

Chlamydia (Psittacosis) Antibody Study in White-tailed Deer 1

Author: DEBBIE, JOHN G.

Source: Bulletin of the Wildlife Disease Association, 3(4) : 152-155

Published By: Wildlife Disease Association

URL: <https://doi.org/10.7589/0090-3558-3.4.152>

The BioOne Digital Library (<https://bioone.org/>) provides worldwide distribution for more than 580 journals and eBooks from BioOne's community of over 150 nonprofit societies, research institutions, and university presses in the biological, ecological, and environmental sciences. The BioOne Digital Library encompasses the flagship aggregation BioOne Complete (<https://bioone.org/subscribe>), the BioOne Complete Archive (<https://bioone.org/archive>), and the BioOne eBooks program offerings ESA eBook Collection (<https://bioone.org/esa-ebooks>) and CSIRO Publishing BioSelect Collection (<https://bioone.org/csiro-ebooks>).

Your use of this PDF, the BioOne Digital Library, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at www.bioone.org/terms-of-use.

Usage of BioOne Digital Library content is strictly limited to personal, educational, and non-commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne is an innovative nonprofit that sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

Chlamydia (Psittacosis) Antibody Study in White-tailed Deer[□]

JOHN G. DEBBIE

Division of Laboratories and Research, New York State Department of Health,
New Scotland Avenue, Albany, New York 12201

ABSTRACT

Deer sera collected during special post-season hunts were tested for group antibody to *Chlamydia* (psittacosis-human pneumonitis) antigen by the complement-fixation test. The deer came from one area in Quebec Province and two areas of New York State. Over 50 percent had complement-fixation titres greater than 1 to 16. Results of these tests are given along with a discussion of their probable implications in wildlife and human health.

Although human illness resulting from a disease of birds has been recognized for over 80 years,¹ only within the last thirty years has disease caused by *Chlamydia* organisms[□] been recognized in domestic animals and wildlife other than avian species. Organisms of the Chlamydial group were isolated from the feces of normal sheep^{8,2,25} and from the feces of normal cattle.^{27,24} Various disease entities in domestic animals caused by these agents are: in cattle, sporadic bovine encephalomyelitis,⁹ pneumonitis,⁷ epizootic abortion^{14,5} and polyarthritides;²¹ in sheep, enzootic abortion,¹⁸ pneumonitis⁸ and polyarthritides;¹⁰ and in pigs, fibrinous pericarditis⁴ and enzootic pneumonia.^{16,22} Agents of this group have been isolated also from wildlife: opossums (*Didelphis marsupialis*),¹³ muskrats (*Ondatra zibethicus*), snowshoe hares (*Lepus americanus*),¹⁷ and fur seals (*Callorhinus ursinus*).³

Storz¹⁹ showed the affinity for placental tissue by members of the Chlamydial group from differing sources by inducing abortion in ewes inoculated with these agents isolated from clinically normal sheep.²⁰ Dungworth and Cordy² induced similar lesions in the lungs of sheep by inoculating fecal, abortion, and pneumonitis strains. Pan¹² induced pneumonitis in pigs with the sheep pneumonitis strain, and Omori et al¹¹ produced pneumonitis lesions in goats and pigs with a bovine encephalomyelitis strain. Dungworth and Cordy² postulated that the agents found in sheep feces are identical to those causing pneumonia and abortion in sheep, and this is supported by the work of Storz.²⁰ Wilson,²³ with a serum neutralization test in eggs, classified ten strains of cattle and sheep Chlamydial agents in three groups: (1) cattle fecal; (2) sheep fecal and sheep pneumonitis; and (3) abortion. How-

□ Presented at the Annual Meeting of the Wildlife Disease Association, University of Illinois, Urbana, Illinois, June 15-16, 1967.

□ The Subcommittee on the Chlamydiae of the Taxonomy Committee of the American Society for Microbiology recently approved the inclusion of the Psittacosis-Lymphogranuloma venereum-Trachoma group of bacterial agents in one genus, *Chlamydia*.

