# Size-Dependent and Temporal Variability in Batch Number and Fecundity of Red Porgy, a Protogynous, Indeterminate Spawner, in the U.S. South Atlantic 

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# Size-Dependent and Temporal Variability in Batch Number and Fecundity of Red Porgy, a Protogynous, Indeterminate Spawner, in the U.S. South Atlantic 

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#### Abstract

Contemporary studies have shown that spawning stock biomass can be a poor estimator of total egg production, with the inclusion of fecundity tending to improve stock-recruit relationships. Since many U.S. fisheries now use spawning potential ratio (SPR) to define overfishing, an understanding of fecundity variation is especially valuable. We studied a commercially important, protogynous sparid, the Red Porgy Pagrus pagrus, off the southeast U.S. coast from 2007 to 2010 to estimate annual fecundity $\left(f_{a}\right)$ as a function of batch fecundity ( $f_{b}$ ) and batch number ( $n_{b}$ ) and to investigate size-based and temporal variation. In total, 1,355 Red Porgy ( $241-478 \mathrm{~mm}$ TL) were collected; subsets of individuals were selected for estimating maturity, $n_{b}$, and $f_{b}$. Throughout the spawning season, the proportion of females that were capable of spawning increased to $100 \%$ on several dates and was generally higher among smaller females than current maturity information would predict. The proportion of females that were actually spawning was also as high as $\mathbf{1 0 0 \%}$ on several dates, suggesting periods of daily spawning. Batch fecundity was positively related to TL, and the relationship varied significantly between years and among months within years. Spawning fraction (and thus $n_{b}$ ) was also size dependent, with relationships varying between years and dependent upon the reproductive indicator used in calculation (i.e., spawning capability, postovulatory follicle presence, or hydrated oocyte presence). Annual fecundity estimates incorporating size-dependent $\boldsymbol{n}_{\boldsymbol{b}}$ differed from estimates assuming size-independent $\boldsymbol{n}_{\boldsymbol{b}}$ and varied between years. Through its effect on $f_{a}$, the size dependence of $n_{b}$ ultimately affected perceptions of how fishing mortality rate influenced the SPR. Our findings show the importance of accounting for size-dependent variation in $n_{b}$ when calculating $f_{a}$ in indeterminate-spawning species and illustrate the potential implications for stock-recruit relationships and stock exploitation status.


Estimates of spawning stock size are incorporated into both stock-recruit relationships and the calculation of biological reference points, such as the spawning potential ratio (SPR; Table 1; Goodyear 1993; Fitzhugh et al. 2012). The number of eggs produced (population fecundity, $f_{\pi}$ ) is the best measurement of reproductive potential (Hilborn and Walters 1992); however, since $f_{\pi}$ is difficult to estimate, fishery models usually incorporate alternative measures like spawning stock biomass (SSB), which is calculated from total mature biomass or mature female biomass. The use of SSB as a proxy for $f_{\pi}$ in stock
assessments involves several assumptions, including (1) all mature females spawn, (2) $f_{\pi}$ is a linear function of total female biomass, and (3) the relationship between $f_{\pi}$ and female biomass does not vary over time (Marshall et al. 2003). These assumptions can be violated for both biological and methodological reasons.

The assumption that all mature females spawn is violated if mature females skip spawning or if some individuals are simply misidentified as mature. Recent evidence indicates that individuals of many species may periodically skip spawning (Rideout

[^0]TABLE 1. Definitions of frequently used symbols and abbreviations.

| Abbreviation | Description |
| :--- | :--- |
| $d$ | Spawning season duration |
| $f_{a}$ | Fecundity, annual |
| $f_{b}$ | Fecundity, batch |
| $f_{\tau}$ | Fecundity, population |
| HO | Hydrated oocyte |
| $n_{b}$ | Batch number |
| $n_{b, I}$ | Batch number, integral method |
| $p$ | Spawning fraction |
| POF $^{\text {POF }} 0$ | Postovulatory follicle of any age |
|  | Postovulatory follicle(s) less than 1 d after |
| SC | ovulation |
| SPR | Spawning capable |
| SSB | Spawning potential ratio |
| TL | Spawning stock biomass |
|  | Total length at the point of inflection of a |
| TW | logistic function |
|  | Total weight |

and Tomkiewicz 2011); thus, even fish that have spawned previously may not spawn in all future years. Determining whether an inactive fish is mature can be technically difficult, so the error rate when assigning maturity phases to inactive fish is likely to be high. Even histological techniques, which are widely accepted as the most accurate methods for determining maturity, cannot usually predict skipped spawning in individuals that have been captured outside of the spawning season. This is because reliable indicators of past or future spawning may not be present at times outside of the main reproductive season. The best way to determine the relationship between fish size and involvement in spawning is to assess maturity either immediately prior to or during the active spawning season, when all females that are about to spawn show clear signs of development and when no (or very few) individuals have completed spawning and regressed (Hunter and Macewicz 2003).

