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HELMINTHS AND ECTOPARASITES OF THE COMMON SNIPE (Capella gallinago L.) FROM SOUTHWEST TEXAS AND MONTE VISTA NATIONAL WILDLIFE REFUGE, COLORADO.

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Abstract: Sixty common snipe, (Capella gallinago), collected from Hudspeth County, Texas and 13 collected from Monte Vista National Wildlife Refuge, Colorado were examined for metazoan parasites. The parasites recovered were Cestoda: Amoebotaenia fuhrmanni, Haploparaxis brachyphallos, Haploparaxis crassirostris, Haploparaxis echinovatum, Haploparaxis sp., Hymenolepis calumnacantha, Hymenolepis sp. I, Hymenolepis sp. II; Trematoda: Cyclocoelum mutabile, Echinostoma revolutum, Tanaisia fedtschenkoi; Nematoda: Capillaria contorta, Cosmocephalus capellae, Tetrameres coloradensis; Acanthocephala: Arhythmorhynchus capellae; Mallophaga: Austromenopon durisetosum and Rhynonirmus scolopacis.

More species of parasites were recovered from fall migrants; (12), than from spring migrants, (10).

The cestode *Haploparaxis echinovatum* was recorded from North America for the first time.

The parasite fauna recorded in this study did show some concentration for dominance by Simpson's index (0.33). This parasite fauna was most similar to that reported by Schmidt from snipe collected in Northern Colorado (67%).

INTRODUCTION

Investigations on the parasites of the common snipe, (Capella gallinago), in North America are limited. Deblock and Rausch⁴ described several species of Haploparaxis from snipe in Alaska, and Schmidt^{8,9,10,11} described species of Hymenolepis, Tetrameres, and Arhythmorhynchus, from snipe collected in Northern Colorado. Threlfall¹⁵ examined common snipe from Newfoundland, Ontario, and Louisiana for parasites and noted differences in geographical distribution of the parasites and reported on their prevalence. Tuck¹⁷ lists species of parasites, geographic location (Palearctic and Nearctic), and abbreviated citations in his monograph The Snipes.

The common snipe is a migratory shore bird who occupies wetland

habitats and is considered an excellent game and table bird. According to Fogarty and Deith⁵ the bird, "is distributed throughout almost the whole North American continent, from Point Hope, Alaska, on the Arctic Ocean, south to Brownsville, Texas, on the Gulf Coast, and through most of central and eastern Mexico, all of Central America and the Northern part of South America in Columbia and Venezuela." Capella gallinago is a common migrant in the Southwest and utilizes the Rio Grande Valley on its annual migrations. The migration from the breeding grounds begins in September and early October. The migration from the wintering grounds begins in April and extends to about the middle of May.7

The Monte Vista National Wildlife Refuge, Colorado, located in the San Juan Basin near the upper reaches of the Rio Grande, hosts both migratory and nesting common snipe.

The purpose of this paper is to report on the species and prevalence of parasites harbored by the common snipe collected in the Rio Grande Valley in Hudspeth County, Texas in the fall and spring and at Monte Vista National Wildlife Refuge, Colorado in August. The geographic distribution of parasites reported in this study and by others is summarized.

METHODS AND MATERIALS

Seventy-three common snipe were examined for metazoan parasites. Sixty were collected from the Rio Grande Valley, Hudspeth County, Texas in the fall of 1976 and 1977 and the spring of 1976, 1977, and 1978. Thirteen were collected from Monte Vista National Wildlife Refuge, Rio Grande County, Colorado in August, 1978. Birds were shot and placed in individual plastic bags. Those not examined within six h were frozen and examined at a later date. All birds were classified as adults because juveniles and adults could not be distinguished. Sex was determined by examination of the gonads. The birds were not examined for blood parasites.

