

Morphology and ultrastructure of Tetranychusturkestani Ugarov & Nikolskii (Acari: Tetranychidae)

Authors: Jiang, Jue-Ying-Qi, Zhang, Yan-Nan, Guo, Dan-Dan, Zhang, Jian-Ping, and Chen, Jing

Source: Systematic and Applied Acarology, 22(8): 1181-1198

Published By: Systematic and Applied Acarology Society

URL: https://doi.org/10.11158/saa.22.8.5

The BioOne Digital Library (<u>https://bioone.org/</u>) provides worldwide distribution for more than 580 journals and eBooks from BioOne's community of over 150 nonprofit societies, research institutions, and university presses in the biological, ecological, and environmental sciences. The BioOne Digital Library encompasses the flagship aggregation BioOne Complete (<u>https://bioone.org/subscribe</u>), the BioOne Complete Archive (<u>https://bioone.org/archive</u>), and the BioOne eBooks program offerings ESA eBook Collection (<u>https://bioone.org/esa-ebooks</u>) and CSIRO Publishing BioSelect Collection (<u>https://bioone.org/csiro-ebooks</u>).

Your use of this PDF, the BioOne Digital Library, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at <u>www.bioone.org/terms-of-use</u>.

Usage of BioOne Digital Library content is strictly limited to personal, educational, and non-commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne is an innovative nonprofit that sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

Article http://zoobank.org/urn:lsid:zoobank.org:pub:84B087A3-E97F-44C9-A5C3-C5E76A25C596

Morphology and ultrastructure of *Tetranychus turkestani* Ugarov & Nikolskii (Acari: Tetranychidae)

JUE-YING-QI JIANG, YAN-NAN ZHANG, DAN-DAN GUO, JIAN-PING ZHANG* & JING CHEN*

College of Agriculture, Shihezi University, Shihezi, Xinjiang 832003, China *Corresponding author: E-mail: zhjp_agr@shzu.edu.cn; chj_agr@shzu.edu.cn

Abstract

Tetranychus turkestani is a serious pest of cotton, corn, vegetables, fruit trees and forests in Xinjiang. All life stages of *T. turkestani* were observed using both light and scanning electron microscope. The egg is smooth and spherical. The eupathidial spinnerets of larva and protonymph are different from that of the deutonymph and adult in shape. The spinneret of adult female is obviously thicker and larger than that in male. The palpfemoral seta of adult male is a short, stout, spine-like process. *T. turkestani* has fourteen pairs of dorsal setae. The number of ventral setae increases regularly in number in each molt. The leg chaetotaxy has a great change from larva to deutonymph.

Key words: Spider mite, adult, egg, larva, protonymph, deutonymph

Introduction

Tetranychus turkestani Ugarov et Nikolskii (Acari: Tetranychidae) is a serious pest on many plants, including cotton (*Gossypium hirsutum* L.), corn (*Zea mays* L.), sorghum (*Sorghum bicolor* L.), medlar (*Achras sapota* L.), tomato (*Lycopersicon esculentum* Mill.), bean (*Phaseolus vulgaris* L.), and other vegetables, fruit trees, and forest trees (Ugarov & Nikolskii 1937; Yu *et al.* 2000; Sohrabi & Shishehbor 2008; Guo *et al.* 2013; Zhang *et al.* 2016). It is mainly distributed in Russia, Kazakhstan, the United States, the Middle East and Xinjiang of China (Hill & Donnell 1991; Ros & Breeuwer 2007; Imani *et al.* 2009; Li *et al.* 2015). Previous research on *T. turkestani* focused on its biology and control (Yuan *et al.* 2008; Yang *et al.* 2012; Yang *et al.* 2013; Li *et al.* 2014; Duan *et al.* 2015; Liu *et al.* 2015). To our knowledge, its morphology has not been well studied, although all life stages of the mite have been simply described (Wang 1981; Lu 1990).

Morphological characters, such as the peritreme, aedeagus, tarsal claws and empodium, have often been used in identifying the species of Tetranychidae, and resolving the phylogenetic relationships between species (Gutierrez & Helle 1985; Lindquist 1985). Jeppson *et al.* (1975), Mollet & Sevacherian (1984) and Hong *et al.* (1994) studied the morphological characters and the density of the integumentary lobes and considered that the integumentary lobes were involved with thermal regulation. Some scholars have described the morphological characteristics of the mouthparts, setae and pedipalps, and inferred their functions from the morphology (Bostanian & Morrison 1973; Hislop & Jeppson 1976; Razaq *et al.* 2000).

The purpose of this study is to describe the morphological characteristics of all life stages of *T*. *turkestani* using light and scanning electron microscope, add ultrastructural details to the characteristics of *T*. *turkestani*.

© Systematic & Applied Acarology Society

Materials and Methods

Specimen preparation

The stock colony of *T. turkestani* was initiated from individuals collected from a cotton field near Huayuan, Shihezi City, Xinjiang Uygur Autonomous Region in 2010. This colony was maintained on potted sword bean (*Semen canavaliae* Gladiatae) in a growth chamber (FLI-2000H) at 26 ± 1 °C, 60% RH, and a 16:8 h (L:D) photoperiod.

Light microscope

Mite eggs of different ages, 24 hr, 48 hr and 72 hr were photographed and measured using a stereo microscope (Zeiss Discovery V20). The post embryonic stages including larva, protonymph, deutonymph, and adult (female and male) were slide mounted, photographed and measured under a compound microscope (Olympus BX53).

Scanning electron microscope

The samples were fixed overnight in fixative (2.5% glutaraldehyde and 10% paraformaldehyde in 0.2 M phosphate buffer pH 7.2) at 4 °C. The fixed samples were rinsed three times with the same buffer, and dehydrated in a graded series of acetone. Samples were then dried under natural conditions for 12 h. The dried samples were mounted on specimen stubs, coated with gold using ion sputter (E-1045), and observed under a scanning electron microscope (LEO 1430VP) at 20 kV and photographed.

