

Chloroplast Microsatellite Markers for Pseudotaxus chienii Developed from the Whole Chloroplast Genome of Taxus chinensis var. mairei (Taxaceae)

Authors: Deng, Qi, Zhang, Hanrui, He, Yipeng, Wang, Ting, and Su, Yingjuan

Source: Applications in Plant Sciences, 5(3)

Published By: Botanical Society of America

URL: https://doi.org/10.3732/apps.1600153

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at <u>www.bioone.org/terms-of-use</u>.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.



PRIMER NOTE

CHLOROPLAST MICROSATELLITE MARKERS FOR *PSEUDOTAXUS* CHIENII DEVELOPED FROM THE WHOLE CHLOROPLAST GENOME OF *TAXUS CHINENSIS* VAR. *MAIREI* (TAXACEAE)¹

QI DENG^{2,3}, HANRUI ZHANG², YIPENG HE², TING WANG^{4,5}, AND YINGJUAN SU^{2,5}

²School of Life Sciences, Sun Yat-sen University, Guangzhou 510275, People's Republic of China; ³School of Medicine, Guangxi University of Science and Technology, Liuzhou 545005, People's Republic of China; and ⁴College of Life Science, South China Agricultural University, Guangzhou 510642, People's Republic of China

- *Premise of the study: Pseudotaxus chienii* (Taxaceae) is an old rare species endemic to China that has adapted well to ecological heterogeneity with high genetic diversity in its nuclear genome. However, the genetic variation in its chloroplast genome is unknown.
- *Methods and Results:* Eighteen chloroplast microsatellite markers (cpSSRs) were developed from the whole chloroplast genome of *Taxus chinensis* var. *mairei* and successfully amplified in four *P. chienii* populations and one *T. chinensis* var. *mairei* population. Of these loci, 10 were polymorphic in *P. chienii*, whereas six were polymorphic in *T. chinensis* var. *mairei*. The unbiased haploid diversity per locus ranged from 0.000 to 0.641 and 0.000 to 0.545 for *P. chienii* and *T. chinensis* var. *mairei*, respectively.
- *Conclusions:* The 18 cpSSRs will be used to further investigate the chloroplast genetic structure and adaptive evolution in *P. chienii* populations.

Key words: chloroplast microsatellite; genetic diversity; Pseudotaxus chienii; Taxaceae; Taxus chinensis var. mairei.

Taxus L. and Pseudotaxus W. C. Cheng are two closely related sister genera with similar appearance in Taxaceae (Fu et al., 1999). Their only distinction is the difference in color in the stomatal bands and aril (Fu et al., 1999). Both T. chinensis (Pilg.) Rehder var. mairei (Lemée & H. Lév.) W. C. Cheng & L. K. Fu and P. chienii (W. C. Cheng) W. C. Cheng are coniferous species endemic to China. Taxus chinensis var. mairei, in particular, has a high medicinal value because it contains the anticancer agent taxol (Li et al., 2008). Pseudotaxus chienii, the sole species in the monotypic genus, is an evergreen shrub or small tree with an average height of 4 m (Su et al., 2009). Due to overexploitation and human activities, the population size of P. chienii is shrinking. The species is categorized as an endangered species in the Red List of Endangered Plants in China (Fu and Jin, 1992). As an "old rare species," P. chienii has adapted well to habitat fragmentation and ecological heterogeneity across a wide range of habitats and is found in Zhejiang, Jiangxi, Hunan,

¹Manuscript received 8 December 2016; revision accepted 7 February 2017.

The authors thank L. Huang, L. M. Cui, and N. Li of the School of Life Sciences, Sun Yat-sen University, for assistance with the experiment. This work was supported by the National Natural Science Foundation of China (grant no. 31070594, 31370364, 31570652, and 31670200), the National Natural Science Foundation of Guangdong Province, China (grant no. 2016A030313320), and the Project of the Department of Science and Technology of Shenzhen City, China (no. JCYJ20160425165447211).