When SSB is used as a direct proxy for reproductive output, $f_{\pi}$ is assumed to be a linear function of the total weight of all females in the stock. This assumes that a given mass of mature females will always produce the same number of eggs, regardless of the size distribution of individuals. If the true relationship is instead curvilinear, the assumption is violated. If, for example, individual fecundity is a power function of weight, then the use of SSB as a proxy for $f_{\pi}$ could simultaneously overestimate and underestimate egg production of the smallest and largest females, respectively (Marshall et al. 2006).

Temporal instability in the relationship between individual fecundity and female body size can also introduce error into the relationship between $f_{\pi}$ and SSB , thereby introducing error into the stock-recruit relationship (Marshall et al. 2003). Despite the general paucity of fecundity data, even for many well-studied species in the Northwest Atlantic (Tomkiewicz et al. 2003),
extensive studies on Atlantic Cod Gadus morhua suggest that temporal variation in fecundity is rather common (Kjesbu et al. 1998; Kraus et al. 2000; McIntyre and Hutchings 2003; Yoneda and Wright 2004; Klibansky 2006). In some instances, variable egg production has been correlated with food availability or water temperature; however, simple environmental correlates have mostly failed to predict observed fluctuations in fecundity (Lambert et al. 2003). Instead, current research has generally been focused on developing simpler methodologies that will allow fecundity-body size relationships to be estimated routinely (Witthames et al. 2009; Armstrong and Witthames 2012).

Estimation of fecundity is often difficult since most fish produce large numbers of small eggs, but it can be particularly challenging to estimate fecundity for species with indeterminate fecundity (hereafter, "indeterminate species"). For such species, fecundity is not fixed prior to the start of spawning, so oocyte counts for individual females do not accurately reflect annual fecundity $\left(f_{a}\right)$. To estimate $f_{a}$ in indeterminate species, the number of batches $\left(n_{b}\right)$ of eggs produced by a female during the season is multiplied by batch fecundity $\left(f_{b}\right)$, which is the number of oocytes spawned in a single batch (Kjesbu 2009). Typically, $n_{b}$ is calculated as the product of spawning season duration $(d)$ and the spawning fraction $(p)$, which is estimated as the proportion of mature females that are found to possess reproductive indicators, such as hydrated oocytes (HOs) or postovulatory follicles (POFs), when caught during the spawning season (Murua et al. 2003). As with $f_{a}$ for determinate species (Duarte and Alcaraz 1989), $f_{b}$ often scales positively with body size among indeterminate species (Harris et al. 2002; McBride and Thurman 2003; Ganias et al. 2004). Although it is commonly assumed that $n_{b}$ does not vary with size or age, a recent review showed that most studies ( 32 of 41) investigating this issue found that spawning frequency, and thus $n_{b}$, varied with size or age (Fitzhugh et al. 2012). In some cases, variation in $n_{b}$ can be the primary cause of variable $f_{a}$ (Collins et al. 1996; Wootton and Fletcher 2009). Therefore, studies estimating $f_{a}$ for indeterminate species should determine the extent of variation in $n_{b}$ as well as $f_{b}$.

The Red Porgy Pagrus pagrus is a protogynous hermaphrodite (Alekseev 1982) that has been shown to be indeterminate (Daniel 2003). The species is found on reefs along continental shelf margins and is broadly distributed in the Atlantic, with three genetically distinct populations (Ball et al. 2003). In the U.S. South Atlantic Bight (SAB), Red Porgy support an economically important fishery that ranked 11th in total (1985-2008) landings among 73 species included in the snapper-grouper management complex (National Oceanic and Atmospheric Administration [NOAA], fisheries statistics; www.st.nmfs.noaa.gov/st1). The SAB stock is considered overfished, and current landings are an order of magnitude lower than landings observed two decades ago. Macroscopic staging and gonadosomatic index data suggest that the main spawning season for Red Porgy in the SAB occurs from January to March (Manooch 1976). Previous histological staging data revealed a decrease in length at maturity between 1979 and 1994 (Harris
and McGovern 1997); age at $50 \%$ maturity was recently estimated at 1.5 years, and size at $50 \%$ maturity was estimated at 289 mm TL (Daniel 2003). A positive relationship between $f_{b}$ and TL has also been noted previously (Daniel 2003).

