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## DETECTION OF ANTIBODIES REACTIVE WITH *EHRLICHIA CHAFFEENSIS* IN THE RACCOON

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**ABSTRACT:** Antibodies reactive with *Ehrlichia chaffeensis* were detected in raccoon (*Procyon lotor*) serum samples by using an indirect immunofluorescence assay. Samples from 411 raccoons trapped in the southeastern United States from 1977 to 1999 were tested. Serologically reactive samples with reciprocal titers of  $\geq 16$  were detected from 83 raccoons (20%) from 13 of 16 counties in eight states, indicating that raccoons are commonly exposed to *E. chaffeensis*. Samples collected as early as 1977 were positive. A polymerase chain reaction assay specific for *E. chaffeensis* failed to detect the presence of ehrlichial DNA in serum samples from 20 representative seroreactive raccoons. Because of serologic cross-reactivity among antigens derived from different *Ehrlichia* spp., additional immunologic, molecular, or culture-based studies will be required to confirm *E. chaffeensis* infections of raccoons in the southeastern United States.

**Key words:** *Ehrlichia chaffeensis*, immunofluorescence assay, *Procyon lotor*, raccoon, survey.

### INTRODUCTION

In the USA, human ehrlichiosis is caused by three distinct *Ehrlichia* spp.: *E. chaffeensis*, *E. ewingii*, and the human granulocytic ehrlichiosis agent (HGE, putatively *E. phagocytophila*). The clinical diseases caused by these three pathogens are similar, although evidence indicates that infection with *E. chaffeensis* may result in more severe disease (Standaert et al., 1998). The greatest number of cases reported have been due to infection with *E. chaffeensis*, which was first recognized in 1986 (Maeda et al., 1987; Eng et al., 1990; Anderson et al., 1991; Fishbein et al., 1994). As of 1997, there had been 742 cases of *E. chaffeensis* reported, and most of these cases occurred in the south-central and southeastern areas of the USA (McQuiston et al., 1999). Several studies suggest that the lone star tick, *Amblyomma americanum*, is the primary vector of *E. chaffeensis* (Anderson et al., 1993; Lockhart et al., 1995, 1996).

It is important to understand the natural history of *E. chaffeensis* so that measures for disease prevention may be designed and implemented; however, knowledge of the maintenance cycle of *E. chaffeensis* is incomplete. Isolation of *E. chaffeensis*

from white-tailed deer (*Odocoileus virginianus*) (Lockhart et al., 1997b), molecular studies of tick vectors (Anderson et al., 1992; Anderson et al., 1993), and studies of experimental transmission between deer and *A. americanum* (Ewing et al., 1995) indicate that deer are important reservoir hosts of *E. chaffeensis* and that the agent may be maintained in a natural cycle involving white-tailed deer and lone star ticks.

Other wildlife species are suspected to play a role in the epizootiology of *E. chaffeensis*. Lockhart et al. (1997a) reported that nine of 43 raccoons (*Procyon lotor*) (21%) trapped at an *E. chaffeensis*-enzootic site in Georgia had antibodies reactive with *E. chaffeensis*. Because the raccoon is heavily parasitized by immature and adult *A. americanum* (Tugwell and Lancaster, 1962; Cooney and Burgdorfer, 1974; Koch and Dunn, 1980; Zimmerman et al., 1988), it is likely that raccoons are routinely exposed to this pathogen and may possibly be involved in its maintenance. We conducted a serosurvey using an indirect immunofluorescence assay (IFA) to evaluate the prevalence of antibodies reactive to *E. chaffeensis* among raccoons in the southeastern USA in order to better define the

role of the raccoon in the natural history of this pathogen.

## MATERIALS AND METHODS

### Acquisition of samples and selection of controls

Archived raccoon serum samples were obtained through the Southeastern Cooperative Wildlife Disease Study (College of Veterinary Medicine, University of Georgia, Athens, Georgia, USA). These samples were collected between 1977 and 1987 and had been stored at  $-20^{\circ}\text{C}$ . Samples from Liberty and Wakulla Counties (Florida, USA) were collected in 1999 from areas in the general vicinity of where several cases of human infection with *E. chaffeensis* have occurred (Sumner et al., 1999). Samples from Pinellas Co. (Florida) were collected in 1998 as part of an oral rabies vaccine study.

