



Biology, Host Preferences, and Potential Distribution of *Calophya terebinthifolii* (Hemiptera: Calophyidae), a Candidate for Biological Control of Brazilian Peppertree, *Schinus terebinthifolia*, in Florida

Authors: Christ, Lindsey R., Cuda, James P., Overholt, William A., Vitorino, Marcelo D., and Mukherjee, Abhishek

Source: Florida Entomologist, 96(1) : 137-147

Published By: Florida Entomological Society

URL: <https://doi.org/10.1653/024.096.0118>

BioOne Complete (complete.bioone.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at www.bioone.org/terms-of-use.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

BIOLOGY, HOST PREFERENCES, AND POTENTIAL DISTRIBUTION
OF *CALOPHYA TEREBINTHIFOLII* (HEMIPTERA: CALOPHYIDAE), A
CANDIDATE FOR BIOLOGICAL CONTROL OF BRAZILIAN PEPPERTREE,
SCHINUS TEREBINTHIFOLIA, IN FLORIDA

LINDSEY R. CHRIST¹, JAMES P. CUDA^{1,*}, WILLIAM A. OVERHOLT², MARCELO D. VITORINO³ AND ABHISHEK MUKHERJEE⁴

¹University of Florida, Department of Entomology and Nematology, P. O. Box 110620
Gainesville, FL 32611-0620, USA

²University of Florida, Biological Control Research and Containment Laboratory, 2199 South Rock Road,
Fort Pierce, FL 34945-3138, USA

³Universidade Regional de Blumenau, Rua Antonio da Veiga, 140-Victor Konder 89012-900-Blumenau,
SC, Brazil

⁴Texas A&M University, 2475 TAMU, College Station, TX 77843

*Corresponding author; E-mail: jcuda@ufl.edu

A pdf file with supplementary material for this article in Florida Entomologist 96(1) (2013) is online at
<http://purl.fcla.edu/fcla/entomologist/browse>

ABSTRACT

Brazilian peppertree, *Schinus terebinthifolia* Raddi (Anacardiaceae), a perennial woody plant native to Brazil, Argentina, Uruguay and Paraguay, has become one of the most invasive weeds in Florida. A leaflet pit galling psyllid, *Calophya terebinthifolii* Burckhardt & Basset (Hemiptera: Calophyidae), has been identified as a potential biological control agent for Brazilian peppertree. However, biological information on the psyllid, including its life history, rearing procedures and potential distribution, is lacking. This type of information is essential when importing an insect for biological control purposes. From May-Aug 2009, field and laboratory research was conducted in Gaspar, Santa Catarina, Brazil with psyllids collected from the Atlantic coastal region of Santa Catarina. Laboratory studies on the psyllid in Brazil focused on: female fecundity (55.3 ± 8.9 eggs/female), the number and size of the immature stages, age-specific survivorship, and mean generation time (43.7 ± 1.2 days). Preliminary evidence from feeding trials suggests this psyllid from the Atlantic coastal region of Santa Catarina is locally adapted to Brazilian peppertree plants of haplotype A, which is one of the genetic types that invaded Florida. Ecological niche modelling with MaxEnt confirmed there was climatic overlap between Florida and the native range of the psyllid in South America. Using collection and survey locations of the psyllid in its native range and point locations for haplotype A plants in Florida, a map was created that predicted Volusia, coastal Pasco and Hernando counties, and a small section of southwestern Polk county as suitable locations for establishment of the psyllid if it is approved for release as a biocontrol agent.

Key Words: weed biocontrol, psyllid, life history, niche modeling

RESUMEN

Brazilian peppertree, *Schinus terebinthifolia* Raddi (Anacardiaceae), un árbol perenne nativo de Brasil, Argentina, Paraguay y Uruguay, es una de las plantas más invasoras en la Florida. El psílido formador de agallas en las hojas, *Calyophya terebinthifolii* Burckhardt & Basset (Hemiptera: Calophyidae), ha sido identificado como un agente potencial de control biológico de Brazilian peppertree. Sin embargo, información biológica sobre el psílido, incluyendo su historial de vida, procedimientos de crianza y distribución potencial no existe. Este tipo de información es esencial cuando se importa un insecto con propósitos de control biológico. De mayo a agosto del 2009, investigaciones de campo y en laboratorio fueron llevadas a cabo en Gaspar, estado de Santa Catarina, Brasil con insectos colectados en la región costera atlántica de Santa Catarina. Los estudios de laboratorio con el psílido en Brasil se concentraron en la fecundidad de las hembras (55.3 ± 8.9 huevos/hembra), el número y tamaño de los estadios inmaduros, sobrevivencia a diferentes edades, y duración promedio de una generación (43.7 ± 1.2 días). Resultados preliminares de los ensayos de alimentación indican que los psíidos originarios de la región costera atlántica de Santa Catarina están adaptados localmente a plantas de Brazilian peppertree haplotipo A, el cual es uno de los tipos genéticos que ha invadido la Florida. El modelo de nicho ecológico utilizando 'MaxEnt' confirmó

que existe una similitud climática entre la Florida y el rango nativo del insecto en América del Sur. Utilizando los datos de las localidades donde se colectó el insecto en su rango nativo y las localidades del haplotipo A en la Florida, se generó un mapa que predijo el condado de Volusia, la región costera de Pasco, el condado de Hernando, y una pequeña sección del suroeste del condado de Polk como localidades adecuadas para el establecimiento del psílido si éste llega a ser aprobado para ser liberado como agente de control biológico.

Palabras Clave: control biológico de malezas, psílicos, historia natural, modelo del nicho

Brazilian peppertree, *Schinus terebinthifolia* Raddi (Sapindales: Anacardiaceae) is a perennial woody shrub/tree native to Brazil, Argentina, Uruguay and Paraguay (Barkley 1944, 1957). It was originally introduced to Florida in the 1840s as an ornamental (Mack 1991), but has since naturalized becoming an invasive weed occupying more than 283,000 ha in central and south Florida (Cuda et al. 2006; Rogers et al. 2012) and is expanding its range northward (Anon. 2007a,b; Mukherjee et al. 2012). Brazilian peppertree having numerous invasive properties such as fast growth, prolific seed production, tolerance to shade, fire, drought, salinity, and vigorous resprouting (Ewel 1979; Nilsen & Muller 1980; Doren et al. 1991; Ewe & Sternberg 2005), displaces native vegetation and reduces the biodiversity of native plants and animals (Bennett et al. 1990). Furthermore, Brazilian peppertree populations in Florida are the result of 2 separate introductions from different parts of the native range resulting in the establishment of 2 cpDNA haplotypes (A and B) and their hybrids (Williams et al. 2005).

In 2011, the South Florida Water Management District spent almost \$1.8 million controlling Brazilian peppertree (Rodgers et al. 2012). Because of its invasive characteristics and resistance to conventional control methods, Brazilian peppertree was targeted for classical biological control in Florida in the late 1970s (Delfosse 1979; Campbell et al. 1980). The lack of native congeners minimizes the risk of damage to non-target plants from introduced natural enemies, making it a good candidate for biological control (Pemberton 2000). Biological control of Brazilian peppertree could provide a more cost effective and sustainable approach for managing this invasive weed, especially when it is integrated with conventional control methods (Cuda et al. 2006). Several insect biological control agents have been studied for release in Florida (e.g., Medal et al. 1999; Martin et al. 2004; Cuda et al. 2005, 2008, 2009; Manrique et al. 2008; Moeri et al. 2009; Mound et al. 2010; Wheeler et al. 2010; Oleiro et al. 2011; McKay et al. 2012; Rendon et al. 2012). *Calophya terebinthifolii* Burckhardt & Basset (Hemiptera: Calophyidae), a leaflet galling psyllid of Brazilian peppertree (Vitorino et al. 2011), is the focus of this study.