Trematodes and cestodes were fixed in alcohol-formalin-acetic acid (AFA), stained in Ehrlich's hematoxylin and mounted in Canada balsam. Nematodes were fixed in 70% ethanol and cleared and examined in temporary lactophenol mounts. Acanthocephalids were fixed in AFA, stained in Van Cleave's hematoxylin and mounted in Permount. Ectoparasites were fixed in 70% ethanol, cleared in 10% KOH and mounted in polyvinyl alcohol or in Lipshaw's mounting media.

Differences between numbers of a species of a parasite harbored by fall and spring migrants were analyzed by applying the Chi-square test without an *a priori* hypothesis and utilizing the Yates correction factor. Significance was assumed for P < .05. Mean intensity

ratios (MIR) were calculated by using the smaller mean intensity from fall or spring as the denominator. In some cases statistical analysis was not done because the number of parasites or their distribution was not appropriate.

Simpson's index was calculated to measure the concentration for dominance of parasite species in snipe collected in Hudspeth County, Texas and Sorenson's index of similarity was used to compare parasite faunas from Southwest Texas to other areas in North America, as used by Stone and Pence.¹³

Two terms proposed by Canaris *et al.*² are used in this manuscript. They are: Parasite Geographic Range (PGR) = the total region that a parasite is transported by the migratory bird host. Parasite Infective Geographic Range (PIGR) = total region where infection of the migratory bird host is known to take place. The Parasite Geographic Range is equal to or greater than the Parasite Infective Geographic Range (PGR \geq PIGR).

Representative slides of parasites have been sent to Dr. Malcom E. McDonald, National Fish and Wildlife Health Laboratory. Fish and Wildlife Service, 1655 Linden Drive, Madison, Wisconsin 53706. Others have been retained in the Zoonoses Laboratory, Dept. Bio. Sci. University of Texas at El Paso, El Paso, Texas 79968.

RESULTS AND DISCUSSION

Eighty three percent of the snipe from Hudspeth County were infected with at least one species of parasite. Fourteen species of helminths and two species of mallophagan lice were recovered (Table 1). Six species of helminths were recorded from fall migrants, four from spring, and four species were ubiquitous.

Twenty-three percent of the birds were infested with lice. Austromenopon durisetosum and Rhynonirmus scolopacis were present on spring and fall migrants from Hudspeth County,

| | FAL | FALL (27) | SPRIN | SPRING (33) | TOTA | TOTAL (60) | | |
|----------------------------|---------------|--------------------------|---------------|--------------------------|---------------|--------------------------|----------------|---------------------------|
| | Numt % In: | Number and % Infected | Numb % Int | Number and % Infected | Numb % Int | Number and % Infected | M Intensity | Mean Intensity & Range |
| CESTODA (7) | | | | | | | | |
| Amoebotaenia fuhrmanni | 2 | 7.4 | 4 | 12 | 9 | 10.0 | 7.3 | 1-33 |
| Haploparaxis brachyphallos | e | 11.1 | 0 | 0 | e | 5.0 | 34.3 | 1-33 |
| Haploparaxis crassirostris | 1 | 3.7 | 0 | 0 | 1 | 1.6 | 20.0 | 20 |
| Haploparaxis echinovatum | 0 | 0 | ę | 6 | e | 5.0 | 2.7 | 1-5 |
| Haploparaxis sp. | e | 11.1 | æ | 24 | 11 | 18.3 | 3.6 | 1-7 |
| Hymenolepis calumnacantha | e | 11.1 | 0 | 0 | e | 5.0 | 6.7 | 6-7 |
| Hymenolepis sp. I | 0 | 0 | 4 | 12 | 4 | 6.6 | 10.5 | 1-15 |
| FREMATODA (3) | | | | | | | | |
| Cyclocoelum mutabile | 1 | 3.7 | 0 | 0 | 1 | 1.6 | 1.0 | 1 |
| Echinostoma revolutum | 0 | 0 | 1 | e | 1 | 1.6 | 2.0 | 2 |
| Tanaisia fedtschenkoi | 7 | 25.9 | 6 | 27 | 16 | 26.6 | 7.1 | 1-56 |
| NEMATODA (3) | | | | | | | | |
| Capillaria contorta | 1 | 3.7 | 0 | 0 | 1 | 1.6 | 2.0 | 2 |
| Cosmocephalus capellae | 1 | 3.7 | 0 | 0 | 1 | 1.6 | 12.0 | 12 |
| Tetrameres coloradensis | 6 | 33.3 | 9 | 18 | 15 | 25.0 | 21.5 | 1-185 |
| ACANTHOCEPHALA (1) | | | | | | | | |
| Arhythmorhynchus capellae | 0 | 0 | 2 | 9 | 2 | 3.3 | 1.0 | 1 |
| MALLOPHAGA (2) | | | | | | | | |
| Austromenopon durisetosum | 5 | 18.5 | 4 | 12 | 6 | 15.0 | 2.2 | 1-9 |
| Rhynonirmus scolopacis | 5 | 18.5 | ę | 6 | 80 | 13.3 | 3.0 | 1-10 |

