

# Leptospira interrogans SEROTYPES Ballum AND Grippotyphosa ISOLATED FROM THE MUSKRAT

Authors: PAUL, JOHN R., HANSON, LYLE E., SCHNURRENBERGER, PAUL R., and MARTIN, RUSSELL J.

Source: Journal of Wildlife Diseases, 8(1): 54-56

Published By: Wildlife Disease Association

URL: https://doi.org/10.7589/0090-3558-8.1.54

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at <a href="https://www.bioone.org/terms-of-use">www.bioone.org/terms-of-use</a>.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

## Leptospira interrogans SEROTYPES Ballum AND Grippotyphosa ISOLATED FROM THE MUSKRAT

JOHN R. PAUL  $^{\circ}$ , LYLE E. HANSON  $^{\circ}$ , PAUL R. SCHNURRENBERGER  $^{\circ}$ , and RUSSELL J. MARTIN  $^{\circ}$ 

- Illinois State Museum, Springfield, Illinois 62706
- University of Illinois, College of Veterinary Medicine
- 3 Illinois Department of Public Health, Springfield

Abstract: Leptospira interrogans serotypes ballum and grippotyphosa were isolated from muskrats (Ondatra zibethicus) taken in central Illinois. This represents the first reported isolation of ballum and the first United States record of grippotyphosa from this species.

#### INTRODUCTION

Muskrats have seldom been found infected with Leptospira; however, a single isolation of L. interrogans serotype icterrohemorrhagiae was made from several muskrats taken in southeastern Pennsylvania. Serotype grippotyphosa was reported from Czechoslovakia, Poland, and Bohemia and Russia. Serotype sejroe isolates have been reported from Poland and serotype kazachstanica II from Russia. This report describes the isolation of serotypes ballum and grippotyphosa from muskrats taken in central Illinois.

#### MATERIALS AND METHODS

The study was made on a private beef and swine feeding farm located approximately 11 km northwest of Springfield, Sangamon County, Illinois, during November, 1968. All animals reported here were taken from a pond of approximately 0.6 hectares surface area which was located near the upper limits of the drainage system of the farm properties. The muskrats were caught in steel traps which were checked daily in the early morning hours. Most animals were dead when removed from the traps.

Blood samples were taken from the heart using sterile disposable needles and syringes. Sera were tested by the microscopic agglutination (MA) technique using live antigens of the following serotypes of L. interrogans: autumnalis, balum, canicola, grippotyphosa, hardjoicterrohemorrhagiae and pomona. A titer of 1:100 or greater was considered to indicate infection.

Approximately ½ a kidney was collected asceptically from each muskrat and expressed through a sterile disposable syringe into a tube containing 4 ml of Ellinghausen-McCullough medium³, and 200 micrograms of 5 fluorouracil per ml.³ The cultures were incubated at 20 C and examined weekly under darkfield for 2 months before being declared negative.

Isolates were identified by use of cross agglutination tests with known antisera. Sera were tested from 14 muskrats. The first seven animals were collected before medium was available; as a result, only seven were cultured.

#### **RESULTS**

The results of serological and cultural tests are given in Table 1. Sera from six of the 14 animals reacted positively to grippotyphosa antigen. Muskrat SA 23, which was serologically negative, was culturally positive for grippotyphosa.

Muskrat SA 19 was serologically and culturally positive for ballum. Thus, 7 of the 14 sera and 2 of 7 kidney cultures were positive. The grippotyphosa isolate was confirmed by Dr. Aaron Alexander

of the Walter Reed Army Institute for Medical Research, Washington, D.C. The ballum isolate was not submitted for confirmation because of the laboratory's extensive experience with this serotype.

TABLE 1. Serologic and cultural results on fourteen Illinois muskrats tested for leptospires, November, 1968.

Number	Sex	Microscopic Agglutination Test	Kidney Cultures
SA 12	F	1:1000 L. grippotyphosa	Not cultured
SA 13	F	1:10,000 L. grippotyphosa	Not cultured
SA 14	M	Negative	Not cultured
SA 15	M	Negative	Not cultured
SA 16	F	Negative	Not cultured
SA 17	M	1:100 L. grippotyphosa	Not cultured
SA 18 ·	F	Negative	Not cultured
SA 19	F	1:100 L. ballum	L. ballum
SA 20	M	1:1000 L. grippotyphosa	Negative
SA 21	M	Negative	Negative
SA 22	M	1:100 L. grippotyphosa	Negative
SA 23	M	Negative	L. grippotyphosa
SA 27	M	1:10,000 L. grippotyphosa	Negative
SA 28	M	Negative	Negative

### **DISCUSSION**

Although L. interrogans serotype grippotyphosa has been reported from muskrats collected at various European and Asian sites, there has been no previous record of this serotype from muskrats in the United States. The discovery of leptospiral serotype ballum represents the first reported isolation of this serotype from muskrats.