To date, the vast majority of data used to determine ageand size-dependent maturity relationships for Red Porgy in the SAB have been collected during periods outside of the spawning season, and this may have compromised accuracy in some cases. Further, no authors have investigated the relationship between $n_{b}$ and body size or evaluated the level of interannual variation in either $f_{b}$ or $n_{b}$. Because the basic reproductive biology of Red Porgy is very similar to that of many other members of the snapper-grouper complex, an improved understanding of the factors influencing egg production in Red Porgy has the potential to benefit the management of multiple valuable fishery resource species.

## METHODS

Field collections.-We sampled the SAB stock of Red Porgy during the spawning season over three consecutive years (20072008, 2008-2009, and 2009-2010); however, in 2007-2008, sampling did not begin until midseason. In the latter 2 years (hereafter, "the complete years"), port sampling began in October and occurred regularly until females with spawning-capable (SC) ovaries (i.e., with vitellogenic oocytes that were visible macroscopically) became abundant in the samples, which triggered the start of at-sea sampling. In 2007-2008, all samples were collected through port sampling. In both 2008-2009 and 2009-2010, we conducted at-sea sampling with local commercial hook-and-line fishers throughout the spawning season. When evidence of spawning activity was minimal, at-sea sampling ceased and port sampling resumed for the remainder of the season. Given resource constraints, sampling was designed to maximize temporal resolution within a limited geographic range offshore from Onslow Bay and Long Bay, North Carolina (Figure 1). Since 2000, $84 \%$ of SAB Red Porgy commercial landings have been taken off the coasts of South Carolina and North Carolina (NOAA, fisheries statistics; www.st.nmfs.noaa.gov/st1). Therefore, we felt that the spatial extent of our sampling was sufficient to ensure that the implications of our findings would be broadly applicable to the stock, and we chose to focus our efforts on obtaining an improved understanding of temporal variation.

Subsampling and laboratory analysis.-We measured TL for all fish sampled. For a large subset ( $92 \%$ ) of fish, sex and maturity were determined macroscopically, and total weight (TW), gutted weight (GW), and gonad weight were each measured before preserving the gonads in a $10 \%$ solution of buffered formalin. A group of these fish-from a roughly even distribution of sample dates throughout all seasons-was then selected for histological analysis. Approximately 20 individuals were selected randomly from among all fish caught on each date. For these fish, tissue from the posterior region of the gonad was embedded in paraffin, sectioned, mounted, and stained (Gill's hematoxylin


FIGURE 1. Spatial distribution of directed trip sampling locations for Red Porgy during 2008-2009 and 2009-2010. The small rectangle in the inset shows the area covered by the larger map.
and eosin Y) by staff at the South Carolina Department of Natural Resources (SCDNR). Sex and maturity were determined based on criteria (developed by SCDNR staff) that are used routinely for reproductive analysis of various species as part of the Marine Resources Monitoring, Assessment, and Prediction program. An abbreviated version of this key was presented by Harris and McGovern (1997), whereas a more complete and updated version was provided by Daniel (2003). We note, however, that during our study, traces of atresia could be present but were not necessary for a female to be designated as regenerating (i.e., mature but inactive). Females that exhibited abundant vitellogenic oocytes with less than $50 \%$ atresia were deemed to have entered their spawning period and were considered SC. When vitellogenic oocytes were present, they were usually abundant ( $>200$ per histology section) but sometimes were rare (1-10 per histology section); females characterized by the latter were not considered SC. We felt that excluding individuals with only rare vitellogenic oocytes produced a more accurate index of true spawning capability than simple presence/absence of vitellogenic oocytes. Reproductive terminology used here largely follows that presented by Brown-Peterson et al. (2011).