Because local climatic conditions can play an important role in the distribution and effectiveness of a weed biocontrol agent (McClay & Hughes 1995; Julien 2000; Robertson et al. 2008), matching ecoclimatic conditions for optimal development of *C. terebinthifolii* in its native range is likely to increase the likelihood of its establishment post-release (Geiger & Gutierrez 2000). Ecological niche modelling is a tool for predicting the fundamental niche of a species (where a species could occur) by establishing a statistical relationship between species known occurrences and relevant predictor variables (for example, climatic variables) (Phillips et al. 2006). Several methods have been developed to model climate based species distributions (reviewed in Franklin 2009). The Maximum Entropy Species Distribution Model (MaxEnt) is a presence only modelling approach that predicts the ecological niche of a species using known occurrence data (latitude/longitude point locations of the species) and environmental layers (Phillips et al. 2006). Using the input data for *C. terebinthifolii*, MaxEnt can generate a prediction of geographical locations climatically suitable for the psyllid to establish persistent populations in Florida.

The objectives of this research were to accelerate testing of *C. terebinthifolii* as a biological control agent for Brazilian peppertree in Florida by 1) investigating the biology of *C. terebinthifolii* in Brazil (fecundity, number/size of instars, and survivorship); 2) comparing the performance of *C. terebinthifolii* on the genotypes of Brazilian peppertree that occur in Florida; and 3) using ecological niche modelling to identify potential release areas in Florida that overlap climatically with the insect's native distribution in South America and locating additional sites in Brazil for collecting *C. terebinthifolii* adapted to Florida Brazilian peppertree genotypes.

MATERIALS AND METHODS

The study was conducted from 11 May-8 Aug 2009 in Gaspar, Santa Catarina, Brazil at the Laboratório de Monitoramento e Proteção Florestal (LAMPF) (S -26.9110° W -48.9362°). LAMPF is affiliated with the Forest Protection and Surveillance Laboratory that is part of the Universidade Regional de Blumenau.

Fecundity, Instar Verification, Size, and Survivorship

A colony of psyllids was established in the laboratory by collecting individuals from the field. Geoposition, corresponding weather conditions (i.e., relative humidity, temperature, wind speed, and precipitation), site description, and number of psyllids collected were recorded for each site. Adults were captured individually in clear gelatin capsules (size "000") from Brazilian peppertrees located along the eastern coast of Santa Catarina, Brazil. The gelatin capsules containing adult psyllids were stored in coolers, and then transported back to LAMPF within 3-5 h of capture. Field collected adults were then released on Brazilian peppertree seedlings. The seedlings used in the studies were grown in 1 liter nursery pots under ambient conditions in a greenhouse at LAMPF and were all approximately 40 cm in height with at least 3 flushes of new leaflets.

To measure fecundity, 1 female and 1 male (newly emerged from the established colony) were released on a Brazilian peppertree seedling ($n = 28$) covered with a 30×60 cm mesh bag and secured at the base of the seedling stem with 2 vinyl-coated twist ties. Leaves were checked daily for eggs. Temperature and relative humidity also were recorded. Female wing length (mm), body length (mm), and head width (mm) were measured at death. Fecundity was regressed against female wing length to examine the relationship between egg production and body size (Geiger & Gutierrez 2000).

To verify the number of instars from laboratory reared ($n = 18$) and field collected nymphs ($n = 177$), the following parameters were measured under a dissecting microscope: body length, body width, and head capsule width (mm). Based on previous studies, this sample size of 195 nymphs was adequate for generating a histogram of instars and size range of each instar for cecidogenous psyllids (Devi & Prabhoo 1995; Alvarez-Zagoya & Cibrian-Tovar 1999). Measurements were compared to those of *Calophya schini* Tuthill recorded by Alvarez-Zagoya and Cibrian-Tovar (1999) as a point of reference and comparison between the 2 congeners.

Using the eggs produced from the female fecundity study, egg cohorts were followed through their development with survivorship and temperature recorded daily. Psyllids were allowed to complete their development on a Brazilian peppertree seedling using the same procedure described for the aforementioned fecundity study. Psyllid eggs ($n = 157$) were kept in environmental growth chambers at 21.9 ± 2.6 °C; RH, 67.9 \pm 7.6%; and 16:8 h (L:D).

Data were analyzed with SAS© software, Version 9.0 (SAS 2002). Sample means \pm SEM were determined for all parameters. After confirming that the data were normally distributed, a linear

regression was performed to correlate fecundity with female size. A frequency distribution was created to determine number of instars. A significance level of $\alpha = 0.05$ was used for all statistical analyses. The means of body length of male and female adult *C. terebinthifolii* and of *C. terebinthifolii* and *C. schini* were statistically compared with t-tests using Proc t-test (SAS 2002). A survivorship curve of the laboratory population was produced and a life table was constructed. Population statistics were calculated using standard techniques (Price 1997).

Performance on Brazilian Peppertree Haplotypes Established in Florida

To test the hypothesis that *C. terebinthifolii* performs better on natal genotypes of Brazilian peppertree, terminal buds were taken from laboratory plants ($n = 22$) in Brazil and preserved in silica gel. Chloroplast haplotype was determined by D. A. Williams, Texas Christian University, Ft. Worth, Texas. Sampled plants were categorized as either a success (females laid eggs, instars developed and emerged as adults) or failure (no eggs were laid or psyllids failed to develop into adults). Categorical data were analyzed with the *G*-test of independence (Sokal & Rohlf 1995) at $\alpha = 0.05$.

Ecological Niche Modeling

Two occurrence datasets were used for the ecological niche modelling. The first dataset included the geolocations of *C. terebinthifolii* in its native South American range ($n = 47$) with points obtained from surveys by Ms. Lindsey Christ, Dr. Greg Wheeler and Mr. Fernando McKay (personal communication), and from Burckhardt & Basset (2000). The second dataset included 50 point locations of Brazilian peppertree (haplotype A) found in Florida provided by Dr. William Overholt (personal communication). Two bioclimatic (bioclim) variables obtained from the WORLDCLIM database (<http://www.worldclim.org>) were used to model the insect's fundamental niche. The climate layers were generated using average climate data collected from weather stations worldwide (Hijmans et al. 2005). ESRI 2.5 arc-minute resolution grids of current climate data were used to run the model (Environmental Systems Research Institute, Inc., Redlands, California). The 2 bioclim variables included were bio14 (precipitation during the driest month) and bio4 (temperature seasonality-standard deviation of monthly temperature).

