few dimensions (Figure 2).

Regarding allometric components, the first three PCs accounted for 46.6% (PC1 + PC2 + PC3 = 18.178% + 16.365% + 12.061%) of the total shape variation, and provided a reasonable estimation of the total variation. The other PCs each accounted for no more than 8.8% of the variation.

Regarding non-allometric components, PC1 held more than 30% of the complete variation between sexes, and the remnant PCs each accounted for no more than 13.6% (Figure 3, 4). Procrustes ANOVA showed that variation in size and symmetric shape between sexes was highly significant (Table 2). To quantify static allometry, multivariate regression of shape (pooled within sexes) was used on centroid size. The results confirmed the occurrence of a strong static allometric effect that accounted for 30.48% of the total shape variance (p < 0.0001).

**Table 2.** Procrustes ANOVA for both centroid size (CS) and shape (SH) of Ophion intricatus. Sums of squares (SS) and mean squares (MS) are in units of Procrustes distances (dimensionless).

|    |            | SS         | MS          | df   | F     | р        | Pillai tr. | p (param) |
|----|------------|------------|-------------|------|-------|----------|------------|-----------|
| Ĩ. | Sex        | 0.000051   | 0.000025    | 2    | 133.5 | < 0.0001 |            |           |
| CS | Individual | 0.000012   | 0.0000001   | 65   | 2.09  | 0.0016   |            |           |
| Co | Side       | 0.000001   | 0.000001    | 1    | 1.55  | 0.217    |            |           |
|    | Ind*Side   | 0.000006   | 0.0000001   | 67   |       |          |            |           |
| SH | Sex        | 0.11457622 | 0.001790254 | 64   | 42.37 | < 0.0001 | 1.68       | < 0.0001  |
|    | Individual | 0.08788273 | 4.22513E-05 | 2080 | 3.5   | < 0.0001 | 22.16      | < 0.0001  |
|    | Side       | 0.00251535 | 7.86046E-05 | 32   | 6.51  | < 0.0001 | 0.83       | < 0.0001  |
|    | Ind*Side   | 0.02587538 | 1.20687E-05 | 2144 |       |          |            |           |

#### Discussion

The results of this study confirmed the presence of sexual variation in wing shape in O. intricatus. A number of studies have found that sexual dimorphism associated with size and shape in insects is adaptive (Nylin and Gotthard 1998; Farbain et al. 2007; Gidaszewski et al. 2009; Benitez et al. 2013). Some of the hypotheses proposed to explain this kind of dimorphism in arthropods do not take into account whether the differences could be due exclusively to sexual dimorphism or to allometric processes (Walker and Rypstra 2001; Esperk et al. 2007; Gidaszewski et al. 2009). According to Zikic et al. (2010), allometry contributes to the variation in body shape among female biotypes of Ephedrus persicae. Several morphological characters, such as size of head, width of petiolus, width of mesoscutum, and length of ovipositor sheath, diverged among the *E. persicae* biotypes. In our study, the influence of size on sexual differences was taken into account, and also if there was some effect on the shape differentiation (Klingenberg 1996; Fairbairn 1997; Debat et al. 2003).

The wings of O. intricatus were evaluated, and it was determined that they were strongly under sexual dimorphism. The results show that females tended to have longer wings than males, as seen in the displacement of landmarks 4, 7, and 8 (the connection of the transversal radial and cubital-anal veins). This type of wing dimorphism is present in almost all parasitoids within Ichneumonidae, as the females' metasomas are likely to be a bit heavier than males because the females carry eggs. This extra weight could result in slight compensatory differences in wing size and shape. This kind of variation may be an adaptive response to flight, and for behavioral activities such as localization, foraging, and oviposition. In contrast, males' wings are wider, which might be linked to the selection of courting sites (longer flight periods in search of sexually active females) and to predator avoidance.

A differentiation of wing shape between two male populations in the same sampling site was found, and two male morphotypes seemed to coexist. The presence of these two male morphotypes might be the effect of sexual differentiation due to females not preferring a particular male wing shape. However, it must be taken into account that male populations used different microhabitats. The light traps used could reasonably have attracted specimens from different microhabitats during the collecting process, and males may have morphologic microadaptation to their habitat. Zikic et al. (2010) reported that wing shape characters related to allometry differed among female specimens of E. persicae that emerged from different aphid hosts, indicating differences in their ecology and physiology.

