

IMMUNOGLOBULIN RESPONSE OF THE ROCKY MOUNTAIN BIGHORN SHEEP

Authors: HUDSON, R., KITTS, W. D., and BANDY, P. J.

Source: Journal of Wildlife Diseases, 7(3) : 171-174

Published By: Wildlife Disease Association

URL: <https://doi.org/10.7589/0090-3558-7.3.171>

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at www.bioone.org/terms-of-use.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

IMMUNOGLOBULIN RESPONSE OF THE ROCKY MOUNTAIN BIGHORN SHEEP

R. HUDSON, W. D. KITTS, Department of Animal Science, University of British Columbia, Vancouver 8, B.C., and

P. J. BANDY, B.C. Fish and Wildlife Branch, Department of Recreation and Conservation, Victoria, B.C.

Abstract: The effects of individual variation, season and parasite activity on the levels of the four major immunoglobulins, IgM, IgA, IgG₁ and IgG₂, were analyzed in four adult Rocky Mountain bighorn ewes. Individual variation was significant for all immunoglobulin classes. Seasonal effects were detected in the levels of IgG₁. The fecal count of *Protostrongylus* larvae was inversely related to IgG₂ concentrations and the fecal *Nematodirus* count was inversely related to IgA. *Muellerius* and coccidia did not demonstrate significant relationships with circulating levels of any of the immunoglobulin classes.

INTRODUCTION

The structural characteristics of ruminant immunoglobulins have been intensively studied, but little is known about their response in disease states. Several reports on the levels of circulating immunoglobulins in domestic animals have appeared.^{6,7} However, they were confined to the quantitation of IgM and total IgG, possibly due to the technical difficulties encountered in the preparation of highly specific anti-immunoglobulin sera required for single radial immunodiffusion.

Recently, an immunoelectrophoretic technique was reported which permits the quantitation of IgM, IgG₁ and IgG₂ and the semiquantitative analysis of IgA.⁴ This method was used, in a preliminary study with four animals, to assess the possible influence of season, parasite activity and individual variability on the circulating levels of all four immunoglobulin classes in captive Rocky Mountain bighorn sheep.

METHODS

Four adult Rocky Mountain bighorn ewes (*Ovis canadensis*), captured in the East Kootenay region of British Columbia, were available for this study. Two

of these animals (Nos. 1 and 2) were taken from overstocked ranges south of Kootenay National Park just prior to a period of significant mortalities due to respiratory disease. The remaining two animals (Nos. 3 and 4) were captured on the Morrisey-Wigwam range where serious disease losses have not been reported recently. Animal No. 1 was studied for 2 years and No. 2, 3 and 4 were examined for a single year.

Immunoglobulin levels were determined for serum samples collected at two week to one month intervals using quantitative immunoelectrophoresis.⁴ Purified reference immunoglobulins, used for calculating concentrations, were measured assuming the following absorption characteristics:¹

$$0.1 \times E \frac{0.5 \text{ cm}}{280 \text{ nm}} = 139 \mu\text{g/ml}$$

Since a standard IgA preparation of satisfactory purity was not obtained, only relative rather than absolute changes of this immunoglobulin were measured.

Fecal parasite counts were conducted using a modified zinc flotation method² on single fecal samples as described previously.⁵

Analysis of covariance was conducted according to Harvey.⁸ Main effects were fit for individuals and seasons (January to June and July to December). Fecal parasite counts of *Muellerius*, *Protostrongylus*, *Nematodirus* and coccidia, quantitatively the major parasites present, were treated as covariables.

RESULTS AND DISCUSSION

Overall least squares means and individual arithmetic mean levels of IgM, IgG₁ and IgG₂ for each of the captive bighorn sheep are summarized in Table 1. Concentrations of IgM were only slightly less than those reported for man¹⁰ but were approximately one-third of levels reported for cattle.⁶ Total IgG levels (the sum of IgG₁ and IgG₂) were intermediate between values of 1290 mg/100 ml reported for cattle by Penhale and Christie⁷ and 2640 mg/100 ml reported by Klaus, Bennett and Jones.⁹ These differences may reflect either true species characteristics or the use of different Ig standards.

An examination of the correlation matrix in the least squares analysis disclosed that the levels of each of the immunoglobulin classes tended to be independent. The balance of increased immunoglobulin synthesis, following antigenic stimulation, and increased catabolism due to increased circulating levels did not appear to be uniform for all immunoglobulin classes.

Multiple regression techniques revealed significant individual effects for all immunoglobulin classes following the removal of variation due to season and parasite activity (Table 2). This was most marked in the mean concentration of IgG₂ exhibited by Animal No. 3, which was less than half of that found in the other bighorn ewes. The exact basis of these individual differences is not known. Since the animals were maintained on the same ration under similar conditions of management, environmental variables would not appear to be of major importance. Possibly the variation was due to differences in genetic regulation; however, further studies may reveal other influences such as age and early pathologic history.

Seasonal effects were significant only for the levels of IgG₁ in the animals studied (Table 2). Levels from January to June were significantly higher than during the rest of the year. This difference may be related to a shift in the relative rates of production and catabolism of IgG₁ although other factors such as changes in the distribution of body water may contribute to the observed effect.