Data partitioning is a technique used to provide test points for model verification and accuracy (Phillips et al. 2006). Both niche models generated in MaxEnt were statistically verified by running 10 random partitions of the dataset with 80% of the points as training data to generate the

models and the remaining 20% as independent test data for extrinsic verification of predictive accuracy. Each training set was run in MaxEnt with the random seeded sub-sampling procedure using 10 replications with 10% as random test percentage for intrinsic model testing. A minimum training threshold dependent one-tailed binomial test was conducted to verify the models generated by the MaxEnt predictions performed significantly better than random (Phillips et al. 2006). The test was based on omission rate (test points falling in pixels predicted as not suitable) and Fractional Predictive Area (FPA, fraction of pixels predicted suitable) (Phillips et al. 2006). Because MaxEnt generates continuous predictions, the minimum training threshold (minimum value received by any training data) was used to perform the threshold dependent binomial test because the binomial test requires the data to be binary (presence / absence), not continuous. The proportion of test points (n_{test}) predicted not suitable (out_{test}) was used as the extrinsic omission rate ($\text{out}_{\text{test}}/n_{\text{test}}$) (Anderson et al. 2003). The binomial test was performed using Proc Freq in SAS© 9.0 (SAS 2002).

MaxEnt used presence data and randomly generated “pseudo-absences” from the study area (also referred to as model background) to generate the predicted distribution. For native occurrences, all of South America was used as the background; for predictions using Florida occurrences of Brazilian peppertree haplotype A, Florida was used as the background. A prediction map showing the potential distribution of *C. terebinthifolii* in Florida was developed by combining the predictions generated using 10 randomly partitioned datasets.

To identify areas in Florida climatically similar to the native distribution of the psyllid, predictions generated using the 2 occurrence datasets were overlapped. The overlapped distribution was classified into 4 major categories: 1) not suitable (absent), 2) suitable based on *C. terebinthifolii* occurrence data from South America, 3) suitable based on Florida distribution of Brazilian peppertree type A plants, and 4) overlap between 2 and 3.

RESULTS AND DISCUSSION

Fecundity, Instar Verification, Size, and Survivorship

In total, 18 of 28 females (64.3%) in cages containing mating pairs of adult *C. terebinthifolii* deposited eggs. Mean number of eggs laid per female was 55.3 ± 8.9 ($n = 18$) with a range of 16-139 eggs (excluding the 10 females that did not lay eggs). These values are similar to those reported for *C. schini*; the average number of eggs laid for this species was 50 (no SEM was given) with a range of 25-110 eggs based on 10 ovipos-

iting females (Alvarez-Zagoya & Cibrian-Tovar 1999). The oblong shaped eggs of *C. terebinthifolii* are 0.212 ± 0.002 mm in length (Table 1) and are a milky, translucent cream color when first laid then turn a black, iridescent color within 24 h. Similar to *C. schini* (Downer et al. 1988), the females laid their eggs on the new leaf flush along the leaflet margins, midribs, leaflet petioles, and leaf buds. The number of eggs laid by a female increased with wing length ($F = 5.15$; $df = 9$; $P = 0.0494$; Fig. 1). The low R^2 value (0.364) reflected the high variability in the data. In total, 11 of the 18 females that laid eggs (61.1%) were recovered for measurement.

To determine the number of instars, a histogram of body length was constructed and examined for distinct peaks. The histogram (Fig. 2) confirmed 5 instars for *C. terebinthifolii*. Automontage images of the different instars and adults are shown in Fig. 3 in black and white and in color in Suppl. Fig. 3. The measurements of all life stages of *C. terebinthifolii* (body length, body width, and head capsule width) are provided in Table 1 and were compared with its congener *C. schini* in Fig. 4. Female body lengths for *C. terebinthifolii* were significantly larger than males ($t = 3.65$; $df = 38$; $P = 0.008$), which is typical for most psyllids (Hodkinson 2009). By comparison, *C. schini* is significantly larger than *C. terebinthifolii* in the following stages: egg ($t = 10.85$; $df = 52$; $P < 0.0001$), 3rd instar ($t = 3.62$; $df = 42$; $P = 0.0008$), 5th instar ($t = 7.63$; $df = 157$; $P < 0.0001$), and adult ($t = 3.54$; $df = 68$; $P = 0.0007$).

A survivorship curve (Fig. 5) was developed using daily observations on survivorship from 4 of the cages ($n = 157$ eggs) held in the environmental growth chambers. Eclosion of 1st instars commenced on day 9, and there was a sharp decrease in survival during the first wk after hatching when the nymphs are in the vulnerable crawler stage. Survivorship leveled off after the crawlers settled into pit galls and then survival gradually decreased during the 3rd, 4th, and 5th instars until adults began to emerge around day 43. The average duration from egg to adult stage was 43.7 ± 1.2 days, which was similar to the 43 days reported for *C. schini* (Alvarez-Zagoya & Cibrian-Tovar 1999).

Performance on Brazilian Peppertree Haplotypes Established in Florida

The results from the DNA analysis are shown in Table 2. From this test sample, psyllids only developed to adults on haplotype A and haplotype O plants, with the best performance on haplotype A plants. A G -test comparing all other haplotypes (O, K, D, and M), with haplotype A plants showed the psyllids performed significantly better on A than other haplotypes ($G = 7.63$; $P < 0.01$). Recent studies on the Brazilian peppertree invasion

TABLE 1. DIMENSIONS OF ALL LIFE STAGES OF THE LEAFLET GALLING PSYLLID *CALOPHYA TEREBINTHIFOLII*.

Stage	n	$\bar{x} \pm \text{SEM}$ (mm)
Egg	30	0.212 \pm 0.002 length 0.099 \pm 0.002 width
1st instar	18	0.228 \pm 0.005 length 0.183 \pm 0.012 width 0.108 \pm 0.004 head capsule width
2nd instar	67	0.307 \pm 0.002 length 0.298 \pm 0.002 width 0.155 \pm 0.002 head capsule width
3rd instar	14	0.446 \pm 0.010 length 0.442 \pm 0.009 width 0.222 \pm 0.006 head capsule width
4th instar	24	0.674 \pm 0.015 length 0.665 \pm 0.015 width 0.320 \pm 0.009 head capsule width
5th instar	72	0.962 \pm 0.006 length 0.942 \pm 0.006 width 0.398 \pm 0.002 head capsule width
Adult	40	1.251 \pm 0.020 body length 1.509 \pm 0.018 wing length 0.441 \pm 0.007 head capsule width
Female	28	1.293 \pm 0.021 body length 1.542 \pm 0.020 wing length 0.453 \pm 0.008 head capsule width
Male	12	1.154 \pm 0.013 body length 1.432 \pm 0.025 wing length 0.415 \pm 0.009 head capsule width

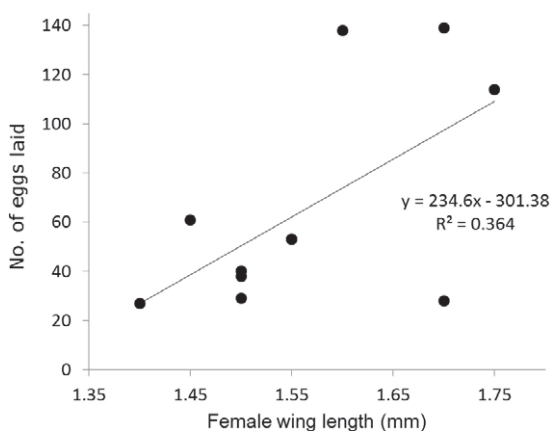


Fig. 1. The relationship between wing length and laboratory fecundity of female *Calophya terebinthifolii* ($n = 11$).

in Florida (Geiger et al. 2011; Mukherjee et al. 2012) and 'fine scale' adaptation by several of its natural enemies (Cuda et al. 2012) suggest that

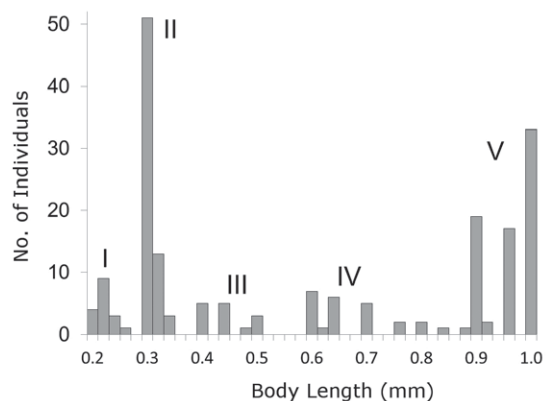


Fig. 2. Frequency distribution of nymphal length measurements of *Calophya terebinthifolii*. Peaks within groupings indicate 5 instars.

it may be necessary to introduce multiple biotype-specific control agents into Florida to achieve the desired impact.