Morphological measurements and statistical analyses

Body length and width, and idiosomal setal length were measured under the light microscope (LM) for 10 specimens, respectively. The data are presented as mean \pm s.e. and analysed using SPSS 17.0 (SPSS, Chicago, IL).

Morphological terminology

Morphological terminology was adopted principally from Wang (1981), Lindquist (1985) and Hong (2012).

Results

Both sexes of *T. turkestani* have five stages, including egg, larva, protonymph, deutonymph and adult (Figs. 1A-1F).

Adult female (n=10)

Body oval in shape, $540.23 \pm 3.40 \ \mu m$ long and $316.83 \pm 5.03 \ \mu m$ wide (Figs. 2A, 2B), light-green to dark-green in color. Two E-shaped dark patches present on both sides of idiosoma (Fig. 1E).

Gnathosoma

Chelicera consist of a pair of whip-like stylet (*St*) (Fig. 2C) and a cystiform stylophore (*Sp*) (Figs. 2D, 3A). *Palpus* (*Pa*) (Fig. 3A) has 5 segments: trochanter (*Tr*), femur (*Fe*), genu (*Ge*), tibia (*Ti*) and tarsus (*Ta*) (Fig. 3B). Palpfemoral seta (*d*) is setiform (Fig. 3A), palptibial claw (*CI*) is well developed (Fig. 3D), palptarsus has striated cuticle and comprises seven smooth projections (*su* ζ , ω , *ul*' ζ , *ul*" ζ , *a*, *b* and *c*) on its tip (Fig. 3D). Eupathidial spinneret (*su* ζ) is cylindrical and its length is approximately two times as long as its width. Solenidion (ω) is peg-shaped. Eupathidia (*ul*' ζ and

1182

SYSTEMATIC & APPLIED ACAROLOGY

VOL. 22

 $ul''\zeta$) are similar in form, elongate with a blunt apex. Three hair-like setae (a, b, c) are attenuate. Subcapitulum (Sub) bears a pair of subcapitular setae (m) (Fig. 3C). The apex of rostrum (Ro) is a large opening surrounded by soft flaps (Fig. 3C). Peritremes (Pe) are embedded in part of the membranous cuticle of stylophore. The last few segments of the peritreme are hooked (Fig. 2D).



FIGURE 1. All stages of *Tetranychus turkestani* (LM). A, Egg; B, Larva; C, Protonymph; D, Deutonymph; E, Female adult; F, Male adult.

Idiosoma

Dorsum: Fourteen pairs of dorsal setae $(v_2, sc_1, sc_2, c_1, c_2, c_3, d_1, d_2, e_1, e_2, f_1, f_2, h_2 \text{ and } h_3)$ present (Figs. 2A, 3E). Eyes (*Eye*) on either side are consistently present between setae sc_1 and sc_2 surrounded by striated integument (Fig. 3A). The striae between the setae v_2 and sc_1 is semiorbicalar, and the integumentary lobes are semi-oblong (Fig. 4A). The striae between the setae c_1, d_1 and e_1 is

transverse, and the integumentary lobes are semi-oblong or triangular (Figs. 4B, 4C). The striae between the seta e_1 is longitudinal and slanting, the integumentary lobes are semi-oblong (Fig. 4D). There is a diamond-shaped area between the setae e_1 and f_1 , the integumentary lobes are semi-oblong or triangular (Fig. 4E). There are three lyrifissure (*Ly*) on the outside of the setae e_2 , d_2 and e_2 , and surrounded by semi-oblong integumentary lobes (Fig. 4F).

Venter: The chaetotaxy includes six pairs of coxisternal setae (*1b*, *1c*, *2b*, *2c*, *3b* and *4b*) (Fig. 2B), three pairs of intercoxal setae (*1a*, *3a* and *4a*) (Fig. 2B), one pair of aggenital setae (*ag*) (Figs. 2B, 3E), two pairs of genital setae (g_1 and g_2) and two pairs of pseudanal setae (p_1 and p_2) (Fig. 3E). The genital opening (*GO*) and anal opening (*AO*) are on the ventral side of the opisthosoma (Fig. 3E). The genital opening is transverse and surrounded by characteristically wrinkled membranous cuticle, and the Semi-circular shaped genital flap (*GF*) is in front of it (Fig. 3E). The striae of the genital flap is transverse, without integumentary lobes (Fig. 3F). The striae in the upper area of the genital flap is longitudinal and have a small number of granules (Fig. 3F).

Legs: Each leg includes coxa (Cx), trochanter (Tr), femur (Fe), genu (Ge), tibia (Ti), tarsus (Ta) and pretarsus (Pr) (Fig. 5A). There are nine tactile setae and one solenidion on tibia I (Fig. 5B). Two pairs of duplex setae (ω' , ft' and ω'' , ft'') are found on the dorsum of tarsus I (Fig. 5C). The longer seta (ω' and ω'') is elongate and tapered, and the shorter seta (ft' and ft'') is barbed and apically forked. Tarsus I also has other kinds of setae, such as tectal setae (tc'), prorals ($p'\zeta$) and primiventrals (pv) (Fig. 5D). The barbed tectal setae have well-developed sockets, while processes (Fig. 5D). The claws of tarsus I are strongly reduced to two pairs of tenent hairs (TH) (Fig. 5D).



FIGURE 2. *Tetranychus turkestani* (adult female, LM). A, Dorsal view; B, Ventral view; C, Stylet (*St*) in ventral view; D, Peritreme (*Pe*) and stylophore (*Sp*) in dorsal view.