□ Psittacosis-Human Pneumonitis, Markham Laboratories, Chicago, Illinois.

□ Standard positive human sera, Markham Laboratories, Chicago, Illinois.

ever, members of the sheep strains he studied could induce pneumonia or abortion.

There have been no reports of isolations of Chlamydial agents or the detection of complement-fixing antibodies to these organisms in white-tailed deer (*Odocoileus virginianus*). Results of tests of deer sera for *Chlamydia* antibodies with a group antigen are presented here.

MATERIALS AND METHODS

Deer Sera

Deer sera were collected during special post-hunting seasons. The following are general descriptions of the areas from which sera were collected:

Anticosti Island, located at the mouth of the St. Lawrence River is a privately owned island in the Province of Quebec, Canada. It has approximately 3100 square miles and an estimated deer population of 30 to 50 deer per square mile. Serum samples were collected between February 23 and March 19, 1966.

Seneca Army Depot is a United States government installation in central New York State of 15.4 square miles. The estimated deer population is 40 per square mile. Deer were collected on January 8, 1966 and January 7 and 14, 1967.

Lordville is a wintering deer yard situated in Delaware County, New York. The hunting area is about 25 square miles, all of which, however, is not used. It is estimated that in the winter the deer population may be 100 or more per square mile with about 75 percent coming from Pennsylvania. This area was hunted between January 10 and 14, 1966.

Dutchess County, where the pertinent area is known as the Clove Valley, is a wintering deer yard with an approximate ten-mile radius. The hunting area consists of about 40 square miles and the deer population is approximately 40 per square mile. This area was hunted on January 30 and 31, 1967.

Antigen

A commercial group antigen supplied in liquid form¹ was used for the complement-fixation test. It is prepared from eggs inoculated with the human pneumonitis agent.

Complement-Fixation Test

The complement-fixation test was performed by the microtiter method¹⁸ with 0.025 ml. of each reagent. Antisera were heated at 56° C. for 30 minutes and two minimum hemolytic doses of preserved guinea pig complement were used. Fixation of complement was effected at 37° C. for 90 minutes before addition of the hemolytic system.

The controls were a commercial standard control serum¹⁹ and known positive serums kindly supplied by Dr. Storz, Colorado State University. The serum titer was considered as the dilution of serum with which greater than 50 percent hemolysis (2+ fixation) occurred.

RESULTS AND DISCUSSION

The results of the complement-fixation tests for *Chlamydia* antibodies are given in Table 1. Serum samples with anti-complementary reactions were discarded. A complement-fixation titer of 1:16 or higher was considered positive. Serum dilutions were carried out as far as necessary for fixation, but the highest dilution giving a positive result was 1:512.

Isolation of *Chlamydia* agents was not attempted because at the time of collections we were unaware of the high titers to the group antigen. Except for a few deer observed to have greatly enlarged spleens, the animals appeared in fair health. The cause of these enlarged spleens is under investigation.

Since the results of the tests are based on only one serum sample per animal, the state of infectivity is unknown. Also unknown is which agent (or agents) in the group is involved. A significant number of deer sera, however, had high titers to *Chlamydia* antigens.

Little can be said as to comparative antibody titers in the years 1966 and 1967, since the Seneca Army Depot area was the only area sampled in each year. The 23 percent positive sera at Anticosti Island in 1966 has no comparison for 1967. However, the sharp decrease in positive sera at Seneca, from 84.7 percent in 1966 to 31.1 in 1967, suggests some significance in the difference between Lordville in 1966 with 88.4 percent positive sera and Dutchess County in 1967 with 6.4 percent. These latter two areas are in southern New York, 70 miles apart.

There is no apparent significant difference in the number of positive sera by age or sex. The number of positive re-

TABLE 1. Results of the complement-fixation tests for *Chlamydia* antibodies in deer sera collected from Anticosti Island, Quebec and New York State.