⁵Authors for correspondence: suyj@mail.sysu.edu.cn; tingwang@scau. edu.cn

doi:10.3732/apps.1600153

and Guangxi provinces (Deng et al., 2013). The previous nuclear inter-simple sequence repeat (ISSR) and simple sequence repeat (SSR) markers have revealed that *P. chienii* possesses high genetic diversity, which provides a large pool of raw material for adaptive evolution (Su et al., 2009; Deng et al., 2013). However, the level of genetic variation in the *P. chienii* chloroplast genome is unknown.

Chloroplast simple sequence repeat (cpSSR) markers, which have been extensively used in population genetics, possess important and unique characteristics such as haploidy, nonrecombination, uniparental inheritance, and a low nucleotide substitution rate (Ebert and Peakall, 2009). cpSSR loci are generally distributed throughout the noncoding regions with higher sequence variations and have conservative flanking regions (Huang et al., 2015). In particular, the chloroplast genome retains ancient genetic patterns and can therefore provide unique insight into evolutionary processes (Provan et al., 2001). Therefore, cpSSR markers can be used to investigate genetic variation in small, fragmented populations and can be transferred to related species (Schaal et al., 1998; Petit et al., 2005; Pan et al., 2014). More important, because cpSSRs are paternally inherited in gymnosperms, they can be used to assess pollen-mediated gene flow, population genetic variation, and phylogeographic patterns. Information revealed by cpSSRs is complementary to that obtained from nuclear SSRs (Powell et al., 1996; Provan et al., 2001). Although no chloroplast genome sequences of P. chienii have been reported, the complete chloroplast genome sequence of *T. chinensis* var. *mairei* is available in the National Center for Biotechnology Information's GenBank (accession no. NC_020321.1). Thus, here we first isolated 18 cpSSRs in

Applications in Plant Sciences 2017 5(3): 1600153; http://www.bioone.org/loi/apps © 2017 Deng et al. Published by the Botanical Society of America. This is an open access article distributed under the terms of the Creative Commons Attribution License (CC-BY-NC-SA 4.0), which permits unrestricted noncommercial use and redistribution provided that the original author and source are credited and the new work is distributed under the same license as the original.

TABLE 1.	Characteristics of 18 cpS	SR markers developed for	or Taxus chinensis var. mairei.