Discrimination of postovulatory follicle age.-Although POF degeneration in Red Porgy generally agreed with descriptions provided by Hunter and Macewicz (1985) and although the sea surface temperatures we recorded during the Red Porgy spawning season $\left(14.7-19.0^{\circ} \mathrm{C}\right)$ were similar to what
those authors reported for Pacific Sardine Sardinops sagax and Northern Anchovy Engraulis mordax (13-19 ${ }^{\circ}$ C), we devised a procedure to help discriminate day- $0 \mathrm{POFs}\left(\mathrm{POF}_{0}\right)$ that were less than 24 h old from older POFs. Discrimination was possible because many Red Porgy ovaries contained two consecutive daily POF cohorts. First, we identified females collected near the daily spawning peak that contained both a cohort of very recent POFs and a cohort of POFs that appeared similar to published descriptions of 24-h-old POFs (Hunter and Macewicz 1985). We then took images of POFs from both cohorts and used them as reference images to define the morphological endpoints of POF degradation during a 24-h period (Figure 2). In subsequent samples, POFs that were morphologically intermediate between these two endpoints were assigned to the day- 0 cohort $\left(\mathrm{POF}_{0}\right)$. A similar procedure was not reliable for discriminating 48-h-old POFs, and temporal discrimination was also not possible for HOs; thus, we considered $\mathrm{POF}_{0}$ to be the most accurate reproductive indicator for Red Porgy.

Estimation of batch fecundity.-For each preserved ovary, we used a syringe to remove subsamples $(\sim 0.2 \mathrm{~g})$ of tissue from the anterior and posterior regions and stored them in vials containing $10 \%$ buffered formalin. Procedures to count oocytes in the most advanced cohort (including germinal vesicle migration [GVM], germinal vesicle breakdown [GVBD], and/or HOs ) generally followed Klibansky and Juanes (2008). Females containing very recent POFs were excluded from $f_{b}$ analysis. Images were captured with a flatbed scanner (Epson Perfection V500) using predefined settings ( $8.9 \times 8.9 \mathrm{~cm}$; 8-bit gray scale at $2,400 \mathrm{dpi} ; 8,400 \times 8,400$ pixels; uncompressed TIFF; 67.2megabyte file size). The varied size and appearance of GVM, GVBD, and HOs in images resulted in poor automated counts, so we manually counted oocytes by using the Cell Counter plug-in, which took very little time ( $5-10 \mathrm{~min} / \mathrm{sample}$ ).

Statistical analysis.-We used logistic regression to examine the size dependence of several binary response variables for Red Porgy, including maturity, sex, spawning capability, $\mathrm{POF}_{0}$ presence, and HO presence. For maturity and sex, we used data from all dates to develop maturation and sex transition ogives. For the latter three variables, we only used data collected within the SC season to calculate size-dependent $n_{b}$. To test for interannual differences in these relationships, we first fit a logistic model that included terms for the intercept, slope, year, and the TL $\times$ year interaction ( $\beta_{0}, \beta_{1}, \beta_{2}$, and $\beta_{3}$, respectively) to validate the assumption of no interaction. If $\beta_{3}$ was not significant, we then fit the model without $\beta_{3}$ to test the significance of $\beta_{2}$ (Hosmer and Lemeshow 1989; Dalgaard 2008). If either term was significant, we concluded that the models differed between years. For each significant two-parameter model, we calculated the point of inflection (TL ${ }_{50}$ ) as $\beta_{0} /-\beta_{1}$ (Jennings et al. 2001). We used linear regression to investigate the relationship between $f_{b}, \mathrm{GW}$, TL, and age. Analysis of covariance (ANCOVA) was used to test for interannual and intermonth differences in $f_{b}$ when withingroup regression slopes were homogeneous. After performing ANCOVA, we calculated relative $f_{b}$ as $\left[\log _{e}\left(f_{b}\right)\right] /(\mathrm{mm} \mathrm{TL})$ to


FIGURE 2. Postovulatory follicles (POFs) from two consecutive cohorts in the same female Red Porgy, defining the morphological endpoints of POF degeneration within the first 24 h of ovulation; examples of the largest and newest POFs from (A) the POF cohort that just appeared after spawning ( 0 h old) and (B) the previous cohort ( 24 h old) are shown $($ scale bar $=100 \mu \mathrm{~m}$ ).
visually compare $f_{b}$ adjusted for body size among months within 2008-2009 and 2009-2010. All statistical analyses were conducted using the base package of R version 2.14.0.

Calculations.-Although our calculations largely follow established standards, we provide details below for clarity. We estimated $f_{b}$ via the method of Hunter et al. (1985) as the number of GVM, GVBD, and HOs per gram of tissue (averaged for two samples) multiplied by the formalin wet weight of both ovaries. We calculated $n_{b}$ as the product of $p$ and $d$ (Murua et al. 2003). We calculated $p$ as the number of spawning females divided by the number of SC females (Hunter and Goldberg 1980). We estimated $p$ separately by using the presence of either $\mathrm{POF}_{0}$ or HOs as the reproductive indicator. We present both estimates to enable comparison with other studies, but we considered $p$ based on $\mathrm{POF}_{0}$ to be more accurate since the stage duration could be
calibrated to 24 h . We estimated $d$ as the number of days between the first and last sampling dates on which at least one female was detected with a reproductive indicator; three separate estimates of $d$ were obtained using spawning capability, HO presence, or $\mathrm{POF}_{0}$ presence as the reproductive indicator. The separate estimates of $p$ and $d$ were used to explore how the selection of HOs or $\mathrm{POF}_{0}$ as the reproductive indicator influenced the calculation of $n_{b}$.