TABLE 2. Parasites of the common snipe (Capella gallinago) from Monte Vista National Wildlife Refuge, Rio Grande County, Colorado.

| Augu | ist 1978 (13) | | | |
|---------------------------|---------------|-------------------|------------------|-------------------|
| | | ber and fected | Mean II and F | ntensity Range |
| CESTODA | | | | |
| Hymenolepis sp. II | 4 | 30.8 | 3.25 | 1-10 |
| TREMATODA | | | | |
| Tanaisia fedtschenkoi | 1 | 7.7 | 16.0 | 16 |
| MALLOPHAGA | | | | |
| Austromenopon durisetosum | 2 | 15.4 | 3.0 | 5 |
| Rhynonirmus scolopacis | 2 | 15.4 | 4.0 | 4 |

Texas and on birds collected in August from Monte Vista National Wildlife Refuge in southern Colorado. There was no significant difference in infestation between spring and fall birds.

Only two species of helminths, *Tanaisia fedtschenkoi* and *Hymenolepis* sp. II, and two species of lice were recovered from birds collected in Colorado. The percent of infected birds (62%) was not as great as those from Hudspeth County, 83% (Table 2).

The cestode, *Haploparaxis* echinovatum, is recorded for the first time from North America.

The mean intensities for the five major groups of parasites and their MIR's are given in Table 3. The mean intensities for the cestodes, nematodes and mallophaga was greater in fall birds. The mean intensities for the trematodes was strongly influenced by the number of kidney trematodes, *T. fedtschenkoi*. This parasite's PIGR may be more southerly and as a consequence would infect more of the spring migrants.

A checklist of helminth parasites reported for *C. gallinago* is given in Table 4. Of the 44 species of helminths recorded for the common snipe in North America, 15 species were recorded from snipe in this study.

The parasite fauna recorded here for the common snipe did show some concentration for dominance by Simpson's index (0.33). Most of the dominance was contributed by the nematode *Tetrameres coloradensis*. This fauna was most similar to that reported by Schmidt¹¹ from snipe collected in northern Colorado. The rank order for similarity among five regions in North America is given in Table 5. The greatest similarity for all regions is to Louisiana and the least is to Northern Colorado.

Cestoda. As a group, the prevalence and, in most instances, the mean intensities were highest for the cestodes. Three

TABLE 3. Mean intensity for parasite groups, fall and spring, common snipe, Hudspeth County, Texas.

| | FALL (27) | SPRING (33) | M.I.R. |
|----------------|-----------|-------------|--------|
| CESTODA | 7.3 | 1.8 | 4.1 |
| TREMATODA | 3.6 | 5.4 | 1.5 |
| NEMATODA | 11.7 | 0.6 | 19.5 |
| ACANTHOCEPHALA | 0.0 | 0.06 | 0.0 |
| MALLOPHAGA | 1.0 | 0.6 | 1.6 |