Locus		Primer sequences $(5'-3')$	Repeat motif	$T_{\rm a}(^{\circ}{\rm C})$	Product size (bp)	Location ^c
PTC-cp01 ^{a,b}	F:	CACCATCCACTGCCTTTG	$(AT)_9$	54	168	trnH-GUG-trnI-CAU (337-354)
•	R:	GTGCGGTCAGAACTTGTCA				
PTC-cp02 ^b	F:	TGCGTGGCTGTGAGATG	(AC) ₅	54	213	trnT-trnE (21,944-21,953)
	R:	GCGGAACCCGTAGTGAA				
PTC-cp03	F:	AAGCCGCCCTGTTTTA	$(TA)_5(AT)_5$	54	232	trnC-GCA-rpoB (25,696-25,715)
	R:	ATCTCATCGCATTGGAAGT				
PTC-cp04 ^b	F:		$(AT)_5$	54	135	rpoC2 (32,846–32,855)
	R:	GCGTGGCAATACATCTCC				
PTC-cp05	F:	TGAACAGGTCCGACAGCA	$(A)_{11}TACAA(AT)_5$	55	376	atpF-atpA (39,353–39,378)
	R:	CCATCCCATCTCCTACTTGA				
PTC-cp08	F:		(AT) ₅ AGAAGTATACTTC(TA) ₅	54	173	rpl36-rps11 (57,373–57,405)
	R:	CGGGGAACTAATCTTCTTGT				
PTC-cp09 ^a	F:	TGTATCAACCAATGCTTCC	$(TC)_6$	54	273	<i>psbT-psbB</i> (62,807 – 62,818)
		ATTCATAGATGTTTTCGCTG				
PTC-cp13 ^b		AGTCCAAAATCTCCCACAC	(AT) ₅	54	180	ndhF (104,893–104,902)
	R:					
PTC-cp15		GCTTGGACCCATTGTTGAA	$(A)_{10}$	55	279	rpoC1-rpoC2 (32,043-32,052)
DEC 16th	R:	CATACTTTAGGTGGCGTTGTTA			225	
PTC-cp16 ^{a,b}	F:	CCCATACTCCCATTTCATAACTT	$(A)_{10}$	55	237	rpoC2 (34,060-34,069)
DEC 10th	R:	AGCACTTGCCCAGGACTAACT			107	
PTC-cp18 ^{a,b}	F:	TCCAGGTGCTGATGCTACTAA	$(A)_{10}$	55	186	rpoC2-atpI (35,652-35,661)
DEC Ath	R:				201	
PTC-cp21 ^b		GGTGGGGTGGGAACG	$(A)_{10}$	55	306	rpl32 (106,169–106,178)
DTC ooth	R:	TTGGGTGAGCCATAGAAAT		~ ~	120	
PTC-cp22 ^{a,b}		AGCAATGTTTGGAAGGGAA	$(A)_{10}$	55	130	rpl32-trnP-GGG (106,257–106,266)
DTC ash		GGTGTAGTCTATTTGGTGGTGTT			201	(2: (0270, 0207)
PTC-cp23 ^b		AACTAATCCCAATGGCTTCA	$(T)_{10}$	55	301	<i>ycf3</i> intron (9378–9387)
DTC 2(sh		CCCTATGCGTGCCTATCA		<i></i>	226	
PTC-cp26 ^{a,b}	F:		$(T)_{10}$	55	326	<i>ycf4</i> (65,483–65,492)
DTC 29		AAACTACGGCGATTTCTTC		55	336	
PTC-cp28	F:	TGTAGTTTGCCGAGTGGTT	(T) ₁₁	55	330	<i>psbE-petL</i> (70,723–70,733)
DTC an 20ah	R:	AATAATAGTAGACATTGGAAGGAC		55	254	$= 2^{\circ} (75, 420, 75, 440)$
PTC-cp29 ^{a,b}		AATAGGTTCTGGAGCGGTTA AGATTTAGTTCGTCACGGGTA	(T) ₁₁	33	234	rps8 (75,439–75,449)
DTC an22ah			(TCTTCC)7	55	284	rps15-chlN (121,797–121,838)
PTC-cp32 ^{a,b}	F:	CCTCGTGCGGATAACTAAA TGGCAAAGATTCCCTGG	$(1C11CC)_7$	55	204	<i>ipsij-cluiv</i> (121, <i>191</i> –121,038)
	K:	IGGUAAAGATTUUUTGG				

Note: T_a = annealing temperature.

^aMonomorphic loci for *Pseudotaxus chienii*.

^b Monomorphic loci for *T. chinensis* var. *mairei*.

^c Locus location (genic or intergenic region); the position amplified by the primers in the *T. chinensis* var. *mairei* chloroplast genome is given in parentheses.

T. chinensis var. *mairei*, then applied 10 polymorphic markers to evaluate the genetic diversity of *P. chienii*. These markers will be further applied to survey the chloroplast genetic variation in *P. chienii*.

METHODS AND RESULTS

In this study, a total of 109 individuals from four populations of *P. chienii* were collected throughout its natural distribution range, including Shuimenjian (Zjsmj) in Zhejiang Province, Zhangjiajie (Hnzjj) in Hunan Province, Zizhuba (Jxzzb) in Jiangxi Province, and Damingshan (Gxdms) in Guangxi Zhuang Autonomous Region, China (Appendix 1). One *T. chinensis* var. *mairei* population was gathered from Fenshui (Jxfs) in Jiangxi Province. Due to its rare and endangered properties, only 11 individuals were sampled. Young leaves were collected and dried in silica gel immediately. Genomic DNA was extracted using a modified cetyltrimethylammonium bromide (CTAB) protocol (Doyle and Doyle, 1987).