Estimates of $n_{b}$ calculated by using the standard approach ( $p \times d$ ) will only be accurate when the initial date and duration of all spawning periods are the same for all females, thereby producing a distribution of $p$ that is uniform across time. Since a strictly uniform distribution is unlikely, the method of piecewise integration used by Hunter and Leong (1981) should be more accurate. We estimated $n_{b}$ using the integral method $\left(n_{b, I}\right)$ as
$n_{b, I}=\sum_{i=1}^{z-1} \frac{\left|p_{i+1}-p_{i}\right|\left(t_{i+1}-t_{i}\right)}{2}+\min \left(p_{i+1}, p_{i}\right)\left(t_{i+1}-t_{i}\right)$,
where $z$ is the total number of sampling dates, $p_{i}$ is the estimate of spawning fraction for the $i$ th sampling date, $\min ()$ is a function indicating that only the minimum of the set of values in parentheses should be used, and $t_{i}$ is the ordinal date for the $i$ th sampling date. We completed calculations separately for each year and also for the combined data from the two complete sampling years.

Standard calculations of $p$ essentially fit a horizontal-line model to $\mathrm{POF}_{0}$ presence or HO presence and TL, leading to a size-independent estimate of $n_{b}$. Building on the work of Collins et al. (1998), who calculated $n_{b}$ for discrete age-groups, and Lowerre-Barbieri et al. (2009), who used logistic regression to test for a relationship between binary reproductive indicators and TL, we calculated three types of size-dependent $n_{b}$ estimate.

For the first two types, we tested for significant logistic fits between $\mathrm{POF}_{0}$ or HO presence and TL in females collected during the spawning season based on spawning capability (hereafter, "SC season"). We used significant models to estimate size-dependent $p\left(p_{\mathrm{TL}}\right)$ and then calculated size-dependent $n_{b}$ as the product of $p_{\mathrm{TL}}$ and $d$, yielding separate estimates for $\mathrm{POF}_{0}$ presence and HO presence. For the third type of estimate, we fit the logistic model to spawning capability and TL and used significant relationships to estimate the size-dependent proportion of $d$ for which females were $\mathrm{SC}\left(\mathrm{P}[d]_{\mathrm{TL}}\right)$. Multiplying $\mathrm{P}(d)_{\mathrm{TL}}$ by $d$ yields a size-dependent estimate of $d\left(d_{\mathrm{TL}}\right)$; multiplying $d_{\mathrm{TL}}$ by size-independent $p$ (based on $\mathrm{POF}_{0}$ ) produced the third type of size-dependent estimate of $n_{b}$.

Since Red Porgy are indeterminate, $f_{a}$ is estimated as the product of $f_{b}$ and $n_{b}$. We estimated the size-dependent $f_{b}$ by using linear regression as described above; we then multiplied the size-dependent $f_{b}$ by the size-independent $n_{b, I}$ and each of the three size-dependent estimates of $n_{b}$, thus producing a total of four types of $f_{a}$ calculation. We then converted TL to TW
based on a TL-TW relationship (SEDAR 2006) to examine the relationship between $f_{a}$ and biomass.

To investigate how different calculations of $f_{a}$ affect the perceived impact of fishing on reproductive potential, we calculated SPR in several ways. Goodyear (1993) defined SPR as potential fecundity per recruit $(f / R)$ in the presence of fishing divided by the $f / R$ in the absence of fishing. We calculated SPR over a range of fishing mortalities by using the four types of $f_{a}$ to estimate $f / R$ and then separately by using $\mathrm{SSB} / R$ based on mature female biomass, a standard proxy for $f / R$. Calculations of SPR followed the method of Gabriel et al. (1989) and used parameter estimates from this study and from the most recent period (1999-2004) described in the 2006 stock assessment (SEDAR 2006).