It is expected that the anatomical design of the wings of insects will be subjected to strong selection. Here, several wing morphotypes emerged, raising the question of whether sexual dimorphism on wing shape may be modulated by natural selection (Betts and Wootton 1988; Dudley 2000).

Behavioral studies are needed in order to clarify what the adaptive significance of the findings of our study are. However, it is worth noting that the results of this study showed that variation in wing shape was an analytical character in the determination of sexual differences in the individuals of *O*. *intricatus*.

#### Acknowledgements

The authors thank Project DIUC 210.113.079-1.0 of the Dirección de Investigación of the Universidad de Concepción for financing this study. We also thank Maria Raquel Lazo de la Vega for her help in the English proofreading of the manuscript, and Carla Sanzana. Carlos Zamora, and Katherine Avendaño for assistance with the graphs of the article.

### References

Andersson M. 1994. *Sexual selection*. Princeton University Press.

Benítez H, Vidal M, Briones R, Jerez V. 2010. Sexual dimorphism and population morphological variation of *Ceroglossus chilensis* (Eschscholtz) (Coleoptera, Carabidae). *Journal of the Entomological Research Society* 12(2): 87–95.

Benítez H, Sanzana M, Jerez V, Parra L, Hernández C, Canales-Aguirre C. 2013. Sexual Shape and Size Dimorphism in Carabid Beetles of the Genus *Ceroglossus*: Is Geometric Body Size Similar Between Sexes Due to Sex Ratio? *Zoological Science* 30(4): 289–295.

Betts C, Wootton R. 1998. Wing Shape and Flight Behaviour in Butterflies (Lepidoptera: Papilionoidea and Hesperioidea): A Preliminary Analysis. *Journal of Experimental Biology* 138: 271–288.

Cepeda-Pizarro J, Vásquez H, Veas H, Colon G. 1996. Relaciones entre tamaño corporal y biomasa en adultos de Tenebrionidae (Coleoptera) de la estepa costera del margen meridional del desierto chileno. *Revista Chilena de Historia Natural* 69: 67–76.

Debat V, Begin M, Legout H, David J. 2003. Allometric and nonallometric components of *Drosophila* wing shape respond differently to developmental temperature. *Evolution* 57: 2773–2784.

Drake A, Klingenberg C. 2008. The pace of morphological change: historical transformation of skull shape in St Bernard dogs. *Proceedings of the Royal Society B-Biological Sciences* 275: 71–76.

Dryden I, Mardia K. 1998. *Statistical Shape Analysis*. Wiley.

Dudley R. 2000. *The Biomechanics of Insect Flight: Form, Function, Evolution*. Princeton University Press.

Esperk T, Tammaru T, Nylin S, Teder T. 2007. Achieving high sexual size dimorphism in insects: females add instars. *Ecological Entomology* 32: 243–256.

Fairbairn D, Blanckenhorn W, Szekely T. 2007. *Sex, size and gender roles: Evolutionary studies of sexual size dimorphism*. Oxford University Press.

Fairbairn D. 1997. Allometry for sexual size dimorphism: Pattern and process in the coevolution of body size in males and females. *Annual Review of Ecology and Systematics* 28: 659–687.

Fairbairn D, Preziozi R. 1994. Sexual selection and the evolution of allometry for sexual size dimorphism in the water strider, *Aquarius remigis. American Naturalist* 144: 101–118.

Forrest T. 1987. Insect size tactics and developmental strategies. *Oecologia* 73: 178–184.

Gauld D. 1978. Notes on the British Ophioriini (Hym., Ichneumonidae). A revised key to species of genus *Ophion* Fabricius. *Entomologist's Gazette* 29: 145– 150.

Gauld D, Bolton B. 1988. *The Hymenoptera*. British Museum (Natural History) and Oxford University Press.

Gidaszewski N, Baylac M, Klingenberg C. 2009. Evolution of sexual dimorphism of wing shape in the *Drosophila melanogaster* subgroup. *BMC Evolutionary Biology* 9: 110.

Hurlbutt B. 1987. Sexual size dimorphism in parasitoid wasps. *Biological Journal of the Linnean Society* 30: 63–89.

Klingenberg C. 1996. Multivariate allometry. In: Marcus LF, Corti M, Loy A, Naylor G, Slice D, Editors. *Advances in morphometrics*. pp. 23–49. Plenum Press.