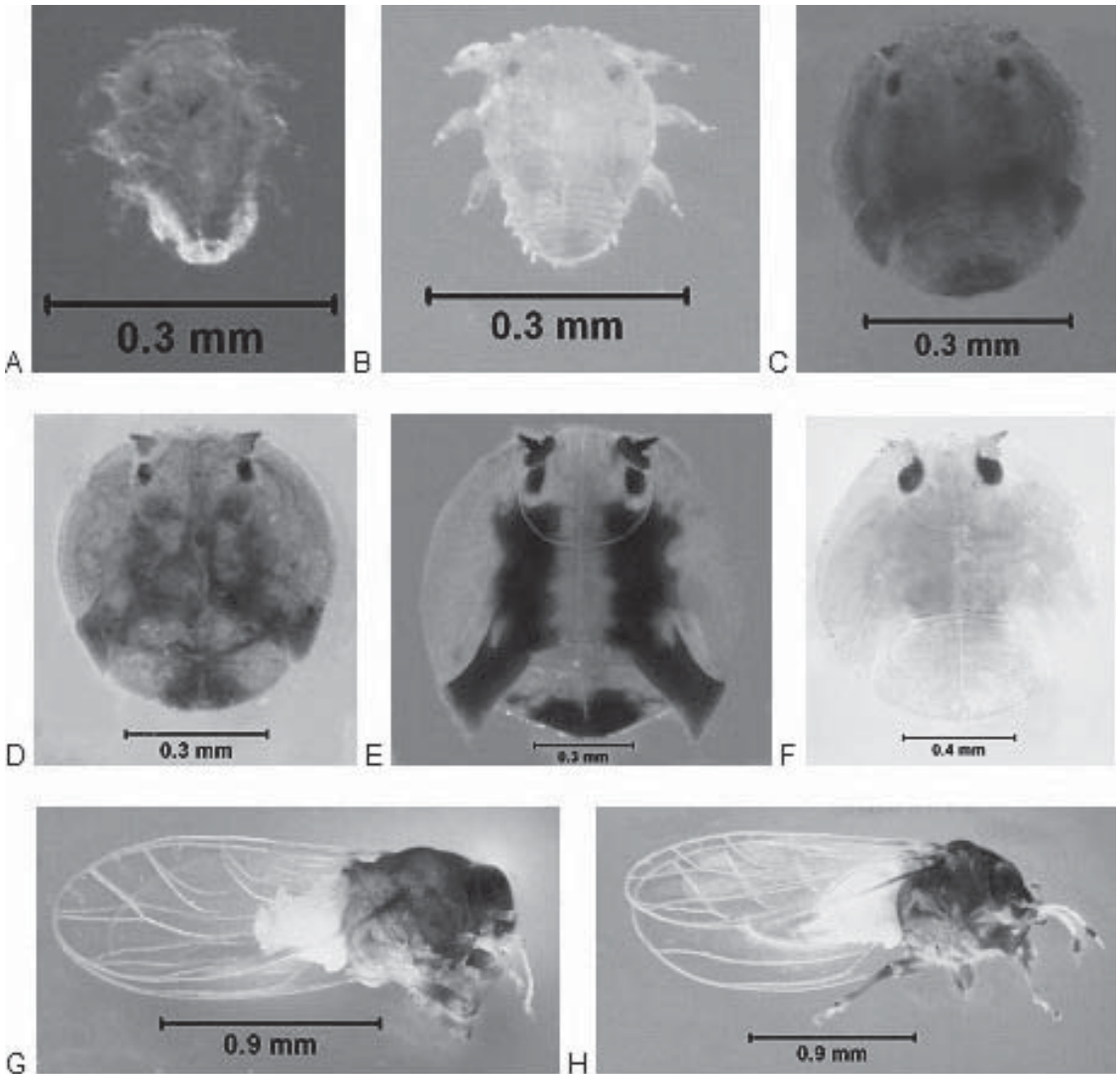


Fig. 3. Automontage images of *Calophya terebinthifolii*. A) 1st instar, B) 2nd instar, C) 3rd instar, D) 4th instar, E) 5th instar, F) 5th instar prior to darkening, G) adult male, and H) adult female. Photo credit L. R. Christ.

Ecological Niche Modelling

Tropical psyllids are especially vulnerable to cold temperatures and drought (Hodkinson 2009). Waring & Cobb (1992) reported that in 73% of published studies, gall-forming species reacted negatively to drought. Young nymphs of several species of gall-inducing psyllids often suffer high mortality before gall formation due to low humidity (Ferreira et al. 1990; personal observation). Because *C. terebinthifolii* reproduces continuously in Brazil, precipitation during the driest month (bio14) is a critical factor to include in the model as reflected by this variable's overall contribution to the model prediction. It is noteworthy

that Weins et al. (2006) found that temperature seasonality (bio4) was an important climate variable for predicting the distribution of tropical tree frogs in temperate North America.

The minimum training threshold dependent binomial test results for Florida (validated using Florida type A Brazilian peppertree points) were highly significant for all the data partitions (Table 3; $z = 3.1623$; $P = 0.0008$). The average FPA (Fractional Predictive Area) was 0.448. The extrinsic omission rates were zero for all partitions. Likewise, the results for the South American tests were similar (Table 4; $z = 2.5-3.2$; $P < 0.0001$), with all data partitions being highly significant. The average FPA for the South American

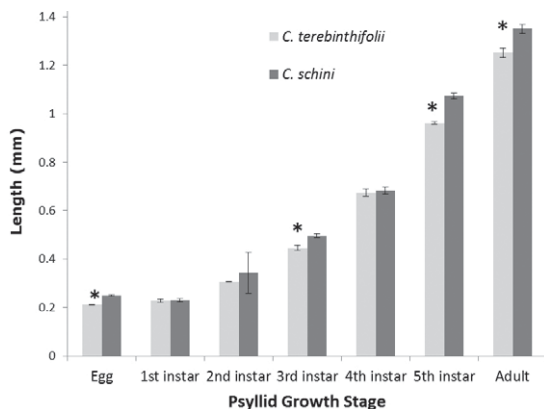


Fig. 4. Comparison of the body lengths of the life stages of *Calophya terebinthifolii* and *C. schini*. Measurements for *C. schini* obtained from Alvarez-Zagoya & Cibrian-Tovar (1999).

tests was 0.173. The extrinsic omission rates were slightly higher with an average of 4%. Overall, the models generated by MaxEnt were accurate with predictions being significantly better than random.