## RESULTS

## Sex and Maturity by Size

In total, 1,355 Red Porgy were returned to the laboratory and examined macroscopically. From these, 823 fish were selected for histological analysis (Table 2). The logistic regression relating sex to TL for the two complete reproductive seasons that we sampled (2008-2009 and 2009-2010) revealed a significant interaction ( $P=0.04, n=636$; Table 3 ) between size and year effects and thus interannual variation in the size at sex transition. Within each year, sex was significantly related to TL $(P<$ $0.001, n=341$ for 2008-2009 and 295 for 2009-2010). All of the Red Porgy females that we encountered were mature; thus, TL at $100 \%$ maturity was not greater than 241 mm TL-the minimum size of Red Porgy females sampled in our study.

## Spawning Capability

Since sampling in 2007-2008 began in the middle of the SC season, $d$ based on spawning capability was not calculated for that year. For the 2008-2009 and 2009-2010 seasons, $d$ based on spawning capability was estimated to be 159 and 165 d , respectively (Table 4), with evidence for a single SC season in each year (Figure 3).

The logistic regression relating spawning capability to TL for the two complete reproductive seasons detected a highly significant interaction ( $P<0.001, n=442$; Table 3) between size and year effects. When examined separately for each year, spawning capability was not related to TL in 2008-2009 ( $P=$ $0.814, n=224$ ) but was positively related to TL in 2009-2010 ( $P<0.001, n=218$; Table 3; Figure 4).

## Spawning Fraction and Batch Number

Several estimates of $p, d$, and $n_{b}$ were generated by using the different reproductive indicators (Table 4). In 2007-2008, the presence of $\mathrm{POF}_{0}$ yielded an estimate of $p$ equal to 0.20 and the presence of HOs yielded a $p$-estimate of 0.34 ; however, since sampling began in the middle of the SC season, $d$ and $n_{b}$ could not be determined (Figure 3). In 2008-2009, $\mathrm{POF}_{0}$ presence yielded an $n_{b, I}$ estimate of 82 , and HO presence yielded an $n_{b, I}$

TABLE 2. Total number of Red Porgy that were phased macroscopically; the number of fish included in the histologically phased subsample by year, month, and sex; and the TL range for each group. Females are further delineated by the presence of reproductive indicators; individual spawning-capable (SC) females may contain day- 0 postovulatory follicles $\left(\mathrm{POF}_{0}\right)$, hydrated oocytes $(\mathrm{HO})$, both, or neither.

| Data set | Month | Total |  |  | Subsample |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  | TL (mm) | Males | Females | TL (mm) | Males | Females | SC | $\mathrm{POF}_{0}$ | HO |
| 2007-2008 | Dec | 361-435 | 16 | 11 | 361-435 | 12 | 8 | 8 | 2 | 3 |
|  | Feb | 292-471 | 32 | 49 | 292-471 | 23 | 36 | 36 | 7 | 7 |
|  | Mar | 303-432 | 27 | 12 | 303-432 | 27 | 12 | 12 | 2 | 9 |
|  | Apr | 326-410 | 6 | 9 | 326-410 | 6 | 9 | 3 | 0 | 0 |
|  | May | 308-370 | 11 | 9 | 308-370 | 13 | 7 | 0 | 0 | 0 |
|  | Jun | 351-441 | 5 | 17 | 351-441 | 6 | 14 | 0 | 0 | 0 |
|  | All | 292-471 | 97 | 107 | 292-471 | 87 | 86 | 59 | 11 | 19 |
| 2008-2009 | Oct | 347-437 | 2 | 3 | 347-437 | 2 | 3 | 0 | 0 | 0 |
|  | Nov | 308-431 | 24 | 20 | 308-431 | 24 | 19 | 12 | 1 | 4 |
|  | Dec | 320-443 | 25 | 34 | 320-443 | 19 | 24 | 24 | 6 | 20 |
|  | Jan | 257-454 | 40 | 97 | 257-409 | 14 | 73 | 73 | 46 | 13 |
|  | Feb | 243-455 | 38 | 106 | 274-420 | 22 | 56 | 53 | 44 | 12 |
|  | Mar | 273-415 | 22 | 40 | 273-415 | 22 | 40 | 35 | 31 | 20 |
|  | Apr | 305-393 | 10 | 18 | 305-393 | 10 | 17 | 3 | 1 | 1 |
|  | All | 243-455 | 161 | 318 | 257-443 | 113 | 232 | 200 | 129 | 70 |
| 2009-2010 | Oct | 317-417 | 6 | 13 |  | 0 | 0 | 0 | 0 | 0 |
|  | Nov | 284-387 | 3 | 30 | 296-387 | 2 | 17 | 6 | 0 | 0 |
|  | Dec | 308-373 | 11 | 16 | 308-373 | 8 | 12 | 9 | 0 | 0 |
|  | Jan | 241-457 | 32 | 227 | 241-457 | 18 | 88 | 86 | 81 | 72 |
|  | Feb | 245-462 | 26 | 72 | 265-462 | 10 | 30 | 28 | 26 | 18 |
|  | Mar | 252-478 | 49 | 105 | 252-416 | 23 | 49 | 16 | 13 | 11 |
|  | Apr | 282-435 | 38 | 43 | 282-426 | 16 | 23 | 9 | 2 | 0 |
|  | All | 241-478 | 165 | 506 | 241-462 | 77 | 219 | 154 | 122 | 101 |