1184

Adult male (n=10)

Body diamond-shaped, $412.10 \pm 3.63 \mu m$ long and $192.53 \pm 2.73 \mu m$ wide (Figs. 6A, 6B), light-green in colour (Fig. 1F).



FIGURE 3. *Tetranychus turkestani* (adult female, SEM). A, Gnathosoma in lateral view, showing eye (*Eye*), palpfemoral seta (*d*), palpus (*Pa*), and stylophore (*Sp*); B, Pedipalp in lateral view, showing trochanter (*Tr*), femur (*Fe*), genu (*Ge*), tibia (*Ti*) and tarsus (*Ta*); C, Gnathosoma in ventral view, showing rostrum (*Ro*), subcapitular setae (*m*) and subcapitulum (*Sub*); D, Palptarsus in ventral view, showing palpotibial claw (*CI*), palpotarsal eupathidial spinneret (*su* ζ), eupathidium (*ul*' ζ and *ul*" ζ), setae (*a*, *b* and *c*), solenidion (ω); E, Opisthosoma in ventral view, showing aggenital setae (*ag*), anal opening (*AO*), genital flap (*GF*), genital opening (*GO*), genital setae (*g*₁ and *g*₂), para-anals (*h*₃), post-anals (*h*₂) and pseudanal setae (*ps*₁ and *ps*₂); F, Genital region.



FIGURE 4. *Tetranychus turkestani* (adult female, SEM). A, The striae and integumentary lobes between the setae v_2 and sc_1 ; B, C The striae and integumentary lobes between the setae c_1 , d_1 and e_1 ; D, The striae and integumentary lobes between the setae e_1 ; E, The striae and integumentary lobes between the setae e_1 and f_1 ; F, lyrifissure (*Ly*).

Gnathosoma

Chelicera: The shape of the stylophore (*Sp*) and stylet (*St*) is similar to that of adult female (Figs. 6C, 7A). *Palpus (Pa)* (Fig. 7A) also has 5 segments (Fig. 7B). The palpfemoral seta (*d*) is a short, stout, spine-like process (Fig. 7B). The palptarsus also has seven smooth-surfaced projections ($su\zeta$, ω , $ul'\zeta$, $ul''\zeta$, *a*, *b* and *c*) (Fig. 7C). Eupathidial spinneret ($su\zeta$) is obviously shortened and smaller than that in female. *Subcapitulum (Sub)*, *rostrum (Ro)* and *peritreme (Pe)* are similar to those of adult female (Figs. 6C, 7C). The rostral gutter (*RG*) is located on the dorsal surface of the rostrum (Fig. 7B).



FIGURE 5. *Tetranychus turkestani* (adult female, SEM). A, Leg I in ventral view, showing coxa (*Cx*), trochanter (*Tr*), femur (*Fe*), genu (*Ge*), tibia (*Ti*), tarsus (*Ta*) and pretarsus (*Pr*); B, Tibia (*Ti*) and Tarsus (*Ta*) I in lateral view; C, Duplex setae (ω' , ft' and ω'' , ft''); D, Tarsus and pretarsus I in lateral view, showing empodium (*Em*), prorals ($p'\zeta$), tenent hairs (*TH*), tectal setae (tc') and primiventrals (pv).

Idiosoma

Dorsum has fourteen pairs of dorsal setae $(v_2, sc_1, sc_2, c_1, c_2, c_3, d_1, d_2, e_1, e_2, f_1, f_2, h_2 \text{ and } h_3)$ (Figs. 6A, 7D).

Venter: The chaetotaxy includes six pairs of coxisternal setae (*1b*, *1c*, *2b*, *2c*, *3b* and *4b*), three pairs of intercoxal setae (*1a*, *3a* and *4a*), one pair of aggenital setae (*ag*) (Fig. 6B), two pairs of genital setae (g_1 and g_2) and two pairs of pseudanal setae (ps_1 and ps_2) (Fig. 7D).

Aedeagus: The shaft (*Sh*) of aedeagus is bent back to form a large terminal knob (*TK*) with an anterior projection that is blunt and a posterior projection that is acute. There is an obvious angle on the posterior projection near the 1/3 region (Fig. 6D).

Legs also have 7 segments. There are nine tactile setae and four solenidion on tibia I (Fig. 8B). The shape and situation of the duplex setae (ω' , ft' and ω'' , ft'') are similar to those of adult female (Figs. 8A, 8C). The primilaterals (pl', pl'') are located laterally and apically on tarsus I (Fig. 8D). The primiventrals (pv) is located ventrally (Fig. 8D). Empodium I (Em) consists of a claw-like structure with a strong mediodorsal spur about one half of the appendage (Fig. 8D). The claws of tarsus I are reduced to two pairs of tenent hairs (TH) (Fig. 8D).



FIGURE 6. *Tetranychus turkestani* (adult male, LM). A, Dorsal view; B, Ventral view; C, Peritreme (*Pe*) and stylophore (*Sp*) in dorsal view; D, Aedeagus in lateral view, showing shaft (*Sh*) and terminal knob (*TK*).

Eggs (*n*=10)

It is smooth and spherical in shape with a diameter of about $129.55 \pm 1.86 \,\mu\text{m}$ (Figs. 1A, 9D). The freshly laid eggs are colorless and transparent (Fig. 9A) and then gradually become canary yellow to dark yellow within one or two days (Figs. 9B, 9C). Eye spots appear redon the third day (Fig. 9C).

Larva (n=10)

Body $223.85 \pm 6.24 \ \mu\text{m}$ long and $139.96 \pm 5.30 \ \mu\text{m}$ wide (Figs. 10A, 10B), from transparent to turn sap green after feeding (Fig. 1B), bearing three pairs of legs.