From the complete chloroplast genome sequence for *T. chinensis* var. *mairei* (GenBank accession no. NC_020321.1), 32 cpSSR loci were identified with the repeat threshold settings of 10 repeats for mononucleotides and five repeats for di-, tri-, tetra-, penta-, and hexanucleotide cpSSRs. Based on their flanking regions, we designed 27 primers using Primer Premier 5.0 software (PREMIER Biosoft International, Palo Alto, California, USA). One individual (SMJ27) from Zjsmj population for *P. chienii* and one individual (FS7) from Jxfs population for *T. chinensis* var. *mairei* were selected to screen these primers. PCR was

performed in a total volume of 20 µL containing 20 ng of genomic DNA, 1× PCR buffer, 5 mM MgCl₂, 0.2 mM dNTPs mixture, 0.25 µM of each primer, and 1 unit *Taq* polymerase (TaKaRa Biotechnology Co., Dalian, China). Reaction conditions included initial denaturation at 94°C for 3 min; followed by 35 cycles at 94°C for 1 min, annealing temperature for 1 min, and 72°C for 1 min; with a final extension at 72°C for 10 min. The annealing temperature was optimized by gradient PCR (Table 1). Amplified products were separated by 6% denaturing polyacrylamide gel electrophoresis and visualized by silver staining. The allele sizes were estimated with a 50-bp DNA ladder (TaKaRa Biotechnology Co.) as size standard. Eighteen of 27 primers (approximately 67%) could produce clear bands in both *P. chienii* and *T. chinensis* var. *mairei*. The 18 cpSSRs were divided into three categories in terms of motif structure: 15 perfect, one imperfect, and two compound repeats. The high frequency of perfect repeats was in accordance with Ebert's description (Ebert and Peakall, 2009).

The utility of these 18 cpSSR primers was further examined in 109 and 11 individuals of *P. chienii* and *T. chinensis* var. *mairei*, respectively. The PCR reactions were conducted as described above. Among these loci, 10 (PTC-cp02, PTC-cp03, PTC-cp04, PTC-cp05, PTC-cp08, PTC-cp13, PTC-cp15, PTC-cp21, PTC-cp23, and PTC-cp28) showed polymorphisms in *P. chienii*, whereas six (PTC-cp03, PTC-cp05, PTC-cp09, PTC-cp09, PTC-cp15, and PTC-cp28) were polymorphic in *T. chinensis* var. *mairei* (Table 1). The genetic parameters, including the number of alleles (*A*), haploid diversity (*h*), and unbiased haploid diversity (*h*_{unb}) for each population, were evaluated with GenAIEx version 6.41 (Peakall and Smouse, 2006). Ten polymorphic cpSSR loci for *P. chienii*, *A* was between one and four, *h* ranged from 0.000 to 0.620, and *h*_{unb} varied from 0.000 to 0.641 (Table 2). Population Zjsmj revealed obviously higher diversity than

	Pseudotaxus chienii									Taxus chinensis var. mairei					
	Z	Zjsmj ($n = 3$	30)	G	xdms ($n =$	30)	1	$\operatorname{Inzjj}(n = 1)$.9)	J	xzzb ($n = 3$	30)		Jxfs ($n = 1$	1)
Locus	Α	h	h _{unb}	Α	h	$h_{\rm unb}$	Α	h	$h_{\rm unb}$	Α	h	$h_{\rm unb}$	Α	h	h_{unb}
PTC-cp02	1	0.000	0.000	1	0.000	0.000	1	0.000	0.000	2	0.320	0.331	NAc	NAc	NAc
PTC-cp03	2	0.180	0.186	1	0.000	0.000	2	0.188	0.199	1	0.000	0.000	2	0.165	0.182
PTC-cp04	2	0.180	0.186	1	0.000	0.000	2	0.100	0.105	2	0.064	0.067	NAc	NAc	NA ^c
PTC-cp05	3	0.127	0.131	3	0.127	0.131	2	0.100	0.105	1	0.000	0.000	3	0.430	0.473
PTC-cp08	2	0.498	0.515	3	0.504	0.522	1	0.000	0.000	1	0.000	0.000	2	0.463	0.509
PTC-cp09	NA ^b	NA ^b	NA ^b	NA ^b	NA ^b	NA ^b	NA ^b	NA ^b	NA ^b	NA ^b	NA ^b	NA ^b	2	0.463	0.509
PTC-cp13	3	0.504	0.522	1	0.000	0.000	1	0.000	0.000	1	0.000	0.000	NAc	NAc	NAc
PTC-cp15	3	0.418	0.432	1	0.000	0.000	1	0.000	0.000	1	0.000	0.000	2	0.496	0.545
PTC-cp21	1	0.000	0.000	2	0.124	0.129	1	0.000	0.000	1	0.000	0.000	NAc	NAc	NA ^c
PTC-cp23	2	0.124	0.129	2	0.064	0.067	2	0.100	0.105	2	0.124	0.129	NAc	NAc	NA ^c
PTC-cp28	4	0.620	0.641	3	0.184	0.191	3	0.421	0.444	2	0.320	0.331	2	0.463	0.509