Significant inverse relationships were detected between levels of IgA and fecal counts of *Nematodirus* and between IgG₂ and *Protostrongylus* (Table 2). Other correlations between circulating immunoglobulins and parasite activity were not apparent at the levels of infection experienced by the experimental animals (Table 3). The cause and effect nature

Table 1. Mean immunoglobulin levels (\pm SE) of captive bighorn sheep. All values are given in mg/100 ml.[†]

Animal No.	Year	IgM	IgG ₁	IgG ₂
No. 1	1968-69	62 \pm 5(9)	1551 \pm 239(9)	495 \pm 39(9)
	1969-70	84 \pm 8(10)	1347 \pm 16(10)	487 \pm 41(10)
No. 2	1968-69	67 \pm 11(14)	1906 \pm 128(14)	515 \pm 20(14)
No. 3	1969-70	99 \pm 12(12)	1529 \pm 251(12)	189 \pm 7(12)
No. 4	1969-70	142 \pm 19(10)	1901 \pm 74(10)	587 \pm 38(10)
Overall least squares means		94(55)	1703(55)	433(55)

[†] Number of values contributing to the means shown in parentheses.

Table 2. Analysis of the influence of individual variation, season and parasite activity on immunoglobulin levels of captive bighorn sheep.

Source of variation	df	F values			
		IgA	IgM	IgG ₁	IgG ₂
Main effects:					
Individuals	3	6.882 ^②	10.233 ^②	6.337 ^②	49.153 ^②
Seasons	1	1.047	0.325	9.269	3.899
Covariables:					
<i>Muellerius</i>	1	0.003	0.940	3.675	0.333
<i>Protostrongylus</i>	1	0.805	1.541	0.165	7.025 ^①
<i>Nematodirus</i>	1	6.059 ^①	0.312	0.228	0.437
Coccidia	1	0.011	0.318	0.817	4.341
Residual:	47				

① P < .05

② P < .01

Table 3. Parasite counts in fecal material collected from captive bighorn sheep. Values are expressed as the number of parasites per gram of air-dried feces.^①

Parasite Species	Mean \pm SE	Range
<i>Muellerius</i>	40.04 \pm 12.01	0-600
<i>Protostrongylus</i>	836.36 \pm 151.417	24-5,900
<i>Nematodirus</i>	15.57 \pm 4.710	0-186
<i>Coccidia</i>	59.21 \pm 16.321	0-1419

① Number of observations contributing to each mean = 55.

of the significant relationships is not known. Although an antibody response, reflected in immunoglobulin levels, may be responsible for controlling parasite activity, it is also possible that the parasite may exert an effect on the host which results in immunoglobulin depression. Recent studies have shown that parasites may have a marked effect on nitrogen balance and on immunologic

responsiveness,⁹ both of which may depress circulating immunoglobulins.

This study was based on four captive bighorn ewes and, therefore, conclusions must be considered of a tentative nature. However, the results encourage the use of immunoglobulin quantitation in further study of the host-parasite relationship.

Acknowledgements

The authors would like to thank Miss Anne Thomas and Mr. David Kitts for performing the fecal counts.

This study was supported, in part, by the National Research Council of Canada, Grant No. A132.

LITERATURE CITED

1. CHAMBERLAIN, M. A., C. G. SHAPLAND, and I. M. ROITT. 1970. Response to autologous IgG in patients with rheumatoid arthritis. *Ann. Rheum. Dis.* 29: 173-177.
2. FAUST, E. C., W. SAWITZ, J. TOBIE, C. ODOM, C. PERES, and D. R. LINCICOME. 1939. Comparative efficiency of various techniques for the diagnosis of protozoa and helminths in feces. *J. Parasitol.* 25: 241-262.
3. HARVEY, W. R. 1960. Least squares analysis of data with unequal sub-class numbers. Publ. ARS. 20-8. S.D.A. Beltsville, Md.
4. HUDSON, R. J., P. J. BANDY, and W. D. KITTS. 1970. Immunochemical quantitation of ovine immunoglobulins. *Amer. J. Vet. Res.* 31: 1231-1236.
5. ———, KITTS, W. D., and P. J. BANDY. 1970. Monitoring parasite activity and disease in the Rocky Mountain bighorn by electrophoresis of seromucoids. *J. Wildl. Dis.* 6: 104-106.
6. KLAUS, G. G. B., A. BENNETT, and E. W. JONES. 1969. A quantitative study of the transfer of colostral immunoglobulins to the newborn calf. *Immunology* 16: 293-299.
7. PENHALE, W. J., and G. CHRISTIE. 1969. Quantitative studies on bovine immunoglobulin. I. Adult plasma and colostrum levels. *Res. Vet. Sci.* 10: 493-501.
8. ROSS, J. G., J. R. TODD, and D. A. PURCELL. 1970. Nutritional balance trials with calves experimentally infected with *Trichostrongylus axei*. *Br. Vet. J.* 126: 393-400.
9. SVET-MOLDAVSKI, G. J., G. S. SHAGHIJAN, I. Y. CHERNYAKHOVSKAYA, D. M. MKHEIDZE, I. A. LITOVCHENKO, N. N. OZERETSKOVSKAYA, and Z. G. KADAGHIDZE. 1970. Inhibition of allograft rejection in *Trichinella*-infected mice. *Transplantation* 9: 69-70.
10. UFFELMAN, J. A., W. E. ENGELHARD, and C. R. JOLLIFF. 1970. Quantitation of immunoglobulins in normal children. *Clin. Chem. Acta* 28: 185-192.

Received for publication February 24, 1971