Palptarsus has seven smoothly surfaced projections $(su\zeta, \omega, ul'\zeta, ul''\zeta, a, b \text{ and } c)$ (Fig. 10C). Eupathidial spinneret $(su\zeta)$ is a slender cylinder structure, and its length is about three times its width. The shapes of the solenidion (ω) , two eupathidia $(ul'\zeta \text{ and } ul''\zeta)$ and three setae (a, b and c) are similar to those of adult female (Fig. 10C).

Dorsum: Larvae have fourteen pairs of dorsal setae $(v_2, sc_1, sc_2, c_1, c_2, c_3, d_1, d_2, e_1, e_2, f_1, f_2, h_2$ and h_3) (Figs. 10A, 10D).

Venter: The chaetotaxy includes one pair of coxisternal setae (*1b*) (Fig. 10B), two pairs of intercoxal setae (*1a* and *3a*) (Fig. 10B) and two pairs of pseudanal setae (ps_1 and ps_2) (Fig. 10D). *Legs*: Leg chaetotaxy as follows: I-1-0-3-4-6-7 + 1 duplex; II- 0-0-3-4-5-7 + 1 duplex; III-0-0-2-2-5-6 (Figs. 11A–11C).



FIGURE 7. *Tetranychus turkestani* (adult male, SEM). A, B, Gnathosoma in dorsal view, showing palpi (*Pa*), rostral gutter (*RG*), stylophore (*Sp*), stylet (*St*), palpfemoral seta (*d*), trochanter (*Tr*), femur (*Fe*), genu (*Ge*), tibia (*Ti*) and tarsus (*Ta*); C, Gnathosoma in ventral view, showing palpotarsal eupathidial spinneret (*su* ζ), eupathidium (*ul'* ζ and *ul''* ζ), rostrum (*Ro*), setae (*a*, *b* and *c*), solenidion (ω), subcapitular setae (*m*) and subcapitulum (*Sub*); D, Opisthosoma in ventral view, showing genital setae (*g*₁ and *g*₂), para-anals (*h*₃) and pseudanal setae (*ps*₁ and *ps*₂).

Protonymph (n=10)

Body $267.87 \pm 5.23 \ \mu\text{m}$ long and $176.69 \pm 5.42 \ \mu\text{m}$ wide (Figs. 12A, 12B), yellow-green after incubation, and it will turn dark green after it's sucking the sap of plants (Fig. 1C).

Palptarsus has seven smooth projections ($su\zeta$, ω , $ul'\zeta$, $ul''\zeta$, a, b and c) which are similar to those of larvae (Fig. 12C).

Dorsum: Protonymph has fourteen pairs of dorsal setae $(v_2, sc_1, sc_2, c_1, c_2, c_3, d_1, d_2, e_1, e_2, f_1, f_2, h_2$ and h_3) (Figs. 12A, 12D).

Venter: In addition to those setae in larva, three pairs of coxisternal setae (1c, 2b and 3b) (Fig. 12B) and one pair of aggenital setae (ag) are added (Fig. 12D).

Legs: Leg chaetotaxy as follows: I-2-0-3-4-6-9 + 2 duplexes; II- 1-0-3-4-5-9 + 1 duplex; III-1-0-2-2-5-8; IV-0-0-2-2-5-6 (Figs. 13A–13D).

Deutonymph (n=10)

Body $372.89 \pm 11.78 \ \mu m$ long and $212.93 \pm 7.10 \ \mu m$ wide (Figs. 14A, 14B); light-green to dark-green in color (Fig. 1D).

Palptarsus has seven smooth projections ($su\zeta$, ω , $ul'\zeta$, $ul''\zeta$, a, b and c) which are similar to those in adult female (Fig. 14C).



FIGURE 8. *Tetranychus turkestani* (adult male, SEM). A, Tarsus and pretarsus I in lateral view; B, Tibia (*Ti*) and Tarsus (*Ta*) I in lateral view; C, Duplex setae (ω' and *ft'*); D, Tarsus and pretarsus I in lateral view, showing empodium (*Em*), primilaterals (*pl'* and *pl''*), primiventrals (*pv*), tenent hairs (*TH*) and tectal setae (*tc'*).



FIGURE 9. Tetranychus turkestani (egg, LM, SEM). A, 24 hr; B, 48 hr; C, 72 hr; D, 48 hr.



FIGURE 10. *Tetranychus turkestani* (larva, LM, SEM). A, Dorsal view; B, Ventral view; C, Palptarsus in ventral view, showing eupathidial spinneret ($su\zeta$), eupathidium ($ul'\zeta$ and $ul''\zeta$), setae (a, b and c), and solenidion (ω); D, Opisthosoma in ventral view, showing para-anals (h_3), post-anals (h_2) and pseudanal setae (ps_1 and ps_2).



FIGURE 11. Tetranychus turkestani (larva). A, Leg I; B, Leg II; C, Leg III.



FIGURE 12. *Tetranychus turkestani* (protonymph, LM, SEM). A, Dorsal view; B, Ventral view; C, Palptarsus in ventral view, showing eupathidial spinneret ($su\zeta$), eupathidium ($ul'\zeta$ and $ul''\zeta$), setae (a, b and c) and solenidion (ω); D, Opisthosoma in ventral view, showing aggenital setae (ag), para-anals (h_3), post-anals (h_2) and pseudanal setae (ps_1 and ps_2).



FIGURE 13. *Tetranychus turkestani* (protonymph). A, Tibia and tarsus I; B, Tibia and tarsus II; C, Tibia and tarsus III; D, Tibia and tarsus IV.