estimate of 54; in 2009-2010, the $n_{b, I}$ estimates were 98 and 76, significant interaction ( $P<0.001, n=442$ ) between size respectively.

The logistic regression relating $\mathrm{POF}_{0}$ presence to TL for the complete years (both years combined) produced a highly and year effects. The response of $\mathrm{POF}_{0}$ presence to TL differed between years, being significantly negative ( $P<0.001$, $n=224$ ) in 2008-2009 and significantly positive ( $P<0.001$,

TABLE 3. Results from logistic models relating binary response variables to Red Porgy TL (mm). Responses are the proportion of all individuals that were male (Male) and the proportion of females that were spawning capable (SC), possessed day-0 postovulatory follicles ( $\mathrm{POF}_{0}$ ), or possessed hydrated oocytes (HO). Model terms for the intercept $\left(\beta_{0}\right)$, slope $\left(\beta_{1}\right)$, year $\left(\beta_{2}\right)$, and TL $\times$ year interaction $\left(\beta_{3}\right)$ are presented; the interaction ( $\beta_{3}$ ) was only evaluated for the multiyear data set $\left(\mathrm{TL}_{50}=\right.$ TL at the point of inflection, calculated as $\left.\beta_{0} /-\beta_{1}\right)$. The $P$-values are for the last parameter listed for each model.

| Data set | Response | $\beta_{0}$ | $\beta_{1}$ | $\beta_{2}$ | $\beta_{3}$ | $\mathrm{TL}_{50}$ | $P$ | $n$ |
| :--- | :--- | ---: | ---: | ---: | ---: | ---: | ---: | ---: |
| $2008-2009$ and 2009-2010 | Male | -9.64 | 0.0255 | -6.07 | 0.01 |  | 0.040 | 636 |
|  | SC | 2.73 | -0.0016 | -12.87 | 0.03 |  | $<0.001$ | 442 |
|  | POF $_{0}$ | 3.53 | -0.0096 | -11.71 | 0.03 |  | $<0.001$ | 442 |
| $2008-2009$ | HO | -3.17 | 0.0070 | -3.91 | 0.01 |  | 0.044 | 442 |
|  | Male | -9.39 | 0.0248 |  |  | 378 | $<0.001$ | 341 |
|  | SC | 2.72 | -0.0016 |  |  | 372 | 0.814 | 224 |
|  | POF | 3.29 | -0.0088 |  |  | 0.026 | 224 |  |
|  | HO | -2.64 | 0.0055 |  |  | 393 | $<0.001$ | 295 |
|  | Male | -15.71 | 0.0400 |  | 313 | $<0.001$ | 218 |  |
|  | SC | -10.47 | 0.0334 |  | 335 | $<0.001$ | 218 |  |
|  | POF |  | -8.26 | 0.0246 |  |  | 355 | $<0.001$ |
|  | HO | -7.09 | 0.0200 |  |  | 218 |  |  |

TABLE 4. Red Porgy spawning fraction ( $p$ ), spawning season duration ( $d$; number of days), and batch number ( $n_{b}$ ) estimates for each data set and each reproductive indicator (indicators: $\mathrm{SC}=$ spawning capability; $\mathrm{POF}_{0}=$ presence of day- 0 postovulatory follicles; $\mathrm{HO}=$ presence of hydrated oocytes). Estimates of $n_{b}$ calculated by using the integral method $\left(n_{b, I}\right)$ are also presented.

| Data set | $p$ |  | $d$ |  |  | $n_{b}$ |  | $n_{b, I}$ |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | $\mathrm{POF}_{0}$ | HO | SC | $\mathrm{POF}_{0}$ | HO | $\mathrm{POF}_{0}$ | HO | $\mathrm{POF}_{0}$ | HO |
| 2007-2008 | 0.20 | 0.34 |  |  |  |  |  |  |  |
| 2008-2009 | 0.72 | 0.36 | 159 | 124 | 147 | 89 | 53 | 82 | 54 |
| 2009-2010 | 0.88 | 0.79 | 165 | 106 | 71 | 93 | 56 | 98 | 76 |
| 2008-2009 and 2009-2010 | 0.79 | 0.51 | 169 | 134 | 147 | 106 | 75 | 86 | 68 |