	Total Tested	Total Negative	Total Positive	Serum Dilutions				
				1/16	1/64	1/256	1/32	1/128
Anticosti Island, Quebec	74	57 (77%)	17 (23%)	7	1	3	1	3
Lordville, New York	124 (12)*	13 (11.6%)	99 (88.4%)	21	24	15	27	11
Seneca Army Depot 1966	59 (3)	9 (15.3%)	50 (84.7%)	6	7	16	14	7
Seneca Army Depot 1967	154	106 (68.9%)	48 (31.1%)	22	12	11	3	0
Dutchess County, N.Y.	126	118 (93.6%)	8 (6.4%)	4	2	0	2	0

* Numbers in parenthesis refer to sera that were anticomplementary and not tabulated in the results.

actors follows closely the total number of deer taken in each age and sex class in the areas studied in both years.

The presumptive evidence of changes in incidence from year to year and the differences in incidence from one area to another suggest the desirability of continued epizootiologic investigation. Since *Chlamydial* agents have been isolated from feces of clinically normal domestic ruminants, the same factors may play a part in wild ruminants. The antibody titers of sheep and cattle with

enteric infection, however, are usually much lower, rarely above 1:64. The findings reported here may represent a systemic infection not readily noticed by gross postmortem examination, resembling the findings of Wilson and Plummer²⁸ in pigs.

The large number of positive reactors and the wide range of high titers warrants continued investigations of white-tailed deer to determine which agents are involved and what effect these may have on deer, other animals, and man.

ACKNOWLEDGEMENTS

The technical assistance of Mrs. Inez Sherman is greatly appreciated as is the assistance of research personnel of the New York State Conservation Department who organized and directed the special deer hunts. Appreciation is extended to Messrs. Milton Friend and Eugene McCaffrey of the New York State Conservation Department and to the Consolidated Paper Corporation of Canada Limited for providing the deer sera from Anticosti Island, Quebec.

LITERATURE CITED

1. BARRETT, P.K.M. and M.J. GREENBERG. 1966. Outbreak of ornithosis. *Brit. Med. J.*, 2: 206-207.
2. DUNGWORTH, D. L. and D. R. CORDY. 1962. The pathogenesis of ovine pneumonia. II. Isolation of virus from faeces; Comparison of pneumonia caused by faecal, enzootic abortion and pneumonitis viruses. *J. Comp. Path.*, 72: 71-79.
3. EDDIE, B., W. J. L. SLADEN, B. K. SLADEN and K. F. MEYER. 1966. Serological studies and isolation of *Bdsonia* agents from Northern Fur Seals on the Pribilof Islands. *Am. J. Epidem.*, 84: 405-410.
4. GUENOV, I. 1961. Etudes sur la pericardite fibrineuse des porcelets due au virus de l'ornithose. *Bull. Off. Int. Epiz.*, 55: 1465-1473.