The predicted distribution of *C. terebinthifolii* in Florida is shown in Fig. 6 in black and white and in color in Suppl. Fig. 6. Establishment of *C. terebinthifolii* is expected to occur in a few coastal regions along the panhandle, west coast, and along the east coast in Florida. Areas of overlap where Brazilian peppertree haplotype A plants were known to occur and predicted to be suitable for establishment of *C. terebinthifolii* included the following Florida counties: Volusia, coastal Pasco and Hernando, and a small section of southwestern Polk. These counties should be targeted for planned releases of *C. terebinthifolii* if it is ap-

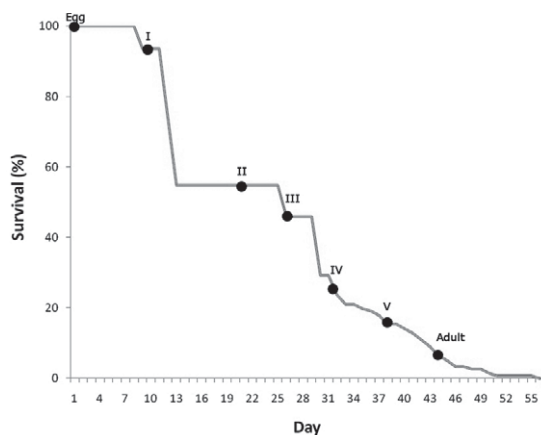


Fig. 5. Survivorship curve of *Calophya terebinthifolii* in laboratory environmental growth chambers in Brazil at 21.9 ± 2.6 °C, RH $67.9 \pm 7.6\%$ and 16:8 h L:D.

proved for release as a biological control agent of Brazilian peppertree. It is noteworthy that the psyllid is predicted to establish in Franklin Co. in the panhandle where Brazilian peppertree was recently discovered (EDDMaps 2012).

Using the known locations of *C. terebinthifolii*, another map was generated in MaxEnt to predict other areas in South America likely to support psyllid populations. The map for South America (available upon request) predicted *C. terebinthifolii* to occur in the following countries: 1) southern Chile, 2) central Bolivia, 3) southeastern Paraguay, 4) Uruguay, and 5) southern Brazil in the states of Rio Grande do Sul, Santa Catarina, Paraná, and coastal São Paulo. These areas (except for Chile and Bolivia where Brazilian peppertree has not been reported) would be good locations to survey for *C. terebinthifolii* because the full native range for the psyllid is still unknown.

CONCLUSIONS

Results of this research provide baseline biological data for evaluating *C. terebinthifolii* as a candidate for biological control of Brazilian peppertree. The 5 instars observed for *C. terebinthifolii* in this study differed from the number reported by Downer et al. (1988) for *C. schini* in California. They observed this congener had only 4 distinct peaks after measuring 1,000 field-collected nymphs in all stages of development over a 10 wk period. More recently, Alvarez-Zagoya & Cibrian-Tovar (1999), who studied *C. schini* in Mexico, found evidence of 5 instars when nymphs were studied in a laboratory setting. It is conceivable that Downer et al. (1988) focused their field sampling efforts on nymphs that had already initiated gall formation, and simply overlooked the first instar crawler stage because of its small size and mobility (Table 1).

Three different studies on both species of *Calophya*; *C. schini* in Chapingo, Mexico (Alvarez-Zagoya & Cibrian-Tovar 1999), *C. schini* in Ventura County, California (Downer et al. 1988), and *C. terebinthifolii* in Santa Catarina, Brazil (Vitorino et al. 2011; personal observation) found the insects to have continuous generations. The polyvoltinism exhibited by *Calophya* psyllids makes them good candidates for biological control because they can provide year round control. Although not gall-forming, other psyllids such as *Boreioglycaspis melaleuca* Moore and *Heteropsylla spinulosa* Muddiman, Hodkinson and Hollis have been successfully used as biological control agents (Kuniata & Korowi 2004; Rayamajhi et al. 2007; Center 2007; Rayamajhi et al. 2008). Additionally, *C. terebinthifolii* may reach higher densities in Florida than Brazil because of escape from specialist parasitoids (Christ 2010).

Brazilian peppertree haplotypes suitable for psyllid development were unknown prior to the

TABLE 2. REARING OF *CALOPHYA TEREBINTHIFOLII* ON 5 DIFFERENT BRAZILIAN PEPPERTREE HAPLOTYPES AT LAMPF, SANTA CATARINA, BRAZIL, MAY-AUG 2009. REARING SUCCESS EXPRESSED AS A PERCENTAGE OF THE TOTAL NUMBER OF PLANTS WITHIN EACH HAPLOTYPE PRODUCING NORMAL ADULTS.

Brazilian peppertree cpDNA haplotype	N	Success	Failure	% Success
A	12	8	4	67%
O	5	1	4	20%
K	2	0	2	0%
D	2	0	2	0%
M	1	0	1	0%

TABLE 3. MINIMUM TRAINING THRESHOLD DEPENDENT BINOMIAL TEST RESULTS OF OMISSION FOR PREDICTING THE DISTRIBUTION OF THE LEAFLET GALLING PSYLLID *CALOPHYA TEREBINTHIFOLII* IN FLORIDA WITH MAXENT.

Data Partition	Fractional Predictive Area	Points Not Suitable (out _{test})	Test Points (n _{test})	Extrinsic Omission Rate (out _{test} /n _{test})	Z
Part - 1	0.406	0	10	0	3.1623**
Part - 2	0.410	0	10	0	3.1623**
Part - 3	0.356	0	10	0	3.1623**
Part - 4	0.475	0	10	0	3.1623**
Part - 5	0.463	0	10	0	3.1623**
Part - 6	0.517	0	10	0	3.1623**
Part - 7	0.419	0	10	0	3.1623**
Part - 8	0.506	0	10	0	3.1623**
Part - 9	0.413	0	10	0	3.1623**
Part - 10	0.510	0	10	0	3.1623**
Average	0.448	0		0	

** $P = 0.0008$

TABLE 4. MINIMUM TRAINING THRESHOLD DEPENDENT BINOMIAL TEST RESULTS OF OMISSION FOR PREDICTING THE DISTRIBUTION OF THE LEAFLET GALLING PSYLLID *CALOPHYA TEREBINTHIFOLII* IN SOUTH AMERICA WITH MAXENT.