TABLE 2.	Genetic properties of 10 poly	morphic cpSSR loci for Pseudotaxus	chienii and six polymorphic loci	for Taxus chinensis var. mairei. ^a
----------	-------------------------------	------------------------------------	----------------------------------	---

Note: A = number of alleles; h = haploid diversity; h_{unb} = unbiased haploid diversity; n = number of individuals sampled.

^aVoucher and locality information are provided in Appendix 1.

^bNo analysis performed because PTC-cp09 was monomorphic in *P. chienii*.

°No analysis performed because PTC-cp02, PTC-cp04, PTC-cp13, PTC-cp21, and PTC-cp23 were monomorphic in T. chinensis var. mairei.

other populations. For *T. chinensis* var. *mairei*, *A*, *h*, and h_{unb} were one to three, 0.000–0.496, and 0.000–0.545, respectively (Table 2).

Analysis of molecular variance (AMOVA) was performed to measure genetic differentiation and the ratio of genetic variations within and among *P. chienii* populations in Arlequin version 3.5 (Excoffier and Lischer, 2010). The results revealed significant difference in partitioning of variation among and within populations (29.03% and 70.97%, respectively; Table 3) and uncovered significant genetic differentiation among all populations ($F_{\rm ST}$ = 0.2903).

CONCLUSIONS

The polymorphic chloroplast SSR loci developed from *T. chinensis* var. *mairei* in this study were verified to be reliable for assessing genetic variation of *P. chienii* populations. Combined with the nuclear SSR loci previously developed (Deng et al., 2013), the 18 cpSSRs will contribute to further exploration of whether the adaptation of *P. chienii* to environmental heterogeneity is driven through nuclear or chloroplast loci. In addition, the conservative nature of cpDNA may allow these markers to be used in other conifers.

LITERATURE CITED

- DENG, Q., Y. J. SU, AND T. WANG. 2013. Microsatellite loci for an old rare species, *Pseudotaxus chienii*, and transferability in *Taxus wallichiana* var. *mairei* (Taxaceae). *Applications in Plant Sciences* 1: 1200456.
- DOYLE, J. J., AND J. L. DOYLE. 1987. A rapid DNA isolation procedure for small quantities of fresh leaf tissue. *Phytochemical Bulletin* 19: 11–15.

TABLE 3. The analysis of molecular variance (AMOVA) within and among populations based on 10 polymorphic cpSSRs in *Pseudotaxus chienii*.

Source of variation	df	Sum of squares	Variance components	Percentage of variation	P value
Among populations	3	22.880	0.25926	29.03%	<0.0001
Within populations	105	66.542	0.63373	70.97%	< 0.0001
Total	108	89.422	0.893	100.00%	

Note: df = degrees of freedom.

EBERT, D., AND R. PEAKALL. 2009. Chloroplast simple sequence repeats (cpSSRs): Technical resources and recommendations for expanding cpSSR discovery and applications to a wide array of plant species. *Molecular Ecology Resources* 9: 673–690.

