

Commensal Aerobic Bacterial Flora of the Gastrointestinal Tract of *Pipistrellus nathusii* (Chiroptera: Vespertilionidae): Lack of *Escherichia coli* in Fecal Samples

Author: Jarzembowski, Tomasz

Source: Acta Chiropterologica, 4(1) : 99-106

Published By: Museum and Institute of Zoology, Polish Academy of Sciences

URL: <https://doi.org/10.3161/001.004.0112>

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at www.bioone.org/terms-of-use.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

**Commensal aerobic bacterial flora of the gastrointestinal tract of
Pipistrellus nathusii (Chiroptera: Vespertilionidae): lack of *Escherichia coli*
in fecal samples**

TOMASZ JARZEMBOWSKI

Department of Medical Microbiology, Medical University of Gdańsk, ul. Do Studzienki 38,
80-227 Gdańsk, Poland; E-mail: doktj@univ.gda.pl

Key words: *Pipistrellus nathusii*, oral cavity, feces, commensal flora, *Escherichia coli*, Poland

INTRODUCTION

The relationships between the macro-organism and commensal bacterial flora are influenced by many factors, including the host specificity and food composition (Jabłoński, 1979). Nevertheless, even basic knowledge about bacterial microflora of wild mammals is insufficient (Akatov, 1984; see also Souza *et al.*, 1999), and bats are not an exception.

These animals seem to be especially interesting for the study of macro- and micro-organism relationships and host specificity of bacteria due to many unusual features, e.g., hypothermia during winter. The recent study of Różalska *et al.* (1998) showed lack of one of the most common fecal bacteria, *Escherichia coli*, in the gastrointestinal track of *Myotis myotis* and *Barbastella barbastellus*. The aim of the present work was to gain information on the composition of the bacterial microflora of *Pipistrellus nathusii*.

MATERIALS AND METHODS

Samples were taken from the oral cavity and feces of *P. nathusii* found in day roosts on Vistula Split (northern Poland) in the 2nd half of October 2000 by using sterile swabs moistened with 0.9% NaCl before sampling. They were inoculated on solid media (blood agar, MacConkey agar, Chapman agar, Sabouraud agar), and incubated 24 hrs at 37°C and then 24 hrs at room temperature. After incubation, micrococci and staphylococci were distinguished according to the method described by Schumacher

(1984). Staphylococcal cultures were identified to species level by using the ID 32 staph kit (Bio-Merieux). The gram-negative bacteria were identified by using ID 32 GN kit (BioMerieux). Antibiotic susceptibility was investigated by the disk-diffusion method on Mueller-Hinton agar. A total of 93 bacterial strains were recovered from 25 oral and 26 fecal samples processed in the Department of Medical Microbiology, Medical University of Gdańsk, Poland.

RESULTS

Most (40%) of all bacterial strains found in fecal samples ($n = 57$) belonged to the family Enterobacteriaceae (*Enterobacter cloacae*, *E. agglomerans*, *Serratia liquefaciens*, *S. fonticola*, *Hafnia alvei*, *Klebsiella pneumoniae*, and *Citrobacter freundii*); 35% of the strains were coagulase-negative staphylococci (*Staphylococcus warneri*, *S. xylosus*, *S. sciuri*, *S. lentus*, and *S. equorum*) and the remaining strains (25%) belonged to the genera *Micrococcus*, *Enterococcus*, and non-fermenting rods (Fig. 1).

Of the strains isolated from oral cavity ($n = 36$), 43% were staphylococci (*S. equorum*, *S. warneri*), 27% micrococci, 18% enterococci, and the remaining strains (12%) belonged to the genus *Bacillus* and the family Enterobacteriaceae (*E. cloacae*, *E. agglomerans*; Fig. 1). There was no dominant species among rods of the family Enterobacteriaceae. However, the staphylococcal flora was dominated by *S. equorum* (15% of all isolates versus up to 3.2% isolates for other species; Table 1).