1192

SYSTEMATIC & APPLIED ACAROLOGY

VOL. 22



FIGURE 14. *Tetranychus turkestani* (deutonymph, LM, SEM). A, Dorsal view; B, Ventral view; C, Palptarsus in ventral view, showing eupathidial spinneret ($su\zeta$), eupathidium ($ul'\zeta$ and $ul''\zeta$), setae (a, b and c) and solenidion (ω); D, Opisthosoma in ventral view, showing aggenital setae (ag), genital setae (g_1), para-anals (h_3), post-anals (h_2) and pseudanal setae (ps_1 and ps_2).



FIGURE 15. *Tetranychus turkestani* (deutonymph). A, Tibia and tarsus I; B, Tibia and tarsus II; C, Tibia and tarsus III; D, Tibia and tarsus IV.

Dorsum bears fourteen pairs of dorsal setae $(v_2, sc_1, sc_2, c_1, c_2, c_3, d_1, d_2, e_1, e_2, f_1, f_2, h_2 \text{ and } h_3)$ (Figs. 14A, 14D).

Venter: In addition to those setae in protonymph, two pairs of coxisternal setae (2c and 4b) (Fig. 14B), one pair of intercoxal setae (4a) (Fig. 14B) and one pair of genital setae (g_1) (Fig. 14D) are added. Genital folds begin to form.

Legs: Leg chaetotaxy as follows: I-2-1-5-5-10-13 + 2 duplexes; II- 2-1-4-4-6-10 + 1 duplex; III-1-1-2-3-5-9; IV-1-0-2-3-5-8 (Figs. 15A-15D).

Comparison of the lengths of dorsal setae of different development stages

In female the lengths of the dorsal setae increase significantly as the mites grow from larva to adult (Table 1). Seta sc_1 is the longest at the adult stage and shortest at the larval stage. All dorsal setae of adult female are significantly longer than those of male. Lengths of v_2 , sc_2 , c_2 , d_1 , d_2 and e_2 don't change significantly in deutonymph and adult male. The length of h_3 shows no significant differences at the stages of larva, protonymph and adult male.

| Length | Development stage | | | | |
|-----------------------|----------------------------|-------------------|-----------------------------|-----------------------------|-----------------------------|
| (µm) | Larva | Protonymph | Deutonymph | Female adult | Male adult |
| v ₂ | 46.69 ± 0.59 a | $50.19\pm0.67~b$ | $59.63\pm0.46~c$ | $75.74 \pm 1.84 \text{ d}$ | $61.03 \pm 0.48 \text{ c}$ |
| sc_1 | 65.76 ± 0.98 a | $89.12\pm0.57~b$ | $107.13 \pm 0.94 \text{ d}$ | $139.91 \pm 1.30 \text{ e}$ | $103.10 \pm 0.60 \text{ c}$ |
| sc_2 | $48.51 \pm 0.59 \text{ a}$ | $59.82\pm0.44\ b$ | $77.51 \pm 0.57 \ c$ | $109.22 \pm 1.39 \text{ d}$ | $75.81 \pm 0.53 \ c$ |
| \mathbf{c}_1 | 53.66 ± 0.81 a | $72.02\pm0.54~b$ | $98.39 \pm 0.97 \text{ d}$ | $132.76 \pm 1.70 \text{ e}$ | $90.70\pm0.88~c$ |
| c ₂ | $52.90\pm0.97~a$ | $70.23\pm0.97~b$ | $92.86 \pm 0.77 \ c$ | $129.71 \pm 0.89 \; d$ | $91.61 \pm 0.62 \ c$ |
| c ₃ | $47.01 \pm 0.58 \text{ a}$ | $59.69\pm1.09\ b$ | $81.69\pm0.84\ d$ | $108.97 \pm 1.71 \text{ e}$ | $78.29 \pm 1.25 \ c$ |
| d_1 | $48.61 \pm 1.30 \text{ a}$ | $69.83\pm0.88~b$ | $92.50\pm0.89\ c$ | $129.06 \pm 1.80 \; d$ | $90.38\pm0.80\ c$ |
| d ₂ | 47.48 ± 0.52 a | $68.49\pm0.82~b$ | $89.70\pm0.41~c$ | $131.68 \pm 1.39 \text{ d}$ | $90.42 \pm 1.17 \text{ c}$ |
| e ₁ | 46.07 ± 0.73 a | $66.62\pm0.60~b$ | $85.84 \pm 0.58 \; d$ | $121.80 \pm 1.51 \text{ e}$ | $83.28\pm0.44~c$ |
| e ₂ | $44.08\pm0.40~a$ | $64.10\pm1.39~b$ | $87.20\pm0.94~c$ | $124.30 \pm 1.20 \text{ d}$ | 85.51 ± 0.46 c |
| f_1 | 37.44 ± 0.90 a | $56.56\pm0.64~b$ | $74.88 \pm 0.65 \ d$ | 106.83 ± 0.77 e | $59.67 \pm 0.94 \ c$ |
| f_2 | 31.36 ± 0.27 a | $46.72\pm0.82\ b$ | $64.77 \pm 0.57 \text{ d}$ | $94.70 \pm 1.15 \text{ e}$ | 55.18 ± 0.73 c |
| h_2 | $19.63 \pm 0.76 \text{ a}$ | $22.97\pm0.67~b$ | $27.35\pm0.87~c$ | $44.73\pm1.06~d$ | $20.40\pm0.45a$ |
| h ₃ | $22.95\pm0.26~a$ | $24.00\pm0.30~a$ | $28.37\pm0.49\ b$ | $43.15\pm0.56\ c$ | 23.33 ± 0.18 a |

TABLE 1. The length of dorsal setae of *T. turkestani* at different development stages (*n*=10).

Note: Data in the table are MEAN \pm SE. Different letter labels in the same line indicate a significant difference (P<0.05).