A raccoon serum sample from Whatcom Co. (Washington, USA) was selected as a negative control because it gave no fluorescence in the IFA and was collected far outside the geographic range of *A. americanum* (Means and White, 1997). A raccoon serum sample empirically selected by screening samples from an area where *E. chaffeensis* is enzootic (Craven Co., North Carolina, USA) served as a positive control. The reciprocal plateau endpoint titer of the sample was high (512), and the pattern of specific fluorescence it gave in the IFA (density, distribution, and intensity) was consistent with that observed with seropositive humans proven to be infected with *E. chaffeensis* by other tests (e.g., bacterial isolation). Aliquots of both control serum samples were frozen to ensure consistency during future testing.

### Antigen preparation and use of IFA

Teflon-templated glass slides were obtained from Erie Scientific Co. (Portsmouth, New Hampshire, USA). *Ehrlichia chaffeensis* (Arkansas strain) was grown in DH82 canine macrophage cells, and infected cells were applied to the slides as described (Nicholson et al., 1997). Briefly, approximately 50  $\mu\text{l}$  of living cells in culture medium (approximately  $3 \times 10^6$  cells per ml) was applied to each well of the antigen slide, and slides were incubated for 15 min at  $37^{\circ}\text{C}$  in a humidified plastic chamber. Culture medium containing any nonadherent cells was then removed by vacuum pipetting, and the slides were air dried, fixed in acetone for 15 min, and stored at  $-70^{\circ}\text{C}$  until used.

The IFA was conducted using conditions and reagents described previously for the HGE agent (Comer et al., 1999a). The diluent was modified by incorporating a 10% suspension of freeze-disrupted normal, uninfected DH82

cells (approximately  $3 \times 10^6$  cells/ml culture medium) and 2% normal heat-inactivated goat serum to reduce potential background staining. Because the expected range of antibody titers could not be predicted, raccoon serum samples were screened at a 1/16 dilution; no difficulty in reading slides due to nonspecific background staining was evident at this dilution. Samples that were reactive at the 1/16 dilution were then titrated to endpoint. Fluorescein isothiocyanate-labeled goat anti-raccoon IgG (heavy and light chain specific) conjugate (Kirkegaard & Perry Laboratories, Inc., Gaithersburg, Maryland, USA) was used at an optimal working dilution (1/100). Multiple dilutions of conjugate were tested against multiple dilutions of positive and negative controls to determine this dilution ("checkerboard titration"). Samples that were contaminated with bacterial or fungal growth were filtered (Spin-X 0.22  $\mu\text{m}$  filter, Costar Corp., Cambridge, Massachusetts, USA) before dilution series were prepared.

All titers  $\geq 16$  were expressed as the greatest reciprocal dilution in which specific ehrlichial reactivity was observed. Titers were log-transformed (base 2) for analysis (Thrusfield, 1995), and geometric mean titers (GMT) were calculated where appropriate. Statistical analysis was conducted using EpiInfo Version 6.04b (Dean et al., 1994). A *P*-value of  $\leq 0.05$  was considered significant.

### Polymerase chain reaction studies

We attempted to amplify *E. chaffeensis* DNA from 20 raccoon serum samples which were selected to be representative of all seroreactive samples tested. These samples originated from three barrier island and three inland sites in five states. DNA extracts were prepared from serum as previously described (Comer et al., 1999b) and evaluated by using a nested polymerase chain reaction (PCR) assay to test for the presence of the *E. chaffeensis* variable length PCR target (VLPT) (Sumner et al., 1999). DNA extracted from *E. chaffeensis*-infected DH82 cells and a water blank were included as positive and negative controls, respectively.

## RESULTS

We tested 411 raccoons collected from 16 counties in eight states (Table 1, Fig. 1). The overall prevalence of reactivity with *E. chaffeensis* antigens was 20% (83 reactive raccoons from 13 counties in all states excluding Washington), but prevalence varied widely by county. Titers

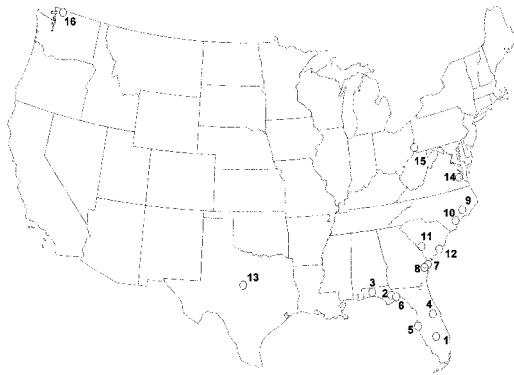


FIGURE 1. Distribution of raccoon collection sites for studies on ehrlichiae. Refer to TABLE 1 for information on individual sites.

ranged from 16 to 1,024, with a modal titer of 64 and an overall GMT of 60 (Fig. 2). Forty-six animals (11%) had a titer that exceeded the GMT for the group.