TABLE 1. The percentage of isolates of different species of bacteria (93 strains) from *P. nathusii*

Gram-negative	Isolates (%)	Gram-positive	Isolates (%)
Enterobacteriaceae		<i>Bacillus</i> sp.	1.1
<i>Serratia liquefaciens</i>	6.5	Cocci	
<i>Hafnia alvei</i>	4.3	<i>Staphylococcus equorum</i>	15.1
<i>Enterobacter cloacae</i>	4.3	<i>S. lentus</i>	3.2
<i>E. agglomerans</i>	3.2	<i>S. xylosus</i>	3.2
<i>Serratia fonticola</i>	2.1	<i>S. warneri</i>	2.1
<i>Citrobacter freundii</i>	1.1	<i>S. sciuri</i>	1.1
<i>Klebsiella pneumoniae</i>	1.1	<i>Staphylococcus</i> (k-) sp.	9.7
Unidentified	3.2	<i>Enterococcus</i> sp.	10.8
Non-fermenting rods		<i>Micrococcus</i> sp.	18.3
<i>Pseudomonas fluorescens</i>	2.1	<i>M. luteus</i>	2.1
<i>P. putida</i>	2.1	<i>M. roseus</i>	1.1
Unidentified	2.1		

As to the frequency of occurrence of different groups of bacteria, the most common isolates in feces of *P. nathusii* ($n = 26$) were gram-negative rods of the family Enterobacteriaceae (62%), and gram-positive cocci of the genera *Staphylococcus* (k-) (58%) and *Enterococcus* (12%) (Fig. 2). In oral cavity samples the gram-positive cocci (*Staphylococcus*, 42%, and *Micrococcus*, 30%) dominated. Enterococci were found in 18% of oral samples (Fig. 2).

DISCUSSION

The results of the study show several differences between aerobic gastrointestinal

flora of bats from the genus *Pipistrellus* and other species of mammals. The most surprising is the lack of *Escherichia coli* among bacterial flora. This bacterium is believed to be one of the most common in the intestinal track of vertebrates. Furthermore, *E. coli* plays an important role in the synthesis of some vitamins (B, K, C) and utilization of food (Jabłoński, 1979). The presence of *E. coli* can also prevent of colonization of gastrointestinal tract by pathogenic species, like *Staphylococcus aureus* and *Salmonella typhimurium* (Ushijima and Ozaki 1986, 1988). As there is no *E. coli* found in feces of *M. myotis* and *B. barbastellus* (Różalska *et al.*, 1998), the lack of

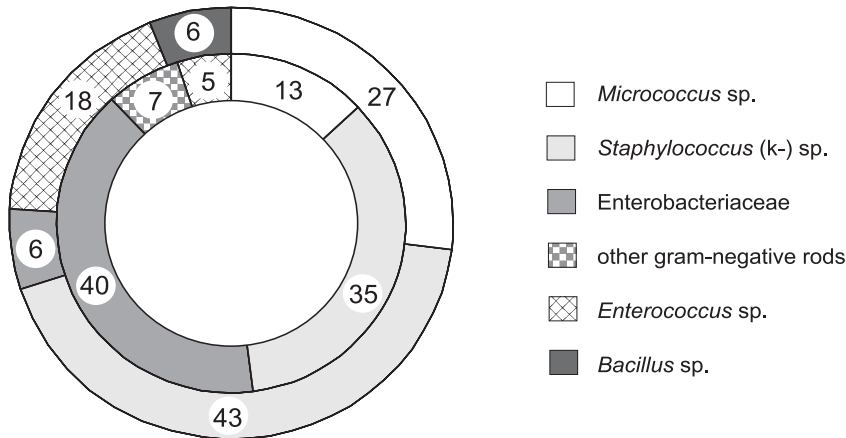


FIG. 1. Proportions of commensal microflora of *P. nathusii* (internal circle — strains isolated from fecal samples, $n = 57$; external circle — from oral samples, $n = 36$)

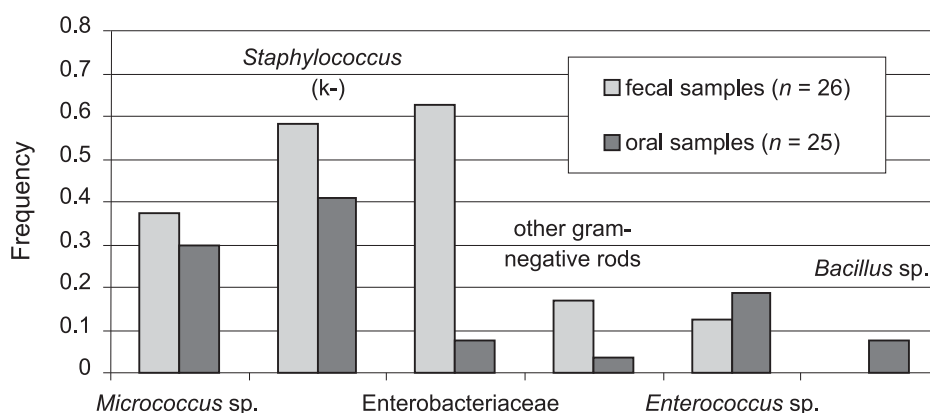


FIG. 2. Frequency of occurrence of different groups of bacteria in samples from *P. nathusii*