Discussion

The shapes of the stylophore, stylets, palpi and rostral gutter of *T. turkestani* are similar to other spider mites, such as *T. atlanticus*, *T. urticae*, *Oligonychus punicae* and *Panonychus citri* (Baker & Connell 1963; Bostanian & Morrison 1973; Hislop & Jeppson 1976; Razaq *et al.* 2000). The stylophore and rostral gutter have the function of protecting the stylets, and the rostral gutter serves as a vessel for stylets to protract and retract (Razaq *et al.* 2000). In comparison to Eriophyoidea which has short stylets and are capable of very shallow penetration into a plant (Jeppson *et al.* 1975), Tetranychoidea possesses long stylets which could easily penetrate plant tissues, so it may kill the plant cells (Evans *et al.* 1961).

The pattern of the striae between setae e_1 and f_1 may be of diagnostic value in adult females in Tetranychidae (Lindquist 1985). The ultrastructure of the striae and integumentary lobes are also of diagnostic importance in distinguishing diapausing and non-diapausing populations of the same

species or between closely related species (Boudreaux & Dosse 1963; Dosse & Boudreaux 1963; Jeppson *et al.* 1975). The morphological characters of the striae and integumentary lobes of *T. urticae* have been reported (Brandenburg & Kennedy 1981; Mollet & Sevacherian 1984). There is also a diamond-shaped area between the setae e_1 and f_1 in *T. urticae*, and the shape of integumentary lobes is different in different regions (Carbonnelle & Hance 2004). The shape of integumentary lobes of *T. turkestani* is similar to that of feeding green females of *T. urticae* collected in Lattes, France (Carbonnelle & Hance 2004).

Spider mites have the habit of spinning webs (Saito 1977, 1983). Adult female of *T. turkestani* covers its eggs and lavae with webs to protect them from predators and other external factors. The eupathidial spinneret is more developed in adult female than in male. The shape of the eupathidial spinneret of *T. turkestani* adults is similar to that of *T. urticae* and *T. truncatus* (Bostanian & Morrison 1973; Sakunwarin *et al.* 2004), but different from that of *T. bunda* and *T. musae* (Flechtmann & Knihinicki 2002; Auger *et al.* 2008). It is much slender, about three times as long as wide in *T. bunda* (Flechtmann & Knihinicki 2002) and as long as wide in *T. musae* (Auger *et al.* 2008).

Many sensory receptors are found on the tip of the mouthparts and legs of mites (Alberti & Coons 1999; Walter & Proctor 1999). Some scholars have studied the sensilla of *T. urticae* and *T. truncatus*, and they concluded that the solenidion and two eupathidia on palptarsus are chemoreceptors, the three setae are mechanosensitive sensilla, and the solenidia (ω' and ω'') of duplex setae are chemosensitive sensilla (Bostanian & Morrison 1973; Sakunwarin *et al.* 2004). The six sensilla on the palptarsus of *T. turkestani* adults are similar in shape and position to those of *T. urticae* and *T. truncatus*.

Most of the articles mainly describe the leg chaetotaxy of female and male mites (Stone 1986; Ehara & Gotoh 1992; Ehara 1995; Flechtmann & Knihinicki 2002), and we studied the leg chaetotaxy of all stages. There is a great change in the leg chaetotaxy of tibia and tarsus from larva to deutonymph, and the chaetotaxy of leg III of larva is consistent with the chaetotaxy of leg IV of protonymph. The shapes of the tarsal claw and empodium in Tetranychidae are important taxonomic characters (Wang 1981). The shape of the empodium I of *T. turkestani* is different from that of *T. musae* (Auger *et al.* 2008). The empodium I of female adult *T. turkestani* have three pairs of minute, attenuate hair-like processes, while that of *T. musae* bear three pairs of proximoventral hairs and a large mediodorsal spurs (Auger *et al.* 2008). The empodium I of male adult *T. turkestani* has a similar shape as the empodium II of that of *T. lintearius*, which consists of three pairs of distally fused proximoventral spurs and possesses a strong mediodorsal spur (Stone 1986). However, the empodium I of male *T. musae* adults consists of a double claw-like structure with a strong mediodorsal spur (Auger *et al.* 2008). To summarize, this paper increases understanding of the external morphological characteristics of *T. turkestani*.

Acknowledgments

We thank Dr. John Richard Schrock from Emporia State University, USA for his generous help with revising the manuscript. We also thank Professor Jeff Geer from College of Agriculture, Shihezi University, Xinjiang for reading the manuscript. We are grateful to Professor Zhaotian Fan from Xinjiang University who helped us take the SEM photos. We appreciate Professor Jianzhen Lin from Institute of Plant Protection, Fujian Academy of Agricultural Sciences and Dr Qing-Hai Fan from Plant Health & Environment Laboratory, Ministry for Primary Industries, New Zealand for sending some literatures to us. This research was supported by Public Welfare Industry (Agriculture) Research Project (No. 201103020).

References

- Alberti, G. & Coons, L.B. (1999) Acari Mites. In: Harrison, F.W. (ed.) Microscopic Anatomy of Invertebrates. Vol. 8c. John Wiley & Sons, Inc., New York, pp. 515–1265.
- Auger, P., Migeon, A. & Flechtmann, C.H.W. (2008) Description of a new *Tetranychus* (Acarina, Prostigmata: Tetranychidae) pest of *Musa* from French Guiana, *Journal of Natural History*, 42(27–28), 1885–1892. http://dx.doi.org/10.1080/00222930802109850
- Baker, J.E. & Connell, W.A. (1963) The morphology of the mouthparts of *Tetranychus atlanticus* and observations on feeding by this mite on soybeans. *Annals of the Entomological Society of America*, 56(6), 733– 736.