Higher prevalences of antibodies were significantly associated with raccoon samples from barrier islands when compared with mainland locations ( $\chi^2 = 106.90$ , odds ratio = 15.70, confidence interval 8.23–30.17,  $P < 0.001$ ). The prevalences of antibodies reactive with *E. chaffeensis* antigens of the three island populations sampled were 67% on Bull Island (South Carolina, USA), 80% on St. Catherines Island (Georgia, USA), and 88% on Ossabaw Island (Georgia).

Seropositive raccoons were detected each year in which raccoons were collected (Table 1). Seropositive raccoons were identified from several states in 1977 and 1978, the earliest years for which samples were available. The highest county-wide GMT observed (128) occurred in Craven County, North Carolina, and Brown County, Texas.

Markedly different prevalences were obtained from sites sampled from within a single state (Table 1). The very low prevalence in Okaloosa County (Florida) in 1979 (1%) was in marked contrast to seroprevalences among raccoons in closely situated counties in the Florida Panhandle in 1999 (Liberty Co., 75%; Wakulla Co., 91%). Pinellas County in central Florida,

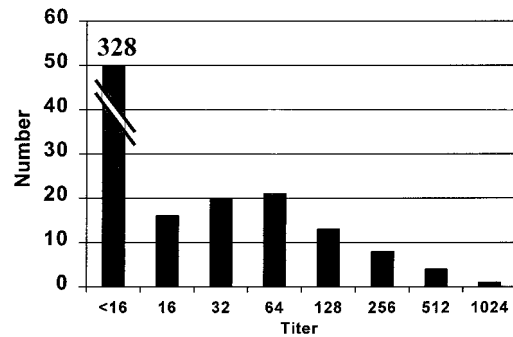


FIGURE 2. Frequency distribution of antibody titers to *Ehrlichia chaffeensis* in 411 raccoons.

sampled in 1998, also had a low prevalence of reactive samples (<1%).

In the PCR assays, positive and negative DNA controls gave the expected results when the VLPT primers were used. However, no products were amplified from the raccoon serum samples. The samples ranged from 32 to 512 in antibody titer (GMT = 104) and came from populations where the prevalence ranged from 18% to 88%.

## DISCUSSION

Overall, antibodies reactive with *E. chaffeensis* were detected from 20% of the raccoons sampled in this study. The prevalence of reactive antibodies on barrier islands was high (67%–88%) and significantly greater than the prevalence from mainland sites, although two inland coastal counties from the Florida Panhandle also had very high prevalences. Pung et al. (1994) speculated that *A. americanum* may be more abundant on barrier islands than on mainland sites when comparing the high tick burdens on raccoons from St. Catherines Island (Georgia) to those from other southeastern mainland counties. However, very high densities of *A. americanum* are common in many mainland regions, and this tick is a notorious pest species to humans and animals. Increased vector density should enhance *Ehrlichia* spp. transmission among vertebrate hosts, yet lower prevalences of *E. chaffeensis*-infected *A. americanum* were found on St.

TABLE 1. Serologic reactivity of 414 raccoons to *Ehrlichia chaffeensis* antigens.