Data Partition	Fractional Predictive Area	Points Not Suitable (out _{test})	Test Points (n _{test})	Extrinsic Omission Rate (out _{test} /n _{test})	Z
Part - 1	0.222	0	10	0.0	3.1623**
Part - 2	0.058	1	10	0.1	2.5298**
Part - 3	0.078	1	10	0.1	2.5298**
Part - 4	0.232	0	10	0.0	3.1623**
Part - 5	0.237	0	10	0.0	3.1623**
Part - 6	0.255	0	10	0.0	3.1623**
Part - 7	0.260	0	10	0.0	3.1623**
Part - 8	0.077	1	10	0.1	2.5298**
Part - 9	0.233	0	10	0.0	3.1623**
Part - 10	0.075	1	10	0.1	2.5298**
Average	0.173	0.400		0.04	

** $P < 0.0001$

laboratory rearing experiments in Brazil. Few replicates were used for some of the haplotypes (K, D, and M) and no type B plants were included, making it difficult to draw clear conclusions regarding the possible suitability of Florida genotype plants. However, it is noteworthy that type O

is most closely related to haplotype B (Mukherjee et al. 2012). If *C. terebinthifolii* is indeed locally adapted to haplotype A, it may have difficulty developing on the novel genotypes in Florida created by the hybridization of haplotypes A and B. In Brazil, these 2 haplotypes do not occur sym-

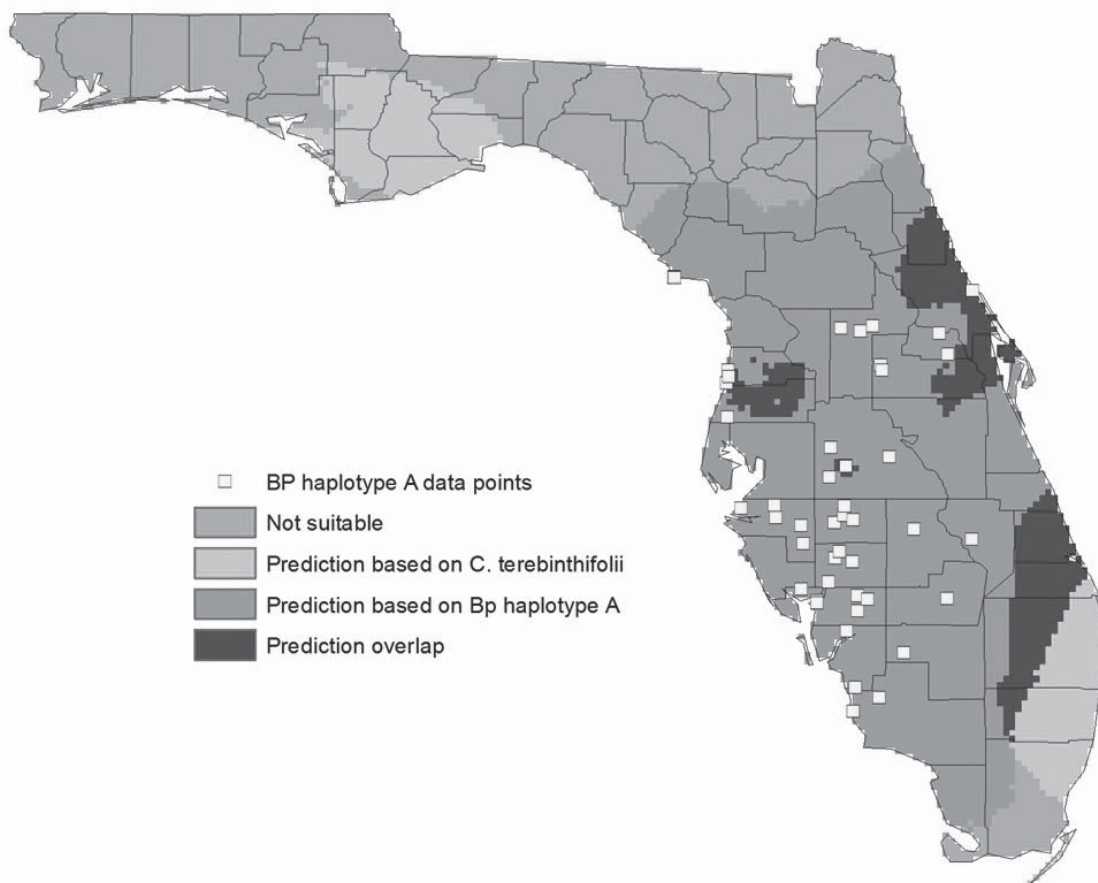


Fig. 6. Map of predicted climatic suitability for the leaflet galling psyllid *Calophya terebinthifolii*, its host plant Brazilian peppertree haplotype A, and their predicted overlap in Florida. Open squares ($n = 50$) indicate locations of Brazilian peppertrees identified as haplotype A through DNA analysis (D. A. Williams, unpubl. data).

patrically (Williams et al. 2005; Mukherjee et al. 2012)). However, if *C. terebinthifolii* from coastal Santa Catarina, Brazil can develop on both haplotypes A and O, it may accept haplotype B as well. Because our preliminary data suggest that *C. terebinthifolii* is locally adapted to specific Brazilian peppertree genotypes, evidence of 'fine scale' adaptation to its host plant is consistent with other small natural enemies of Brazilian peppertree having limited dispersal capability (see Cuda et al. 2012).

The 2 predicted distribution maps produced in this study were constrained by the paucity of data points for *C. terebinthifolii* in its native range. Consequently, the maps generated should be considered preliminary. One of the assumptions made when generating niche models is that the entire native range of the species is known and is accounted for in the point locations (Phillips et al. 2006). However, the complete native range of *C. terebinthifolii* is still unknown. As more information about the native range distribution of the

psyllid becomes available, a more accurate model can be developed. Nevertheless, these preliminary models are reasonable for selecting locations for future surveys in South America and potential habitats in Florida where the psyllid could be released as a biocontrol agent for Brazilian peppertree.

Finally, the most compelling argument for using *C. terebinthifolii* as a biological control agent for Brazilian peppertree in Florida is based on the discovery of the adventive *C. schini* in California in the mid 1980s (Downer et al. 1988). *Calophya schini*, which was first reported from Los Angeles County, California in Jul 1984, dispersed rapidly from San Diego County to the San Francisco Bay (a distance of about 800 km) in less than 4 yr (Downer et al. 1988). This congener of *C. terebinthifolii* caused extensive damage on Peruvian peppertree, *Schinus molle* L. In southern California where both *Schinus* species co-occur, Downer et al. (1988) reported that *C. schini* attacked only Peruvian peppertree but not its close relative

Brazilian peppertree. Assuming *C. terebinthifolii* would respond similarly if it were introduced into Florida without its natural enemies, this leaflet galla psyllid may be a promising candidate for the biological control of Brazilian peppertree.

ACKNOWLEDGMENTS

Assistance in Brazil was given by Maria F. Pollnow, Liliam Beal, André Buss, and Tatiana Reichert. We would also like to thank Dr. Daniel Burkhardt for his assistance in psyllid identification and Dr. Dean Williams for DNA analysis. We thank the following organizations for their financial support: the South Florida Water Management District, the Florida Fish and Wildlife Conservation Commission, Invasive Plant Management Division, the Florida Exotic Pest Plant Council, and the Julia Morton Invasive Plant Research Grant Program.