Gillespie and Ryno' examined 14 muskrats from stream waters known to be contaminated with leptospiral serotype pomona from infected cattle. They postulated that the muskrat would have been a suitable species to reflect the presence of pathogenic leptospires in such contaminated waters, but concluded that the muskrat was resistant to water-borne infection by leptospires. Although the source of the leptospires was not determined in the present study, a water-borne source would seem to be the most logical explanation. Domestic animals penned

upstream and various wildlife species having access to the stream could have served as a source of the organisms. Because of the nature of known leptospiral infection mechanisms, i.e., the shedding of infected urine, maintenance of the organism in wet sites, and high prevalence of infection in rodents, it seems logical that the muskrat would be an animal that could provide an indication of the presence of such organisms in its environment.

The isolation of serotype grippotyphosa is of interest because of the recent increase of reports of the importance of this serotype for domestic animals. It appears that more detailed examination of a greater number of wildlife species may reveal this type to be quite common in Illinois animals. The isolation of serotype ballum is important as a new host record. Antibody titers to this serotype are found occasionally in domestic animals.

#### LITERATURE CITED

- ANAN'YIN, V. V., and E. V. KARASEVA. 1950. (Cited without reference by Shapiro, D. M., Leptospirosis of silver fox and muskrat at the Balkhash Fur Farm. From Natural Nidality of Disease & Questions of Parasitism. Univ. III. Press, 1968).
- CLARK, L. G., J. I. KRESSE, R. R. MARSHAK, and C. J. HOLLISTER. 1962. Natural occurrence of Leptospira icterohemorrhagiae infection in the common muskrat (Ondatra zibethicus zibethicus). A preliminary report. Cornell Vet. 52: 3-5.
- ELLINGHAUSEN, H. C., and W. G. McCULLOUGH. 1965. Nutrition of Leptospira pomona and growth of 13 other serotypes: Frostination of leic albumin complex and a medium of bovine albumin and polysorbate 80. Amer. J. Vet. Res. 26: 45-51.
- GILLESPIE, R. W. H., and J. RYNO. 1963. Leptespirosis in cattle may not spread to associated muskrats. Am. J. Vet. Res. 24: 634-637.
- 5. JOHNSON, R. C., and P. ROGERS. 1964. 5-fluorouracil as a selective agent for growth of leptospirae. J. Bact. 87: 422-426.
- KMETY, E., E. CHYLO, and I. PLESKO. 1957. Pripad ochorenia nu plonu horucku prenesenu z ondatry. Brat. Lek. Listy 371: 473. (Cited in Leptospiral Serotype Distribution Lists, Natl. Comm. Dis. Center, 1966).
- KREPKOGORSKAYA, T. A., and D. M. SHAPIRO. 1956. (Cited by Shapiro, D. M. Leptospirosis of silver fox & muskrat at the Balkhash Fur Farm. From Natural Nidality of Disease & Questions of Parasitism. Univ. III. Press, 1968).
- 8. PARNAS, J., and T. DOMBROWSKI. 1957. Travaux complexes sur les foyers naturals de tularemie, de leptospirose, et de quelques autres anthropozoonoses en Poland. Arch. Inst. Pasteur de Tunis 34: 351-380. (Cited by Reilly, J. R.: The Role of Predator Prey Relationships in the Nidality of Leptospirosis. Ph.D. Thesis, Univ. of Ill., Urbana, 302 pages, 1966.)
- POLORNY, B., F. LEHKY, Z. SEBEK, and J. VOSTA. 1958. Priodni ohniska a reservoiry leptospir v kotline trebonske v roce 1956. Sborn, Cs. aked. Zemedelsk Ved. 3: 263-270. (Cited by Reilly, J. R.: The Role of Predator-Prey Relationships in the Nidality of Leptospirosis. Ph.D. Thesis, Univ. of Ill., Urbana, 302 pages, 1966.)
- U.S. Livestock Sanitary Association. 1960. Report of the committee on leptospirosis. Proc. U.S. Livestock San. Assoc. 63: 140-142.
- 11. ZWIERZ, J., I. DURLAKOWA, K. KARMANSKU, J. ZWIERZCKOWSKI, K. LAZUGA, and A. KORCZYNSKA. 1958. Badania fauny w ogniskach epidemii leptospirozy w powiecie tomaszow lubelski. Med. weteryn 14: 647-657. (Cited by Reilly, J. R.: The Role of Predator-Prey Relationships in the Nidality of Leptospirosis. Ph.D. Thesis, Univ. of Ill., Urbana, 302 pages, 1966.)

Received for publication August 9, 1971