http://dx.doi.org/10.1093/aesa/56.6.733

- Bostanian, N.J. & Morrison, F.O. (1973) Morphology and ultrastructure of sense organs in the two-spotted spider mite (Acarina: Tetranychidae). *Annals of the Entomological Society of America*, 66(2), 379–383. http://dx.doi.org/10.1093/aesa/66.2.379
- Boudreaux, H.B. & Dosse, G. (1963) The usefulness of new taxonomic characters in females of the genus *Tetranychus* Dufour (Acari: Tetranychidae). *Acarology*, 5, 13–33.
- Brandenburg, R.L. & Kennedy, G.G. (1981) Differences in dorsal integumentary lobe densities between *Tetranychus urticae* Koch and *Tetranychus cinnabarinus* (Boisduval) (Acarina: Tetranychidae) from Northeastern North Carolina. *International Journal of Acarology*, 7(1–4), 231–234. http://dx.doi.org/10.1080/01647958108683266
- Carbonnelle, S. & Hance, T. (2004) Cuticular lobes in the *Tetranychus urticae* complex (Acari: Tetranychidae): a reliable taxonomic character? *Belgian Journal of Zoology*, 134(2/1), 51–54.
- Duan, X.K., Li, Y.T., Chen, L., Zhang, Y.N., Wang, Y.Q. & Zhang, J.P. (2015) Sublethal effects of a novel acaricide Cyflumetofen on *Tetranychus turkestani*. *Journal of Environmental Entomology*, 37(2), 372–380. (in Chinese)
- Dosse, G. & Boudreaux, H.B. (1963) Some problems of spider mite taxonomy involving genetics and morphology. *In*: J.A. Naegele (Editor), *Advances in Acarology*, Cornell University Vol. 1., pp. 343–349.
- Ehara, S. (1995) A new species of *Tetranychus* (Acari, Tetranychidae) from the Ryukyu Islands. *Japanese Journal of Entomology*, 63(1), 229–233.
- Ehara, S. & Gotoh, T., (1992) Descriptions of two *Panonychus* spider mites from Japan, with a key to species of the genus in the world (Acari: Tetranychidae). *Applied Entomology and Zoology*, 27, 107–115.
- Evans, G.O., Sheals, J.G. & Macfarlane, D. (1961) The Terrestrial Acari of the British Isles. An Introduction to their Morphology, Biology and Classification. Vol. I. Introduction and Biology. Bartholomew Press, Lon don, 219 pp.
- Flechtmann, C.H.W. & Knihinicki, D.K. (2002) New species and new record of *Tetranychus* Dufour from Australia, with a key to the major groups in this genus based on females (Acari: Prostigmata: Tetranychidae). *Austral Entomology*, 41(2), 118–127.

http://dx.doi.org/10.1046/j.1440-6055.2002.00289.x

Guo, Y.L., Jiao, X.D., Xu, J.J., Yang, S., Duan, X.K. & Zhang, J.P. (2013) Growth and reproduction of *Tetrany-chus turkestani* and *Tetranychus truncatus* (Acari: Tetranychidae) on cotton and corn. *Systematic and Applied Acarology*, 18(1), 89–98.

http://dx.doi.org/10.11158/saa.18.1.10

- Gutierrez, J. & Helle, W. (1985) Evolutionary changes in the Tetranychidae. *In*: Helle, W. & Sabelis, M.W. (ed.) *Spider mites: their biology, natural enemies and control*. Elsevier, Amsterdam, pp. 91–107.
- Hill, R.L. & Donnell, D.J. (1991) Reproductive isolation between *Tetranychus lintearius* and two related mites, *T. urticae* and *T. turkestani* (Acarina: Tetranychidae). *Experimental and Applied Acarology*, 11(4), 241– 251.
- Hislop, R.G. & Jeppson, L.R. (1976) Morphology of the mouthparts of several species of phytophagous mites. Annals of the Entomological Society of America, 69(6), 1125–1135. http://dx.doi.org/10.1093/aesa/69.6.1125

Hong, X.Y. (2012) Agricultural Acarology. China Agriculture Publishers, Beijing, pp. 1–51.

- Hong, X.Y., Wang, Y.C. & You, Z.P. (1994) Scanning electron microscopic observation on the body surface of *Tetranychus cinnabarinus* (Boisduval). *Journal of Nanjing Agricultural University*, 17(2), 48–53. (in Chinese)
- Imani, Z., Shishehbor, P. & Sohrabi, F. (2009) The effect of *Tetranychus turkestani* and *Eutetranychus Orientalis* (Acari: Tetranychidae) on the development and reproduction of *Stethorus gilvifrons* (Coleoptera: Coc-

1196

SYSTEMATIC & APPLIED ACAROLOGY

VOL. 22

cinellidae). *Journal of Asia-Pacific Entomology*, 12(4), 213–216. http://dx.doi.org/10.1016/j.aspen.2009.05.004

- Jeppson, L.R., Baker, E.W. & Keifer, H.H. (1975) *Mites Injurious to Economic Plants*. University of California Press, Berkeley, 614 pp.
- Li, G.Y., Li, J.J., Xia, W., Qu, H.L., Yang, S. & Zhang, J.P. (2014) Effects of Bt+CpTI transgenic cotton on the performance of *Tetranychus turkestani* (Acari: Tetranychidae). *Systematic and Applied Acarology*, 19(2), 236–247.