Klingenberg C. 2011. MorphoJ: an integrated software package for geometric morphometrics. *Molecular Ecology Resources* 11: 353–357.

Klingenberg C, Barluenga M, Meyer A. 2002. Shape analysis of symmetric structures: Quantifying variation among individuals and asymmetry. *Evolution* 56: 1909–1920.

Klingenberg C, Duttke S, Whelan S, Kim M. 2012. Developmental plasticity, morphological variation and evolvability: a multilevel analysis of morphometric integration in the shape of compound leaves. *Journal of Evolutionary Biology* 25: 115–129.

Klingenberg C, McIntyre G. 1998. Geometric morphometrics of developmental instability: Analyzing patterns of fluctuating asymmetry with procrustes methods. *Evolution* 52: 1363–1375.

Mackauer M. 1996. Sexual size dimorphism in solitary parasitoid wasp: influence of host quality. *Oikos* 76: 265–272.

Mitrovski-Bogdanović A, Ivanovic A, Tomanovic Z, Zikic V, Stary P, Kavallieratos NG. 2009. Sexual dimorphism in *Ephedrus persicae* (Hymenoptera: Braconidae: Aphidiinae): intraspecific variation in size and shape. *Canadian Entomologist* 141: 550–560.

Moller A, Zamora-Muñoz C. 1997. Antennal asymmetry and sexual selection in a cerambycid beetle. *Animal Behaviour* 54: 1509–1515.

Moutia LA, Courtois CM. 1952. Parasites of the moth borers of sugarcane in Mauritius. *Bulletin of Entomological Research* 43: 325–359.

Nylin S, Gotthard K. 1998. Plasticity in life history traits. *Annual Review of Entomology* 43: 63–83.

O'Neill K. 2001. *Solitary wasps*. Cornell University Press.

Pretorius E. 2005. Using geometric morphometrics to investigate wing dimorphism in males and females of Hymenoptera - a case study based on the genus *Tachysphex* Kohl (Hymenoptera:

Sphecidae: Larrinae). *Australian Journal of Entomology* 44: 113–121.

Price PW. 1975. The Evolutionary Strategies of Parasitic Insects and Mites. Plenum Press.

Reeve J, Fairbairn D. 1999. Change in sexual size dimorphism as a correlated response to selection on fecundity. *Heredity* 83: 697–706.

Rohlf F. 2008. *TPSUtil*, version 1.57. New York State University.

Rohlf F. 2008b. *TPSdig*, version 2.12. New York State University.

Rohlf F, Slice D. 1990. Extensions of the Procrustes method for the optimal superimposition of landmarks. *Systematic Zoology* 39: 40–59.

Shreeves G, Field J. 2008. Parental care and sexual size dimorphism in wasps and bees. *Behavioral Ecology and Sociobiology* 62: 843–852.

Townes H. 1971. The genera of Ichnelimotiidae, part 4. *Memoirs of the American Entomological Institute* 17: 1–372.

Vickery RA. 1929. Studies on the fall armyworm in the Gulf Coast district of Texas. *Technical Bulletin of the United States Department of Agriculture* 138: 1–63. Walker S, Rypstra A. 2001. Sexual dimorphism in functional response and trophic morphology in *Rabidosa rabida* (Aranae: Lycosidae). *The American Midland Naturalist* 146: 161–170.

Yu DS, Horstmann K. 1997. A catalogue of World Ichneumonidae, part 1: Subfamilies Acaenitinae to Ophioninae. *Memoirs of the American Entomological Institute* 58(1): 1– 763.

Zelditch M, Swiderski D, Sheets D. 2012. Geometric Morphometrics for Biologists: A Primer. Elsevier.

Zikic V, Tomanovic Z, Ivanovic A, Kavallieratos NG, Stary P, Stanisavljevic LZ, Rakhshani E. 2009. Morphological Characterization of *Ephedrus persicae* Biotypes (Hymenoptera: Braconidae: Aphidiinae) in the Palaearctic. *Annals of the Entomological Society of America* 102: 1– 11.

Zikic V, Tomanovic Z, Kavallieratos NG, Stary P, Ivanovic A. 2010. Does allometry account for shape variability in *Ephedrus persicae* Froggatt (Hymenoptera: Braconidae: Aphidiinae) parasitic wasps? *Organisms Diversity and Evolution* 10: 373– 380.