5. HOWARTH, J. A., J. E. MOULTON and L. M. FRAZIER. 1956. Epizootic bovine abortion characterized by fetal hepatopathy. J.A.V.M.A., 128: 441-449.
6. KAWAKAMI, T., T. OMORI, S. FUKUHARA, G. TAKUDA, S. ISHII and M. MATUMOTO. 1955. Studies on the disease of cattle caused by a Psittacosis-Lymphogranuloma group virus (Miyagawanella). VII. Isolation of virus, identified as a member of the Psittacosis-Lymphogranuloma group of viruses, from feces of cattle. Jap. J. Exptl. Med., 25: 51-63.
7. MATUMOTO, M., T. OMORI, K. HARADA, Y. INAHA, T. MORIMOTO, R. ISHITANI and S. ISHII. 1955. Studies on the diseases of cattle caused by a Psittacosis-Lymphogranuloma group virus (Miyagawanella). VI. Bovine pneumonia caused by this virus. Jap. J. Exptl. Med., 25: 23-34.
8. McKERCHER, D. G. 1952. A virus possibly related to the Psittacosis-Lymphogranuloma-Pneumonitis group causing a pneumonia in sheep. Science, 115: 543-544.
9. McNUTT, S. H. 1940. A preliminary report on an Infectious Encephalomyelitis of Cattle. Vet. Med., 35: 228-231.
10. MENDLOWSKI, B., W. H. KRAYBILL and D. SEGRE. 1960. Polyarthritides in sheep. II. Characterization of the causative virus. Amer. J. Vet. Res., 21: 74-80.
11. OMORI, T., M. MATUMOTO, Y. KAWAKAMI, S. FUKUHARA and S. ISHII. 1955. Studies on the disease of cattle caused by a Psittacosis-Lymphogranuloma group virus (Miyagawanella). V. Experimental infection of laboratory animals with the isolate from bovine encephalomyelitis. Bull. Nat. Inst. Anim. Health (Japan), 39: 89-97.
12. PAN, I. C., and D. R. CORDY. 1952. The response of swine to an ovine pneumonia virus. Bull. Inst. Zool. Academica Sinica, 1: 29-40.
13. ROCA-GARCIA, M. 1949. Viruses of the Lymphogranuloma-Psittacosis group isolated from Opossums in Colombia. Opossum virus A. J. Infect. Dis., 85: 275-289.
14. SCHOOP, G. and E. GAUKER. 1956. Zur Isolierung von Miyagawanellen aus abortierten Rinderfeten. Deutsch Tierärztl Wschr., 63: 233-235.
15. SEVER, J. L. 1962. Application of a microtechnique to viral serological investigations. J. Immunology, 88: 320-329.
16. SORODOC, G., C. SURDAN and D. SARATENEAU. 1961. Identification of pararickettsia causing enzootic virus pneumonia in pigs. Stud. Cerc. Inframier, 12, Supp.: 355-364.
17. SPALATIN, J., C. E. O. FRASER, R. CONNELL, R. P. HANSON and D. T. BERMAN. 1966. Agents of Psittacosis-Lymphogranuloma venereum group isolated from muskrats and snowshoe hares in Saskatchewan. Can. J. Comp. Med. Vet. Sci., 30: 260-264.
18. STAMP, J. T., A. D. McEWEN, J. A. WATT and D. I. Nisbet. 1950. Enzootic abortion in ewes. I. Transmission of the disease. (Possibly rickettsial infection). Vet. Rec., 62:251-254.
19. STORZ, J. 1961. The diaplacental transmission of Psittacosis-Lymphogranuloma group viruses in Guinea Pigs. J. Infect. Dis., 109: 129-135.
20. STORZ, J. 1963. Superinfection of pregnant ewes latently infected with a Psittacosis-Lymphogranuloma agent. Cornell Vet., 53: 469-480.
21. STORZ, J. 1964. Über eine natürliche infektion eines Meerschweinchenbestandes mit einem erregere aus der Psittakose-Lymphogranuloma-Gruppe. Zentralbl. Bakt., I. Orig. 193: 432-446.
22. SURDAN, C., V. CARAMAN, G. SORODOC and A. BERCAN. 1961. Clinical and pathological study of enzootic virus pneumonia in pigs. Stud. Cerc. Inframier, 12, Supp.: 335-353.
23. WILSON, M. R. 1966. The differentiation of some Psittacosis-Lymphogranuloma venereum group agents isolated from sheep and cattle. J. Infect. Dis., 116: 319-323.
24. WILSON, M. R. 1962. Isolation of Psittacosis-Lymphogranuloma venereum group viruses from the faeces of clinically normal cattle in Britain. Vet. Rec., 74: 1430-1431.
25. WILSON, M. R. and D. L. DUNGWORTH. 1963. Psittacosis-Lymphogranuloma-venereum group viruses in sheep. Comparison between a faecal and an enzootic abortion strain. J. Comp. Path., 73: 277-284.
26. WILSON, M. R. and P. PLUMMER. 1966. A survey of pig sera for the presence of antibodies to the Psittacosis-Lymphogranuloma-venereum group of organisms. J. Comp. Path., 76: 427-433.
27. YORK, C. J. and J. A. BAKER. 1951. A new member of the Psittacosis-Lymphogranuloma group of viruses that causes infection in calves. J. Exptl. Med., 93: 587-603.