REFERENCES CITED

- ALVAREZ-ZAGOYA, R., AND CIBRIAN-TOVAR, D. 1999. Biology of the peppertree psyllid *Calophya rubra* (Blanchard) (Homoptera: Psyllidae). *Rev. Chapingo Serie Ciencias Forestales y del Ambiente* 5: 51-57.
- ANDERSON, R. P., LEW, D., AND PETERSON, A. T. 2003. Evaluating predictive models of species' distributions: criteria for selecting optimal models. *Ecol. Modeling* 162: 211-232.
- ANONYMOUS. 2007a. Other news: Brazilian pepper expands its range. *Wildland Weeds* 10: 29.
- ANONYMOUS. 2007b. Panhandlers beware! *Wildland Weeds* 11: 22.
- BARKLEY, F. A. 1944. *Schinus* L. *Brittonia* 5: 160-198.
- BARKLEY, F. A. 1957. A study of *Schinus* L. *Lilloa Rev. Botonica*, Tomo 28. Universidad Nacional de Tucumén, Argentina. 110 pp.
- BENNETT, F. D., CRESTANA, L., HABECK, D. H., AND BERTI-FILHO, E. 1990. Brazilian peppertree - prospects for biological control. Rome Italy: Istituto Sperimentale per la Patologia Vegetale, Ministero dell'Agricoltura e delle Foreste. Proc. VII Intl. Symp. Biol. Control Weeds. pp. 293-297.
- CAMPBELL, G. R., CAMPBELL, J. W., AND WINTERBOTHAM, A. L. 1980. The First Fund of Animals, Inc. *Schinus terebinthifolius* Brazil Expedition, July 1980- Interim Report. [Unpubl.].
- CENTER, T. D., PRATT, P. D., TIPPING, P. W., RAYAMAJHI, M. B., VAN, T. K., WINERITER, S. A., AND DRAY JR., F. A. 2007. Initial impacts and field validation of host range for *Boreioglycaspis melaleucae* Moore (Homoptera: Psyllidae), a biological control agent of the invasive tree *Melaleuca quinquenervia* (Cav.) Blake (Myrtales: Myrtaceae: Leptospermoideae). *Environ. Entomol.* 36: 569-576.
- CHRIST, L. R. 2010. Biology, population growth, and feeding preferences of *Calophya terebinthifolii* (Homoptera: Psyllidae), a candidate for biological control of Brazilian Peppertree, *Schinus terebinthifolius* (Sapindales: Anacardiaceae) M.Sc. thesis, Univ. Florida, Gainesville, Florida. 102 pp.
- CUDA, J. P., MEDAL, J. C., VITORINO, M. D., AND HABECK, D. H. 2005. Supplementary host specificity testing of the sawfly *Heteroperreyia hubrichi*, a candidate for classical biological control of Brazilian peppertree, *Schinus terebinthifolius*. *BioControl* 50: 195-201.
- CUDA, J. P., FERRITER, A. P., MANRIQUE, V., AND MEDAL, J. C. [eds.]. 2006. Florida's Brazilian Peppertree Management plan, 2nd edition: Recommendations from the Brazilian Peppertree Task Force, Florida Exotic Pest Plant Council, Apr 2006. <http://ipm.ifas.ufl.edu/pdf/BPmanagPlan.pdf>.
- CUDA, J. P., GILMORE, J. L., MEDAL, J. C., AND PEDROSA-MACEDO, J. H. 2008. Mass rearing of *Pseudophilothrips ichini* (Thysanoptera: Phlaeothripidae), an approved biological control agent for Brazilian peppertree, *Schinus terebinthifolius* (Sapindales: Anacardiaceae). *Florida Entomol.* 91: 338-340.
- CUDA, J. P., CHRIST, L. R., MANRIQUE, V., OVERHOLT, W. A., WHEELER, G. S., AND WILLIAMS, D. A. 2012. Role of molecular genetics in identifying 'fine tuned' natural enemies of the invasive Brazilian peppertree, *Schinus terebinthifolius*: a review. *BioControl* 57: 227-232.
- CUDA, J. P., MEDAL, J. C., GILMORE, J. L., HABECK, D. H., AND PEDROSA-MACEDO, J. H. 2009. Fundamental host range of *Pseudophilothrips ichini* sensu lato (Thysanoptera: Phlaeothripidae), a candidate biological control agent of *Schinus terebinthifolius* (Sapindales: Anacardiaceae) in the USA. *Environ. Entomol.* 38: 1642-1652.
- DELFOSE, E. S. 1979. Biological control: A strategy for plant management, pp. 83-86 *In* R. Workman [ed.], *Schinus*- Tech. Proc. Tech. for Control of *Schinus* in South Florida: A Workshop for Nat. Area Managers, 2 Dec 1978. The Sanibel Captiva Conservation Foundation, Inc., Sanibel, FL.
- DEVI, S. B., AND PRABHOO, N. R. 1995. Biology of leaf gall forming psyllid *Paurosylla turberculata* (Homoptera). *J. Ecobiol.* 7: 75-77.
- DOREN, R. F., WHITEAKER, L. D., AND LAROSA, A. M. 1991. Evaluation of fire as a management tool for controlling *Schinus terebinthifolius* as secondary successional growth on abandoned agricultural land. *Environ. Mgt.* 15: 121-129.
- DOWNER, J. A., SVIHRA, P., MOLINAR, R. H., FRASER, J. B., AND KOEHLER, C. S. 1988. New psyllid pest of California USA pepper tree. *Calif. Agric.* 42: 30-32.
- EDDMAPS. 2012. Early Detection & Distribution Mapping System. The University of Georgia - Center for Invasive Species and Ecosystem Health. Available online at <http://www.eddmaps.org/>; accessed 29 Oct 2012.
- EWEL, S., AND STERNBERG, L. 2005. Growth and gas exchange responses of Brazilian pepper (*Schinus terebinthifolius*) and native south Florida species to salinity. *Trees-Struct. Funct.* 19: 119-128.
- EWEL, J. J. 1979. Ecology of *Schinus*, pp. 7-21 *In* R. Workman [ed.], *Schinus*- Tech. Proc. Tech. for Control of *Schinus* in South Florida: A Workshop for Nat. Area Managers, 2 Dec 1978. The Sanibel Captiva Conservation Foundation, Inc., Sanibel, FL.
- FERREIRA, S. A., FERNANDES W. G., AND CARVALHO, L. G. 1990. Biology and natural history of *Euphaleurus ostreoides* (Homoptera: Psyllidae) a gall former on *Lonchocarpus guilleminiaus*. *Rev. Brasileira Biol.* 50: 417-424.
- GEIGER, C. A., AND GUTIERREZ, A. R. 2000. Ecology of *Heteropsylla cubana* (Homoptera: Psyllidae): Psyllid damage, tree phenology, thermal relations, and parasitism in the field. *Environ. Entomol.* 29: 76-86.
- GEIGER, J. H., PRATT, P. D., WHEELER, G. S., WILLIAMS, D. A. 2011. Hybrid vigor for the invasive exotic Brazilian peppertree (*Schinus terebinthifolius* Raddi., Anacardiaceae) in Florida. *Intl. J. Plant Sci.* 172: 655-663.

