

# Sarcocystis leporum IN COTTONTAIL RABBITS AND ITS TRANSMISSION TO CARNIVORES

Authors: FAYER, R., and KRADEL, D.

Source: Journal of Wildlife Diseases, 13(2): 170-173

Published By: Wildlife Disease Association

URL: https://doi.org/10.7589/0090-3558-13.2.170

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at <a href="https://www.bioone.org/terms-of-use">www.bioone.org/terms-of-use</a>.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

# Sarcocystis leporum IN COTTONTAIL RABBITS AND ITS TRANSMISSION TO CARNIVORES

R. FAYER, United States Department of Agriculture, Animal Parasitology Institute, Agricultural Research Service, Beltsville, Maryland 20705, USA

D. KRADEL, Animal Diseases Laboratory, Department of Veterinary Science, The Pennsylvania State University, University Park, Pennsylvania 16802, USA

Abstract: Muscle from Sarcocystis-infected cottontail rabbits (Sylvilagus floridanus) was fed to coccidia-free cats (Felis domestica) and dogs (Canis familiaris). Only cats became infected and shed sporocysts in their feces. The prepatent period ranged from 10 to 25 days and the patent period from 3 to 46 days. Sporocysts were fully sporulated when shed. They contained 4 sporozoites and a coarse granular residuum and averaged 9.4 by 13.6  $\mu$ m (N=55). Doses of 200-75,000 sporocysts were orally administered to 5 domestic rabbits (Oryctolagus cuniculus). Domestic rabbits did not become infected, suggesting a strict host specificity for the intermediate host S. floridanus.

### INTRODUCTION

Sarcocystis is commonly found in the cardiac and skeletal muscles of many vertebrates. The life cycle recently has been elucidated for some species of Sarcocystis in livestock, wildlife and rodents.3,4,8,9,10 In general, after infected muscle is ingested by a suitable carnivore, zoites are liberated from the intramuscular cysts and enter cells in the carnivore's intestine. Here they develop into gametes and then oocysts. Oocysts sporulate in situ. Fully-developed isosporan sporocysts containing 4 sporozoites are shed in the carnivore's feces. Sporocysts ingested by the appropriate intermediate host initiate an asexual cycle leading to the development of intramuscular cysts.

The life cycle of Sarcocystis in cottontail rabbits (Sylvilagus floridanus) is unknown. The scant literature on Sarcocystis in rabbits, reviewed in great part by Vande Vusse, has dealt with either the morphology of the cyst stage or the prevalence in rabbits based on finding the cyst stage. Although both microscopic and macroscopic cysts have been found in cottontail musculature, Crawley states that cyst size is a function of age and is of no diagnostic value with regard to speciation.

The present study was undertaken to determine (1) the role of carnivores in the life cycle of Sarcocystis leporum Crawley, 1914 of cottontail rabbits and (2) the feasibility of establishing a laboratory model with the domestic rabbit (Oryctolagus cuniculus) fed sporocysts from a cottontail-carnivore cycle.

# MATERIALS AND METHODS

Eighteen cottontail rabbit carcasses acquired at the Veterinary Clinic of the Pennsylvania State University were refrigerated and shipped in insulated containers to the Animal Parasitology Institute (API) at various intervals. Fifteen were used in the following experiments.

Upon arrival at the API, musculature was trimmed from the carcass. Representative pieces of muscle were frozen and sectioned with a cryostat, and other pieces were fixed in 10% neutral buffered formalin and prepared for histologic examination. The remaining muscle

was minced into small pieces and fed to either cats (Felis domestica) or dogs (Canis familiaris), or both, to determine the infectivity for the carnivore host (Table 1).

Eighteen cats and eight dogs were used in four experiments (Table 1). Each was housed in a separate cage to facilitate fecal collection. All cats and dogs were coccidia-free at the beginning of each experiment. All were fed dry pelleted feed except at the time rabbit muscle was fed. Total feces were collected daily from each animal beginning on or before the day of feeding rabbit muscle and ending at the termination of the experiment.