| Amoebotaenia fuhrmanni1.3.6.10.Amoebotaenia sp.1.3.3.Anomotaenia citrus1.2.3.4.Anomataenia variabilis4.Anomataenia sp.1.3.Choanotaenia cingulifera3.Choanotaenia manipurensis4.Diorchis sp.1.3.Haploparaxis brachyphallos9.10.Haploparaxis clerci6.Haploparaxis clerci6.Haploparaxis genetrans1.2.3.Haploparaxis genetrans2.Haploparaxis sp.1.2.3.Haploparaxis sp.1.2.3.Haploparaxis sp.1.2.3.Haploparaxis sp.1.2.3. | ပိ |
|--|-----|
| Amoebotaenia fuhrmanni1.3.6.10.Amoebotaenia sp.1.3.3.Anomotaenia citrus1.2.3.4.Anomataenia variabilis4.Anomataenia sp.1.3.Choanotaenia cingulifera3.Choanotaenia manipurensis4.Diorchis sp.1.3.Haploparaxis brachyphallos9.10.Haploparaxis clerci6.Haploparaxis clerci6.Haploparaxis grassirostris1.2.3.Haploparaxis parafilum9.Haploparaxis penetrans2.Haploparaxis sp.1.2.3.Haploparaxis sp.1.2.3. | 11. |
| Amoebotaenia sp.1.3.Anomotaenia citrus1.2.3.Anomataenia variabilis4.Anomataenia sp.1.3.Choanotaenia cingulifera3.Choanotaenia manipurensis4.Diorchis sp.1.3.Haploparaxis brachyphallos9.10.Haploparaxis clerci6.Haploparaxis crassirostris1.2.3.10.Haploparaxis filum1.2.3.10.Haploparaxis parafilum9.Haploparaxis sp.1.2.3.10. | |
| Anomataenia variabilis4.Anomataenia variabilis4.Anomataenia sp.1.Choanotaenia cingulifera3.Choanotaenia manipurensis4.Diorchis sp.1.Biorchis sp.1.Haploparaxis brachyphallos9.Haploparaxis clerci6.Haploparaxis crassirostris1.2.3.Haploparaxis filum1.2.3.Haploparaxis parafilum9.Haploparaxis sp.1.2.3.Haploparaxis sp.1.2.3. | |
| Anomataenia sp.1.3.Choanotaenia cingulifera3.Choanotaenia manipurensis4.Diorchis sp.1.Haploparaxis brachyphallos9.Haploparaxis clerci6.Haploparaxis echinotatum10.Haploparaxis filum1.2.3.Haploparaxis parafilum9.Haploparaxis sp.1.2.3.Haploparaxis ponetrans2.Haploparaxis sp.1.2.3.Haploparaxis ponetrans2.Haploparaxis sp.1.2.3. | |
| Choanotaenia cingulifera3.Choanotaenia manipurensis4.Diorchis sp.1.Haploparaxis brachyphallos9.Haploparaxis clerci6.Haploparaxis echinotatum10.Haploparaxis filum1.2.3.Haploparaxis parafilum9.Haploparaxis sp.1.2.3.Haploparaxis ponetrans2.Haploparaxis sp.1.2.3.Haploparaxis sp.1.2.3. | |
| Choanotaenia manipurensis4.Diorchis sp.1.Haploparaxis brachyphallos9.Haploparaxis clerci6.Haploparaxis crassirostris1.2.3.Haploparaxis filum1.2.3.Haploparaxis parafilum9.Haploparaxis sp.1.2.3.Haploparaxis sp.1.2.3.Haploparaxis ponetrans2.Haploparaxis sp.1.2.3.Haploparaxis sp.1.2.3. | |
| Diorchis sp.1.3.Haploparaxis brachyphallos9.10.Haploparaxis clerci6.Haploparaxis crassirostris1.2.3.10.Haploparaxis filum1.2.3.10.Haploparaxis parafilum9.Haploparaxis penetrans2.Haploparaxis sp.1.2.3.10.10. | |
| Haploparaxis brachyphallos9.10.Haploparaxis clerci6.Haploparaxis crassirostris1.2.3.10.Haploparaxis echinotatum10.Haploparaxis filum1.2.3.Haploparaxis parafilum9.Haploparaxis penetrans2.Haploparaxis sp.1.2.3.10.10. | |
| Haploparaxis clerci6.Haploparaxis crassirostris1.2.3.Haploparaxis echinotatum10.Haploparaxis filum1.2.3.Haploparaxis parafilum9.Haploparaxis penetrans2.Haploparaxis sp.1.2.3.10.2.3.10. | |
| Haploparaxis crassirostris1.2.3.10.Haploparaxis echinotatum10.Haploparaxis filum1.2.3.Haploparaxis parafilum9.Haploparaxis penetrans2.Haploparaxis sp.1.2.3.10.10. | |
| Haploparaxis echinotatum10.Haploparaxis filum1.2.3.Haploparaxis parafilum9.Haploparaxis penetrans2.Haploparaxis sp.1.2.3.10. | |
| Haploparaxis filum1.2.3.Haploparaxis parafilum9.Haploparaxis penetrans2.Haploparaxis sp.1.2.3.10. | |
| Haploparaxis filum1.2.3.Haploparaxis parafilum9.Haploparaxis penetrans2.Haploparaxis sp.1.2.3.10. | |
| Haploparaxis penetrans2.Haploparaxis sp.1.2.3.10. | |
| Haploparaxis sp. 1.2.3. 10. | |
| | |
| | |
| Hymenolepis calumnacantha 6. 10. | |
| Hymenolepis capellae 1. 3. | |
| <i>Hymenolepis</i> sp. 1. 6. 10. | 11. |
| Paricterotaenia decacantha 6. | |
| Paricterotaenia paradoxa 4.5. | |
| Paricterotaenia stellifera 4. | |
| Paricterotaenia sp. 1.2.3. | |
| Profimbriaria multicanalis 4. | |
| TREMATODA (16) 1.2.3. 4.5. 6. 7.8. 9. 10. | 11. |
| Cotylurus brevis 4. | |
| Cotylurus cornutus 1. 4. | |
| Cotylurus leidyi 1. | |
| Cyclocoelum mutabile 1. 3. 6. 10. | |
| Cyclocoelum obscurum 4. | |
| Cyclocoelum sp. 5. | |
| Echinostoma operosum 4. | |
| Echinostoma revolutum 1. 3. 5. 6. 10. | |
| Hypoderaeum sp. 4. | |
| Pulvinifer macrostomum 1. 3. 5. | |
| Pulvinifer singularis 6. | |
| | 11. |
| Tanaisia sp. 4. | |
| Wardianum wilsoni 4. | |
| Zygocotyle lunata 4. | |