this bacterium differs vesperilionid bats from other vertebrates (including pteropids — Heard *et al.*, 1997), and the means of regulation of commensal flora in the intestinal tract of these bats remains unknown.

In contrast to Różalska *et al.* (1998), we have not isolated *Bacillus* strains from feces originating from *P. nathusii*. Nevertheless, the staphylococcal flora of *P. nathusii* seems to be more diverse than that of wild birds where only two species of coagulase-negative staphylococci were discovered (Akato *et al.*, 1984).

The frequency of recovery of gram-negative rods from feces of *P. nathusii* was similar to that observed in *M. myotis*, but it was higher than that in *B. barbastellus* (Różalska *et al.*, 1998). In contrast to *M. myotis* and *B. barbastellus*, non-fermenting gram-negative rods have been not isolated from the oral cavity samples of *P. nathusii*. On the other hand, the gram-negative rods of the family Enterobacteriaceae were less frequent in *P. nathusii* than in the other bat taxa, whereas enterococci were more frequent.

The observed interspecific differences in the specific composition of bacteria may result from slight differences in bats' diets, although *Myotis*, *Barbastella*, and *Pipistrellus* are insectivorous. In addition,

differences in bacterial flora composition can be observed before and after hibernation (Różalska *et al.*, 1998). Unfortunately, because there are no winter roosts of *P. nathusii* in the study area, it is impossible to make this comparison.

ACKNOWLEDGEMENTS

I thank Prof. J. Galiński and Prof. W. Bogdanowicz, and two anonymous reviewers for their helpful comments on an earlier version of this manuscript.

LITERATURE CITED

- DZIERŻANOWSKA, D., and K. MADALIŃSKI. 1999. Flora fizjologiczna. Pp. 17–19, in *Zakażenia szpitalne* (D. DZIERŻANOWSKA and J. JELASZEWICZ, eds.). Medica Press, Bielsko Biala, 255 pp.
- HEARD, D. J., J. L. DE YOUNG, B. GOODYEAR, and G. A. ELLIS. 1997. Comparative rectal bacterial flora of four species of flying fox (*Pteropus* sp.). *Journal of Zoo and Wildlife Medicine*, 28: 471–475.
- JABŁOŃSKI, L. 1979. *Podstawy mikrobiologii lekarskiej*. PWN, Warszawa, 520 pp.
- RÓŻAŁSKA, B., G. RADZICKI, B. SADOWSKA, J. MARKOWSKI, and W. RUDNICKA. 1998. Aerobic microflora of *Myotis myotis* (Borkhausen, 1797) and *Barbastella barbastellus* (Schreber, 1774). *Bulletin of the Polish Academy of Science, Biological Sciences*, 46: 21–26.
- SOUZA, V., M. ROCHA, A. VALERA, and L. E. EGUIARTE. 1999. Genetic structure of natural populations of *Escherichia coli* in wild hosts on different continents. *Applied and Environmental Microbiology*, 65: 3373–3385.

- USHIJIMA, T., and Y. OZAKI. 1986. Potent antagonism of *Escherichia coli*, *Bacterioides ovatus*, *Fusobacterium varium* and *Enterococcus faecalis*, alone or combination, for enteropathogens in anaerobic continuous cultures. *Journal of Medical Microbiology*, 22: 157–163.
- USHIJIMA, T., and Y. OZAKI. 1988. Factors influencing potent antagonistic effect of *Escherichia coli* and *Bacterioides ovatus* on *Staphylococcus aureus* in anaerobic continuous flow cultures. *Canadian Journal of Microbiology*, 34: 645–650.
- VIRELLA, G., and G. SCHMIDT. 2000. Enterobacteriaceae i inne pałeczki gram-ujemne. Pp. 161–177, in *Mikrobiologia i choroby zakaźne* (G. VIRELLA, ed.). Urban and Partner, Wrocław, 644 pp.

Received 5 October 2001, accepted 28 January 2002