Feces found to contain Sarcocystis sporocysts were refrigerated until many

fecal specimens had accumulated. Sporocysts were then cleaned of fecal debris as previously described, resuspended in tap water, and refrigerated until used as inoculum.

Eight 3 to 6-month-old domestic rabbits were used in two experiments. In one experiment, two rabbits were orally inoculated with either 200 or 1000 sporocysts; two others served as uninoculated controls. All were examined daily for signs of disease until they were killed 34 days later. At this time, a thorough postmortem examination was made, and tissues from nearly all organs were fixed in neutral buffered formalin and prepared for histologic examination. In the second experiment, three rabbits were orally inoculated with 25,000, 50,000, or

TABLE 1. Sarcocystis-infected cottontail rabbit muscle fed to cats and dogs.

Exp.	Carnivore fed cottontail muscle	Carnivore identification No.	Rabbit carcass No.	Sporocysts shed by carnivore	
				Prepatent period (day after c	Patent period arcass fed)
1	Felis domestica	1	A	20	3
	t cos domesica	2	В	15	24
		3-12	None	<del></del>	_
	Canis familiaris	1	Α		
		2	В	_	
		3- 6	None	_	
2	F. domestica	1	C,D,E	_	_
		2	F,G	12	5
		3-12	None	_	-
3	F. domestica	13	н	10	46
		14	I		
		15	J	11	44
		16	K	12	46
		17,18	None	_	
4	F. domestica	14	L,M,N,O	25	15
		17,18	None	_	_
	C. familiarts	7,8	L,M,N,O	_	_

75,000 sporocysts and examined daily until they were killed 86, 50, or 30 days later, respectively; one rabbit served as an uninoculated control and was killed on day 86. Tissues were taken as in the previous experiment.

# RESULTS

Microscopic examination of skeletal muscle from 18 cottontail rabbits submitted to the Veterinary Diagnositc Laboratory revealed that 15 were infected with Sarcocystis. Carcasses A-K contained only microscopic cysts. Carcasses L-O contained both macroscopic and microscopic cysts (Table 1). A typical microscopic cyst is shown in Figure 1.

Three cats fed Sarcocystis-infected muscle from cottontail rabbits became infected and passed Sarcocystis sporocysts in their feces (Exp. 1, two cats; Exp. 4, one cat; Table 1). In contrast, none of four dogs fed parts of the same infected muscle nor any of four unfed dogs and 10 unfed cats passed sporocysts in their feces.

Cats that had passed sporocysts in Experiment 1 were again fed Sarcocystis-infected cottontail rabbit muscle and one became reinfected and again passed sporocysts, (Exp. 2, Table 1). None of the unfed cats passed sporocysts.

Three of four additional cats fed Sarcocystis-infected cottontail rabbit muscle passed sporocysts, but none of the unfed cats passed sporocysts (Exp. 3, Table 1).

Within the 4 experiments, prepatent periods ranged from 10 to 25 days after feeding. Patent periods ranged from 3 to 46 days. Sporocysts most often were seen as individuals in fecal flotations (Fig. 2) although occasional pairs were observed. Sporocysts were ellipsoid. Each contained four sporozoites and a compact granular residuum. No Stiedae body was present. Fifty-five sporocysts averaged 9.4 by 13.6 µm, with a range of 9.3-11.1 µm by 13.0-16.7 µm.

None of the 5 experimentally inoculated rabbits nor the 3 uninoculated controls showed any signs of illness during the 2 experiments. No lesions were observed at postmortem, nor were any parasites found in histological sections.