TABLE 4. Checklist of helminth parasites of Capella gallinago from North America.

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| TABLE 4. (continued) | | | | | | |
|----------------------------|---|--|-----------------------------------|------------------------------------|---|--------------------------------------|
| | Threlfall ¹⁵ Newfoundland Ontario Louisiana | Threifall ¹⁶ USA Canada | Schmidt ¹¹ Colorado | Byrd & Denton' Texas Georgia | Deblock & Rausch ⁴ Alaska | This Study S.W. Texas Colorado |
| Dicrocoeliids unidentified | 1. | | | | | |
| NEMATODA (3) | | | | | | |
| Capillaria contorta | 3. | | 6. | | | 10. |
| Cosmocephalus capellae | 2.3. | | 6. | | | 10. |
| Tetrameres coloradensis | | | 6. | | | 10. |
| ACANTHOCEPHALA (1) | | | | | | |
| Arhythmorhynchus capellae | | | 6. | | | 10. |

species were present in fall migrants, two in spring migrants and one was ubiquitous (Table 1). There was no significant difference for species of cestodes between fall and spring birds.

Ten percent of the snipe were infected with Amoebotaenia fuhrmanni. Schmidt¹¹ reported A. fuhrmanni from birds collected in northern Colorado. Threlfall¹⁵ reported it in 8.2% of 24 snipe from Newfoundland and 16% of 268 snipe from Louisiana, but he did not observe this tapeworm in snipe from Ontario. The PGR and PIGR appears to be extensive.