- HABECK, D. H., BENNETT, F. D., AND BALCIUNAS, J. K. 1994. Biological control of terrestrial and wetland weeds, pp. 523-548 *In* D. Rosen, F. D. Bennett and J. L. Capinera [eds.], Pest Management in the Subtropics: Biological Control - a Florida Perspective. Intercept Ltd., Andover, UK.
- HJLMANS, R. J., CAMERON, S. E., PARRA, J. L., JONES, P. G., AND JARVIS, A. 2005. Very high resolution interpolated climate surfaces for global land areas. *Intl. J. Climatol.* 25: 1965-1978.
- HODKINSON, I. D. 2009. Life cycle variation and adaptation in jumping plant lice (Insecta: Hemiptera: Psyllodea): a global synthesis. *J. Nat. Hist.* 43: 65-179.
- KUNIATA, L. S., AND KOROWI, K. T. 2004. Bugs offer sustainable control of *Mimosa invisa* and *Sida* spp. in the Markham Valley, Papua New Guinea. Canberra Australia: CSIRO Entomol.
- MACK, R. N. 1991. The commercial seed trade: An early disperser of weeds in the United States. *Econ. Bot.* 45: 257-273.
- MANRIQUE, V., CUDA, J. P., OVERHOLT, W. A., WILLIAMS, D. A., AND WHEELER, G. S. 2008. Effect of host-plant genotypes on the performance of three candidate biological control agents of *Schinus terebinthifolius* in Florida. *Biol. Control* 47: 167-171.
- MARTIN, C. G., CUDA, J. P., AWADZI, K. D., MEDAL, J. C., HABECK, D. H., AND PEDROSA-MACEDO, J. H. 2004. Biology and laboratory rearing of *Episimus utilis* (Lepidoptera: Tortricidae), a candidate for classical biological control of Brazilian peppertree, *Schinus terebinthifolius* (Anacardiaceae). *Florida Entomol.* 33: 1351-1361.
- McKAY, F., OLEIRO, M., VITORINO, M. D., AND WHEELER, G. 2012. The leafmining *Leurocephala schinusae* (Lepidoptera: Gracillariidae): Not suitable for the biological control of *Schinus terebinthifolius* (Sapindales: Anacardiaceae) in continental USA. *Biocontrol Sci. & Tech.* 22: 477-489.
- MEDAL, J. C., VITORINO, M. D., HABECK, D. H., GILLMORE, J. L., PEDROSA, J. H., AND DESOUSA, L. D. 1999. Host specificity of *Heteroperreyia hubrichi* Malaise (Hymenoptera: Pergidae), a potential biological control agent of Brazilian Peppertree (*Schinus terebinthifolius* Raddi). *Biol. Control.* 14: 60-65.
- MOERI, O. E., CUDA, J. P., OVERHOLT, W. A., BLOEM, S., AND CARPENTER, J. E. 2009. F1 sterile insect technique: A novel approach for risk assessment of *Episimus unguiculatus* (Lepidoptera: Tortricidae), a candidate biological control agent of *Schinus terebinthifolius* in the continental USA. *BioControl Sci. & Tech.* 19: 303-315.
- MOUND, L. A., WHEELER, G. S., AND WILLIAMS, D. A. 2010. Resolving cryptic species with morphology and DNA; thrips as a potential biocontrol agent of Brazilian peppertree, with a new species and overview of Pseudophilothrips (Thysanoptera). *Zootaxa.* 2432: 59-68.
- MUKHERJEE, A., WILLIAMS, D. A., WHEELER, G. S., CUDA, J. P., PAL, S., AND OVERHOLT, W. A. 2012. Brazilian peppertree (*Schinus terebinthifolius*) in Florida and South America: evidence of a possible niche shift driven by hybridization. *Biol. Invasions* 14: 1415-1430.
- NILSEN, E. T., AND MULLER, W. H. 1980. A comparison of the relative naturalizing ability of two *Schinus* species (Anacardiaceae) in southern California. II Seedling establishment. *Bull. Torrey Bot. Club* 107: 232-237.
- OLEIRO, M., MCKAY, F., AND WHEELER, G. S. 2011. Biology and host range of *Tecmessa elegans* (Lepidoptera: Notodontidae) a leaf-feeding moth evaluated as a potential biological control agent for *Schinus terebinthifolius* (Sapindales: Anacardiaceae) in the USA. *Environ. Entomol.* 40: 605-613.
- PEMBERTON, R. W. 2000. Predictable risk to native plants in weed biological control. *Oecologia* 125: 489-494.
- PHILLIPS, S. J., ANDERSON, R. P., AND SCHAPIRE, R. E. 2006. Maximum entropy modeling of species geographic distributions. *Ecol. Modeling* 190: 231-259.
- PRICE, P. W. 1997. *Insect Ecology*. 3rd ed. New York: Wiley.
- RAYAMAJHI, M. B., PRATT, P. D., CENTER, T. D., TIPPING, P. W., AND VAN, T. K. 2008. Aboveground biomass of an invasive tree melaleuca (*Melaleuca quinque-nervia*) before and after herbivory by adventive and introduced natural enemies: A temporal case study in Florida. *Weed Sci.* 56: 451-456.
- RAYAMAJHI, M. B., VAN, T. K., PRATT, P. D., CENTER, T. D., AND TIPPING, P. W. 2007. *Melaleuca quinque-nervia* dominated forests in Florida: Analyses of natural-enemy impacts on stand dynamics. *Plant Ecol.* 192: 119-132.
- RENDON, J., CHAWNER, M., DYER, K., AND WHEELER, G. S. 2012. Life history and host range of the leaf blotcher *Eucosomophora schinusivora*: A candidate for biological control of *Schinus terebinthifolius* in the USA. *Biocontrol Sci. & Tech.* 22: 711-722.
- RODGERS, L., BODLE, M., BLACK, D., AND LAROCHE, F. 2012. Status of nonindigenous species, Chapter 7: pp. 7-1 to 7-35 *In* 2012 South Florida Environmental Report, Vol. I. The South Florida Environment. South Florida Water Mgt. District, West Palm Beach, FL. http://my.sfwmd.gov/portal/page/portal/pg_grp_sfwmd_sfer/portlet_prevreport/2012_sfer/v1/chapters/v1_ch7.pdf.
- SAS. 2002. SAS System for Windows. Cary, NC, USA.
- SCHMITZ, D. C. 2007. Florida's invasive plant research: Historical perspective and the present research program. *Nat. Areas J.* 27: 251-253.
- SOKAL, R. R., AND ROHLF, F. J. 1995. *Biometry: The principles and practice of statistics in biological research*. New York, W. H. Freeman.
- VITORINO, M. D., CHRIST, L. R., BARBIERI, G., CUDA, J. P., AND MEDAL, J. C. 2011. *Calophya terebinthifolii* (Hemiptera: Calophyidae), a candidate for biological control of *Schinus terebinthifolius* (Sapindales: Anacardiaceae): feeding preferences and impact studies. *Florida Entomol.* 94: 694-695.
- WARING, G. L., AND COBB, N. S. 1992. The impact of plant stress on herbivore population dynamics, pp. 167-227 *In* E. A. Bernays [ed.], *Plant-Insect Interactions*. CRC Press, Boca Raton, Florida
- WHEELER, G. S., GEIGER, J. H., MCKAY, F., RENDON, J., CHAWNER, M., AND PRATT, P. D. 2010. Defoliating broad nosed weevil, *Plectrophoroides lutra*; not suitable for biological control of Brazilian Pepper. *Biocontrol Sci. Technol.* 21: 89-91.
- WIENS, J. J., GRAHAM, C. H., MOEN, D. S., SMITH, S. A., AND REEDER, T. W. 2006. Evolutionary and ecological causes of the latitudinal diversity gradient in Hylid frogs: treefrog trees unearth the roots of high tropical diversity. *Amer. Nat.* 168: 579-596.
- WILLIAMS, D. A., OVERHOLT, W. A., CUDA, J. P., AND HUGHES, C. R. 2005. Chloroplast and microsatellite DNA diversities reveal the introduction history of Brazilian peppertree (*Schinus terebinthifolius*) in Florida. *Mol. Ecol.* 14: 3643-3656.