http://dx.doi.org/10.11158/saa.19.2.14

- Li, Y.T., Jiang, J.Y.Q., Huang, Y.Q., Wang, Z.H. & Zhang, J.P. (2015) Effects of temperature on development and reproduction of *Neoseiulus bicaudus* (Phytoseiidae) feeding on *Tetranychus turkestani* (Tetranychidae). *Systematic and Applied Acarology*, 20(5), 478–490. http://dx.doi.org/10.11158/saa.20.5.4
- Lindquist, E.E. (1985) External anatomy. In: Helle, W. & Sabelis, M.W. (eds.) Spider Mites: Their Biology, Natural Enemies and Control. World Crop Pests IA. Elsevier Publications, Cambridge, U.K., pp. 3–26.
- Liu, M., Li, Y.T., Li, T., Su, J., Duan, X.K., Wang, Z.H. & Zhang, J.P. (2015) Effects of sub-lethal concentrations of Azocyclotin on the movement rates of *Tetranychus turkestani* (Ugarov et Nikolskii). *Chinese Journal of Applied Entomology*, 52(3), 593–599. (in Chinese)
- Lu, S.L. (1990) Preliminary report on *Tetranychus turkestani* (Acari: Tetranychidae). *Journal of Xinjiang Agricultural Science*, (3), 118–119. (in Chinese)
- Mollet, J.A. & Sevacherian, V. (1984) Effect of temperature and humidity on dorsal strial lobe densities in *Tetranychus* (Acari: Tetranychidae). *International Journal of Acarology*, 10(3), 159–161. http://dx.doi.org/10.1080/01647958408683370
- Razaq, A., Ohbayashi, N., Shiraishi, M., Ono, H. & Fujibuchi, M. (2000) Scanning electron microscopic observations on the mouthparts of *Panonychus citri* (McGregor) (Acari: Tetranychidae) and *Agistemus terminalis* (Quayle) (Acari: Stigmaeidae) on Satsuma Mandarin. *Applied Entomology and Zoology*, 35(1), 189–198.

https://doi.org/10.1303/aez.2000.189

Ros, V.I.D. & Breeuwer, J.A.J. (2007) Spider mite (Acari: Tetranychidae) mitochondrial COI phylogeny reviewed: host plant relationships, phylogeography, reproductive parasites and barcoding. *Experimental* and Applied Acarology, 42(4), 239–262.

http://dx.doi.org/10.1007/s10493-007-9092-z

- Saito, Y. (1977) Study on spinning behavior of spider mites (Acarina: Tetranychidae). I. Method for quantitative evaluation of the mite webbing, and the relationship between webbing and walking. *Japanese Journal of Applied Entomology and Zoology*, 21(1), 27–34.
- Saito, Y. (1983) The concept of "life types" in Tetranychinae. An attempt of classify the spinning behaviour of Tetranychinae. *Acarologia*, 24, 377–391.
- Sakunwarin, S., Baker, G.T. & Chandrapatya, A. (2004) Structure of sensilla on the palptarsus and the tarsus I of *Tetranychus truncatus* Ehara (Acari: Tetranychidae). *Systematic and Applied Acarology*, 9(1), 133–140.

http://dx.doi.org/10.11158/saa.9.1.18

Sohrabi, F. & Shishehbor, P. (2008) Effects of host plant and temperature on growth and reproduction of the strawberry spider mite *Tetranychus turkestani* Ugarov & Nikolskii (Acari: Tetranychidae). *Systematic and Applied Acarology*, 13, 26–32.

https://doi.org/10.11158/saa.13.1.2

- Stone, C. (1986) An investigation into the morphology and biology of *Tetranychus lintearius* Dufour (Acari: Tetranychidae). *Experimental and Applied Acarology*, 2(2), 173–186. https://doi.org/10.1007/BF01213760
- Ugarov, A.A. & Nikolskii, V.V. (1937) Systematic study of spider mites from Central Asia. *Trudy Sredneazi*atskoi Stantsii Zashchaty Rastenii, 2, 26–64.
- Walter, D.E. & Proctor, H.C. (1999) *Mites: Ecology, Evolution and Behaviour*. CABI Publishing, New York, 322 pp.
- Wang, H.F. (1981) Economic Insect Fauna of China. Fasc. 23. Acari: Tetranychoidea. Science Press, Beijing, China, pp. 125–126.
- Yang, S., Guo, Y.L., Jiao, X.D., Wang, P.L., Yang, D.S. & Zhang, J.P. (2012) Comparison of susceptibility to acaricides between *Tetranychus turkestani* and *Tetranychus truncates*. Acta Agriculturae Boreali-occidentalis Sinica, 21(12), 188–191. (in Chinese)
- 2017 JIANG ET AL.: MORPHOLOGY AND ULTRASTRUCTURE OF TETRANYCHUS TURKESTANI 1197

Yang, S., He, Y.F., He, H., Li, T., Wei, Q.Y., Feng, P. & Zhang, J.P. (2013) Comparison of the predation of Stethorus punctillum on Tetranychus turkestani and Tetranychus truncates. Journal of Shihezi University (Natural Science), 31(1), 10–13. (in Chinese)

Yuan, H.X., Zhang, J.P., Yang, X.H., Feng, X.H. & Li, Q. (2008) The comparison on fecundity of *Tetranychus turkestani* and *Tetranychus truncatus*. Acta Arachnologica Sinica, 17(1), 35–38. (in Chinese)

Yu, J.N., Wang, D.Y. & Yuan X.G. (2000) Study on the ecology of laboratory populations and the fecundity life table of *Tetranychus turkestani*. Acta Arachnologica Sinica, 9(2), 78–81. (in Chinese)

Zhang, Y.N., Guo, D.D., Jiang, J.Y.Q., Zhang, Y.J. & Zhang, J.P. (2016) Effects of host plant species on the development and reproduction of *Neoseiulus bicaudus* (Phytoseiidae) feeding on *Tetranychus turkestani* (Tetranychidae). *Systematic and Applied Acarology*, 21(5), 647–656. http://dx.doi.org/10.11158/saa.21.5.6

Submitted: 2 Dec. 2016; accepted by Qing-Hai Fan: 15 Jul. 2017; published: 28 Jul. 2017