Haploparaxis echinovatum was present in 3% of the snipe. Deblock and Rausch⁴ recovered and described this species from snipe, (C. gallinago), collected in Iran. *H. echinovatum* is recorded for the first time from North America. The PIGR may be more southerly since it was absent in birds collected in August from Monte Vista National Wildlife Refuge, Colorado and from all fall migrants.

Five percent of the snipe were infected with Haploparaxis brachyphallos. This parasite was reported by Deblock and Rausch⁴ from the common snipe, (C. gallinago), collected near Anchorage, Alaska and by Webster¹⁸ from the redbacked sandpiper, (Erolia alpina pacifica), from southeastern Alaska. This parasite was not recorded from spring migrants or from birds collected in August in southern Colorado. Schmidt¹¹ did not report this parasite

TABLE 5. Rank order of similarity-helminths, common snipe, North America.

| Louisiana | Newfoundland | S.W. Texas | Ontario | Northern Colorado |
|------------------------------|---------------|------------|---------|----------------------|
| NF 83 | LA 83 | NCO 67 | LA 60 | SWT 67 |
| ONT 60 | ONT 56 | LA 48 | NF 56 | LA 40 |
| SWT 48 | SWT 35 | NF 35 | SWT 21 | NF 26 |
| NCO 40 | NCO 26 | ONT 21 | NCO 11 | ONT 11 |
| $\Sigma = 231$ | 200 | 171 | 148 | 144 |
| $\overline{\mathbf{x}} = 58$ | 50 | 43 | 37 | 36 |

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from the common snipe, (C. gallinago), collected in northern Colorado. It is not clear how much of his material came from fall migrants. The PIGR based on the above, may be more northerly.

Only one snipe was infected with Haploparaxis crassirostris = (Aploparaksis rauschi Webster). It was a fall migrant and a total of 20 specimens were recovered from the small intestine. Threlfall¹⁵ reported a prevalence of 1.8% in Louisiana, 10.5% in Ontario, and 12.3% in Newfoundland for this parasite. Webster¹⁸ recovered this species of tapeworm from the redbacked sandpiper, E. a. pacifica, in the Stikine River flats near Wrangel, southeastern Alaska. The absence of this parasite from spring migrants in this study and the low prevalence in Louisiana as reported by Threlfall¹⁵ indicate that the PIGR is perhaps more northerly.

Schmidt¹⁰ described Hymenolepis calumnacantha from 27 specimens recovered from the small intestine and caeca of snipe collected in northern Colorado. In this study H. calumnacantha infected 5% of the snipe. The PIGR may be more northerly since it was not present in spring migrants or in birds collected at Monte Vista National Wildlife Refuge in southern Colorado.

Hymenolepis sp. I was recovered only from spring migrants. Species identification was not possible because specimens were in poor condition. Hymenolepis sp. I resembled Hymenolepis deblocki described by Schmidt and Neiland¹² from the short-billed dowitcher, Limnodromus griseus, collected near Bristol Bay, Alaska. The main difference between Hymenolepis sp. I and H. deblocki was the hooks. Hymenolepis sp. I had 10 hooks, each 80 µm long arranged in a circle. A guard, a long blade and long handle with a small knob on the end were present. H. deblocki had 10 hooks averaging 100-102 μ m, arranged in a circle, with the guard almost absent, and a long blade and long handle with a knob on the end present. The PGR is at least from Alaska to southwest Texas.

Hymenolepis sp. II was recovered from snipe collected in southern Colorado (Table 2). Four birds were infected with the cestode. Not enough material was obtained for an identification of the species.