| State          | County     | Map site | Location (map coordinates)               | Year(s) collected | Number positive/number tested <sup>b</sup> | Percent positive | Range of titers | GMT <sup>a</sup> |
|----------------|------------|----------|------------------------------------------|-------------------|--------------------------------------------|------------------|-----------------|------------------|
| Florida        | Glades     | 1        | Fisheating Creek (26°59'N, 81°22'W)      | 1977              | 1/12                                       | 8                | 32              | —                |
| Florida        | Liberty    | 2        | Torreya State Park (30°17'N, 84°83'W)    | 1999              | 6/8                                        | 75               | 16–128          | 51               |
| Florida        | Okaloosa   | 3        | Crestview (30°41'N, 86°59'W)             | 1979              | 1/88                                       | 1                | 16              | —                |
| Florida        | Orange     | 4        | unknown (28°34'N, 81°26'W)               | 1977              | 0/3                                        | 0                | —               | —                |
| Florida        | Pinellas   | 5        | unknown (27°53'N, 82°72'W)               | 1998              | 2/160                                      | 1                | 16–64           | 32               |
| Florida        | Wakulla    | 6        | Wakulla Springs Park (30°8'N, 84°39'W)   | 1999              | 10/11                                      | 91               | 16–1,024        | 104              |
| Georgia        | Chatham    | 7        | Ossabaw Island (31°58'N, 81°12'W)        | 1981              | 21/34                                      | 88               | 16–256          | 46               |
| Georgia        | Liberty    | 8        | St. Catherines Island (31°49'N, 81°48'W) | 1977              | 8/10                                       | 80               | 16–128          | 54               |
| North Carolina | Craven     | 9        | Cherry Point (35°7'N, 77°05'W)           | 1979              | 5/11                                       | 45               | 32–512          | 128              |
| North Carolina | Pender     | 10       | unknown (34°30'N, 77°89'W)               | 1978              | 5/9                                        | 56               | 16–256          | 42               |
| South Carolina | Barnwell   | 11       | Savannah River Plant (33°17'N, 81°48'W)  | unknown           | 0/1                                        | 0                | —               | —                |
| South Carolina | Charleston | 12       | Bull Island (32°51'N, 79°86'W)           | 1987              | 16/24                                      | 67               | 16–512          | 73               |
| Texas          | Brown      | 13       | Brownwood (31°46'N, 98°93'W)             | 1977, 1978        | 3/17                                       | 18               | 64–256          | 128              |
| Virginia       | James City | 14       | Williamsburg (37°18'N, 76°75'W)          | 1978              | 1/10                                       | 10               | 16              | —                |
| West Virginia  | Ohio       | 15       | unknown (40°6'N, 80°63'W)                | 1977              | 4/11                                       | 36               | 32–128          | 38               |
| Washington     | Whatcom    | 16       | unknown (48°49'N, 121°76'W)              | unknown           | 0/2                                        | 0                | —               | —                |
| Total          |            |          |                                          |                   | 83/411                                     | 20.2             | 16–1,024        | 60.4             |

<sup>a</sup> Geometric mean titers of  $\geq 16$ .<sup>b</sup> Positive titer  $\geq 16$ .

Catherines Island (0.9%) and neighboring Sapelo Island (0.0%) than at a nearby mainland site (9.3%) (Whitlock et al., 2000). As a tick-borne disease, a focal distribution of ehrlichiosis would be expected, and differences in prevalence among collection sites presumably reflects the distribution of *E. chaffeensis* and the abundance of suitable tick vectors. The effect of vector density on transmission of *E. chaffeensis* is not fully understood.

A high antibody prevalence was noted in 1999 among raccoons from two counties

in the Florida Panhandle from sites near locations where several human cases of ehrlichiosis due to *E. chaffeensis* have been reported (Paddock et al., 1997; Sumner et al., 1999). However, a low seroprevalence was detected in nearby Okaloosa County (using samples collected in 1979). This low prevalence may reflect a difference in the true prevalence of infection or a difference in the ecology of tick or reservoir host populations two decades ago (Lockhart et al., 1995).

Reactive samples were found in rac-

coons collected in the late 1970s, indicating that *E. chaffeensis* may have been circulating among raccoons a decade before human infections with *E. chaffeensis* were initially recognized (Maeda et al., 1987). Furthermore, seroreactive raccoons were distributed in several southeastern states, indicating a widespread distribution of *E. chaffeensis*. Because they are readily fed upon by *A. americanum* and other ticks, are found throughout most of North America, and are easily trapped, raccoons may prove useful for surveillance of ehrlichiae and other tick-borne pathogens.

We used a minimum dilution of 1/16 in our screening assay of the raccoons. Other studies of *E. chaffeensis* infection in wildlife (Lockhart et al., 1997a,b; Little et al., 1998) have used 1/64 as a seropositive screening dilution. Because the spectrum of illness and magnitude of the antibody response associated with *E. chaffeensis* infection in raccoons is not known and could not be predicted, we reported the full range of titers obtained rather than assign an arbitrary cutoff value. Examination of Figure 2 reveals a normal distribution of log-transformed titers, a pattern well established for serologic data (Thrusfield, 1995). This evidence supports our approach. The distribution further suggests that a lower dilution might have been appropriate for screening our samples. Our assay has a strikingly low level of background staining and can be used to test lower dilutions of samples. Had a screening titer of 64 been used, the resulting frequency distribution would have been distinctly non-normal.