- EXCOFFIER, L., AND H. E. L. LISCHER. 2010. Arlequin suite ver 3.5: A new series of programs to perform population genetics analyses under Linux and Windows. *Molecular Ecology Resources* 10: 564–567.
- FU, L. G., AND J. M. JIN. 1992. Red list of endangered plants in China. Science Press, Beijing, China.
- FU, L. G., N. LI, AND R. R. MILL. 1999. Taxaceae. In Z. Y. Wu and P. H. Raven [eds.], Flora of China, vol. 4, 89–98. Science Press, Beijing, China, and Missouri Botanical Garden Press, St. Louis, Missouri, USA.
- HUANG, J., X. T. YANG, C. M. ZHANG, X. YIN, S. P. LIU, AND X. G. LI. 2015. Development of chloroplast microsatellite markers and analysis of chloroplast diversity in Chinese jujube (*Ziziphus jujuba* Mill.) and wild jujube (*Ziziphus acidojujuba* Mill.). *PLoS ONE* 10: e0134519.
- LI, C. F., C. H. Huo, M. L. ZHANG, AND Q. W. SHI. 2008. Chemistry of Chinese yew, Taxus chinensis var. mairei. Biochemical Systematics and Ecology 36: 266–282.
- PAN, L., Y. LI, R. GUO, H. WU, Z. H. HU, AND C. Y. CHEN. 2014. Development of 12 chloroplast microsatellite markers in *Vigna unguiculata* (Fabaceae) and amplification in *Phaseolus vulgaris*. Applications in *Plant Sciences* 2: 1300075.
- PEAKALL, R., AND P. E. SMOUSE. 2006. GenAlEx 6: Genetic analysis in Excel. Population genetic software for teaching and research. *Molecular Ecology Notes* 6: 288–295.
- PETIT, R. J., J. DUMINIL, S. FINESCHI, A. HAMPE, D. SALVINI, AND G. G. VENDRAMIN. 2005. Comparative organization of chloroplast, mitochondrial and nuclear diversity in plant populations. *Molecular Ecology* 14: 689–701.
- POWELL, W., G. C. MACHRAY, AND J. PROVAN. 1996. Polymorphism revealed by simple sequence repeats. *Trends in Plant Science* 1: 215–222.
- PROVAN, J., W. POWELL, AND P. M. HOLLINGSWORTH. 2001. Chloroplast microsatellites: New tools for studies in plant ecology and evolution. *Trends in Ecology & Evolution* 16: 142–147.
- SCHAAL, B. A., D. A. HAYWORTH, K. M. OLSEN, J. T. RAUSCHER, AND W. A. SMITH. 1998. Phylogeographic studies in plants: Problems and prospects. *Molecular Ecology* 7: 465–474.
- SU, Y. J., T. WANG, AND P. Y. OUYANG. 2009. High genetic differentiation and variation as revealed by ISSR marker in *Pseudotaxus chienii* (Taxaceae), an old rare conifer endemic to China. *Biochemical Systematics and Ecology* 37: 579–588.

Appendix 1.	Collection locality and	voucher information for Pseu	dotaxus chienii and Ta	axus chinensis var. mair	ei populations used in this study.

Species	Population code	Collection locality	Geographic coordinates	Voucher specimen ^a
Pseudotaxus chienii (W. C. Cheng) W. C. Cheng	Zjsmj	Shuimenjian, Zhejiang Province	28°43′42″N, 118°57′32″E	YJ Su 201303, SMJ27
Pseudotaxus chienii	Jxzzb	Zizhuba, Jiangxi Province	26°27′18″N, 114°06′22″E	YJ Su 201303, ZZB12
Pseudotaxus chienii	Hnzjj	Zhangjiajie, Hunan Province	29°23'12"N, 110°28'56"E	YJ Su 201303, ZJJ09
Pseudotaxus chienii	Gxdms	Damingshan, Guangxi Zhuang Autonomous Region	23°29′54″N, 108°26′12″E	YJ Su 201303, DMS17
<i>Taxus chinensis</i> (Pilg.) Rehder var. <i>mairei</i> (Lemée & H. Lév.) W. C. Cheng & L. K. Fu	Jxfs	Fenshui, Jiangxi Province	28°56′31″N, 108°02′12″E	WB Liao 201108, FS1

^aVoucher specimens are deposited at the herbarium of Sun Yat-sen University (SYSU).

http://www.bioone.org/loi/apps