Trematoda. Three species of trematodes were recorded and are listed in Table 1. Tanaisia fedtschenkoi is parasitic in the genito-urinary tract of aquatic and semiaquatic birds. This parasite was common in both fall and spring migrants including birds from Monte Vista National Wildlife Refuge, Colorado (Table 2). There was no significant difference in prevalence between fall and spring migrants but spring migrant's mean intensity was 20 and fall was 14 with an MIR of 1.4. Schmidt¹¹ reported this trematode from $C_{\rm c}$ gallinago collected in northern Colorado and Byrd and Denton¹ recorded it for Capella delicata (= C. gallinago) from Texas and Georgia. Cheatum³ described Tanaisia pelidnae, from a red-backed sandpiper, E. a. sakhalina, collected near Caseville, Michigan. This parasite was synonymized with T. fedtschenkoi by Byrd and Denton.¹ The PGR is extensive and it has been recorded from the orders Passeriformes, Gruiformes, and Charadriiformes from Russia, Turkestan, Siberia and Macedonia.

Cyclocoelum mutabile was found in the thoracic cavity of one fall migrant and only one parasite was recovered. Schmidt¹¹ reported it from *C. gallinago* collected in northern Colorado and Threlfall¹⁵ recorded prevalences of 7.3% from Newfoundland and 19% from Louisiana. The PGR appears to be extensive.

Two specimens of *Echinostoma revolutum* were recorded from one spring migrant. This parasite was also recorded by Schmidt¹¹ from northern Colorado and by Threlfall¹⁵ from Newfoundland and Louisiana with a 7.3% and 10.8% prevalence respectively. The PGR and the number of different hosts for this parasite are known to be extensive. The reason for the very low prevalence recorded in this study is not known.

Nematoda. Three species of nematodes were recovered, all from the proventriculus (Table 1). Cosmocephalus capellae and Capillaria contorta were reported by Schmidt¹¹ from northern Colorado. Threlfall¹⁵ noted a prevalence of 42.1% from Ontario and 0.3% from Louisiana for C. capellae. The prevalence for C. contorta was 1.4% and was reported only from Louisiana. In another study Threlfall¹⁴ examined two snipe from Newfoundland and found a single infection of C. contorta. There was no significant difference between fall and spring migrants for these two species of nematodes. The low prevalence of these two species in Texas and Louisiana suggest that the PIGR may be more northerly.

Tetrameres coloradensis was a ubiquitous parasite and there was no significant difference in prevalence between fall and spring migrants, but fall migrant's mean intensity was 33 and spring migrant's was 3.5. This yielded an MIR of 9.4. Schmidt* described this species of parasite from C. gallinago collected in northern Colorado. Its PGR, as it is presently understood, appears to be from northern Colorado, southward. Its PIGR may be confined to northern Colorado or further north. Evidence that supports this is the apparent loss of nematodes in returning spring migrants, fall migrants with both female and male nematodes present but spring migrants with single sex infections, spring migrants with a lower mean intensity than fall migrants, and the fact that the parasite was not recorded from snipe collected from Monte Vista National Wildlife Refuge in southern Colorado.

Acanthocephala. During the spring of 1976 and 1978, single infections of Arhythmorhynchus jeffreyi (Schmidt 1963) Schmidt 1973, were recorded from two spring migrants. Schmidt9 recovered and described this species from six snipe collected in northern Colorado. This parasite has not been reported from other geographic areas or from other hosts. Its PGR extends from northern Colorado to southwest Texas and since it was in returning spring migrants its PGR may extend further south. This parasite was not recovered from birds collected at Monte Vista National Wildlife Refuge in southern Colorado. Therefore, its PGR may be more northerly.

Mallophaga. Two species of Mallophaga, Austromenopon durisetosum and Rhynonirmus scolopacis infested the common snipe, Table 1. There was no significant difference in prevalence for either species between spring and fall migrants. The mean intensity for A. durisetosum was greater for spring migrants with an MIR of 2.4. The mean intensity for R. scolopacis was greater in the fall with an MIR of 3.1. Geist⁶ reported a prevalence of 50% for Mallophaga from charadriiform birds collected in Ohio. He observed a decline in adult Mallophaga during late summer and early fall with few or no adults observed during the winter months, and an increase of adults during the late spring and early summer. The prevalence for Mallophaga in this study was lower, 23.2%, but the population trend was generally similar to that reported by Geist.

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