Serologic reactivity to *E. chaffeensis* antigens among the raccoons sampled does not preclude the possibility that some of the reactivity noted may have been caused by other, antigenically related, *Ehrlichia* spp. Potential for serologic cross-reaction also has been recognized for seropositive white-tailed deer (Dawson et al., 1996). Raccoons are host to immature and adult stages of at least three tick species which are recognized vectors of five *Ehrlichia*

spp., although not all are known from the locations tested. Raccoons are heavily parasitized by both immature and adult *A. americanum* (Tugwell and Lancaster, 1962; Cooney and Burgdorfer, 1974; Koch and Dunn, 1980; Zimmerman et al., 1988), a putative vector of *E. chaffeensis* to deer and humans (Anderson et al., 1993; Lockhart et al., 1997b), *E. ewingii* to humans and dogs (Anziani et al., 1990; Buller et al., 1999), and the white-tailed deer *Ehrlichia* agent among deer (Dawson et al., 1996; Brandsma et al., 1999). Raccoons are also frequently parasitized by adult *Dermacentor variabilis* (Tugwell and Lancaster, 1962; Rabinowitz et al., 1983; Mock et al., 1991; Kollars, 1993; Kollars and Ladine, 1999), a tick species that has been incriminated as a secondary vector in the transmission of *E. canis* to dogs (Johnson et al., 1998). Furthermore, *E. chaffeensis* DNA was amplified from one of four *D. variabilis* removed from an opossum (*Didelphis virginiana*) in Arkansas, USA (Anderson et al., 1992). Raccoons are also important hosts for *Ixodes scapularis* larvae and nymphs (Fish and Daniels, 1990; Manelli et al., 1993), a vector of the HGE agent to animals and humans in the northeastern USA (Vignes and Fish, 1997). Multiple ehrlichial species have been reported from a single coastal island site in Georgia (Little et al., 1998). Thus, infection of raccoons by more than a single ehrlichial species is possible in certain locations where the pathogens and vector tick populations occur together.

Serologic cross-reactivity within the genus *Ehrlichia*, especially among species within the same genogroup (Dumler et al., 1995), has been useful in diagnostic testing. *Ehrlichia canis* was used as a surrogate antigen to detect human infection with *E. chaffeensis* before the latter organism was adapted to continuous cell culture (Eng et al., 1990). *Ehrlichia canis* and *E. chaffeensis* are presently used as surrogate antigens to detect *E. ewingii* infection in dogs and humans, respectively, because *E. ewingii* has not yet been propagated in

continuous cell culture (Buller et al., 1999). Antibodies to the white-tailed deer agent (also not yet propagated in continuous cell culture) may cross-react with *E. chaffeensis* (Dawson et al., 1996), and at present only molecular methods are able to reliably detect the presence of this organism and *E. ewingii*. It is possible that some of the reactivity to *E. chaffeensis* antigens observed among raccoon serum samples in this study may represent reactivity to related species of *Ehrlichia*.

Further immunologic studies may provide more definitive confirmatory evidence that these raccoons were exposed to *E. chaffeensis*. Cultivation of the agent would be most desirable, and *E. chaffeensis* has been isolated from other animals, including white-tailed deer (Lockhart et al., 1997b) and goats (Dugan et al., 2000). However, other ehrlichial agents which potentially might have induced the antibody response observed (e.g., *E. ewingii* and the white-tailed deer *Ehrlichia* agent) have not yet been successfully isolated in cell culture. PCR amplification and sequencing might also be used to identify the etiologic agent. Adsorption with specific antigens or blocking with specific antibodies may help improve confirmatory assays (Comer et al., 1999c). Additional analysis of the immune response by Western immunoblotting methods would help to confirm the etiologic agent (Rikihisa et al., 1994).

Our inability to amplify ehrlichial DNA from serum specimens was not unexpected, as this substrate has yielded limited positive results when applied to acute-phase human serum samples (Comer et al., 1999b) or deer serum samples (Little et al., 1998). Furthermore, the stage of ehrlichial infection in the raccoons at the time of serum collection was unknown. More useful samples, such as EDTA-anticoagulated blood or splenic tissue, likely would have improved our molecular evaluation but were not available for testing. Experimental studies with captive raccoons would also be helpful in resolving

the role that these hosts play in the maintenance of agents of human ehrlichiosis in nature and their transmission to humans. We hope that this serosurvey will stimulate additional research into these topics.

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