## DISCUSSION

This is the first report of transmission of *S. leporum* from its intermediate host, the cottontail rabbit, to a definitive host, the domestic cat. In the present study, *F. domestica* was the only felid tested as a possible definitive host. When we consider the recent finding that dogs and

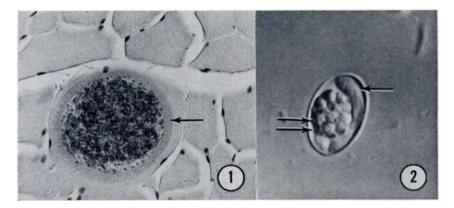


FIGURE 1. Hematoxylin-eosin stained section of cottontail rabbit muscle containing a large cyst.

FIGURE 2. Sporocyst in feces of a cat fed Sarcocystis leporum-infected cottontail rabbit muscle.

coyotes serve as final host for a species of *Sarcocystis* in the bovine and that foxes and raccoons also may serve as final host for the same species,<sup>5</sup> we can reasonably speculate that other felids also may serve as the final host for *S. leporum*.

Failure to infect two cats (Exps. 2 and 3) may have been caused by deterioration or death of the parasite. Rabbit carcasses shipped from Pennsylvania to Maryland may have been dead as long as a week before they were fed.

Reinfection of a cat that had stopped shedding sporocysts from a previous infection parallels findings in previous studies.<sup>2</sup> The carnivore final host does not appear to develop immunity to the parasite.<sup>2</sup>

The inability to infect domestic rabbits with *S. leporum* sporocysts from cats may reflect a specificity that parasites of this genus have for the intermediate host. Sporocysts of *S. fusiformis* from dogs fed infected beef were not infectious for sheep nor were *S. tenella* sporocysts from dogs fed infected lamb infectious for calves. Thus, the domestic rabbit apparently will not provide us with a suitable laboratory model with which to study *S. leporum*.

#### LITERATURE CITED

- CRAWLEY, H. 1914. Two new sarcosporidia. Proc. Acad. Nat. Sci. Phila. 66: 214-218.
- 2. FAYER, R. 1974. Development of Sarcocystis fusiformis in the small intestine of the dog. J. Parasit. 60: 660-665.
- 3. ——— and A. J. JOHNSON. 1973. Development of Sarcocystis fusiformis in calves infected with sporocysts from dogs. J. Parasit. 59: 1135-1137.
- 4. —— and ——. 1974. Sarcocystis fusiformis: development of cysts in calves infected with sporocysts from dogs. Proc. Helm. Soc. Wash. 41: 105-108.
- 5. ——, —— and HILDEBRANDT. 1976. Oral infection of mammals with Sarcocystis fusiformis bradyzoites from cattle and sporocysts from dogs and coyotes. J. Parasit. 62: 10-14.
- GESTRICH, R., A. O. HEYDORN and N. BAYSU. 1975. Beitrage zum Lebenszyklus der Sarkosporidien VI. Untersuchungen zur Artendifferenzierung bei Sarcocystis fusiformis and Sarcocystis tenella. Berl. Munch. Tierarzt. Wochen. 88: 191-197.
- 7. HAMMOND, D. M., B. CHOBOTAR and J. V. ERNST. 1968. Cytological observations on sporozoites of *Eimeria bovis* and *E. auburnensis* and an *Eimeria* species from the Ord kangaroo rat. J. Parasit. 54: 550-558.
- 8. HUDKINS-VIVION, G., T. P. KISTNER and R. FAYER. 1976. Possible species differences between *Sarcocystis* from mule deer and cattle. J. Wildl. Dis. 12: 86-87.
- 9. ROMMEL, M., A. O. HEYDORN and F. GRUBER. 1972. Beitrage zum Lebenszyklus der Sarkosporidien. I. Die Sporozyste von S. tenella in den Fazes der Katze. Berl. Munch. Tierarzt. Wochen. 85: 121-123.
- RZEPCZYK, C. M. 1974. Evidence of a rat-snake life cycle for Sarcocystis. Internat. J. Parasit. 4: 447-449.
- 11. VANDE VUSSE, F. J. 1965. Sarcocystis infections in relation to age of Iowa cottontails (Protozoa:Sarcocystidae). Iowa Acad. Sci. 72: 524-528.

Received for publication 22 October 1976