$n=218$ ) in 2009-2010 (Table 3; Figure 4). The logistic regression relating HO presence to TL for the complete years (combined) also produced a significant interaction ( $P=0.044$, $n=442$ ) between size and year effects. The relationship between HO presence and TL was not significant for 2008-2009 ( $P=0.127, n=224$ ) but was positive and highly significant ( $P<0.001, n=218$ ) for 2009-2010 (Table 3; Figure 4).

## Batch Fecundity

When data were pooled across all three years, TL was a better predictor of $f_{b}\left(r^{2}=0.142\right)$ than was GW $\left(r^{2}=0.112\right)$. Natural log transformation of $f_{b}$ removed heteroscedasticity and improved both fits ( $r^{2}=0.155$ and 0.136 , respectively; Table 5). Since TL and $\log _{e}\left(f_{b}\right)$ had the strongest relationship, we used


FIGURE 3. Proportion of female Red Porgy on each sampling date that were spawning capable (SC), possessed day-0 postovulatory follicles $\left(\mathrm{POF}_{0}\right)$, or possessed hydrated oocytes (HO) during (A) 2007-2008, (B) 2008-2009, and (C) 2009-2010. Tick marks along the $x$-axis indicate the first day of each month.
these variables in further analyses. The ANCOVA revealed a significant effect of year on $f_{b}$ when all years were included ( $P<0.001, n=285$; Table 6; Figure 5). Year remained a significant factor when only the two complete sampling years were included ( $P=0.002, n=276$; Table 6 ). Separate ANCOVA models for each year detected significant effects of month within 2008-2009 ( $P<0.001, n=82$ ) and 2009-2010 $(P<0.001, n=$ 193; Table 6). Box plots showed that relative $f_{b}$ was higher in the middle of the season and lower during earlier and later portions of the season (Figure 6). Variation in TL explained much higher proportions of variation in $\log _{e}\left(f_{b}\right)$ for several monthly data sets than for the data set that included all years (Table 5). The two highest $r^{2}$ values were observed for January $2010\left(r^{2}=0.344\right.$, $n=140)$ and February $2010\left(r^{2}=0.427, n=36\right)$.


FIGURE 4. Presence or absence of reproductive traits as a function of body size for female Red Porgy sampled in (A) 2008-2009 and (B) 2009-2010. Lines are fitted logistic functions; points are observed proportions (within size-bins) of spawning-capable females (SC), females that possessed day-0 postovulatory follicles $\left(\mathrm{POF}_{0}\right)$, or females that possessed hydrated oocytes $(\mathrm{HO})$.

TABLE 5. Batch fecundity $\left(f_{b}\right)$ regressions for Red Porgy in each data set. Models estimating $\log _{e}\left(f_{b}\right)$ are presented in exponential form (GW $=$ gutted weight, $\mathrm{g} ; \mathrm{TL}=$ total length, $\mathrm{mm} ; r^{2}=$ coefficient of determination). The $P$-values are for the regression slope in each model.

| Data set | Predictor | Model | $\beta_{0}$ | $\beta_{1}$ | $P$ | $r^{2}$ | $n$ |
| :--- | :---: | :--- | :---: | ---: | ---: | ---: | ---: |
| All years | GW | Linear | $-5,100$ | 26.798 | $<0.001$ | 0.112 | 275 |
| All years | TL | Linear | $-31,869$ | 148.24 | $<0.001$ | 0.142 | 285 |
| All years | GW | Exponential | 8.83 | 0.00159 | $<0.001$ | 0.136 | 275 |
| All years | TL | Exponential | 6.79 | 0.00835 | $<0.001$ | 0.155 | 285 |
| 2008-2009 and 2009-2010 | TL | Exponential | 6.58 | 0.00891 | $<0.001$ | 0.172 | 276 |
| 2007-2008 | TL | Exponential | 6.19 | 0.01190 | 0.029 | 0.518 | 9 |
| $2008-2009$ | TL | Exponential | 6.71 | 0.00902 | $<0.001$ | 0.141 | 83 |
| 2009-2010 | TL | Exponential | 6.14 | 0.00995 | $<0.001$ | 0.234 | 193 |
| Nov 2008 | TL | Exponential | 11.40 | -0.00610 | 0.458 | 0.293 | 4 |
| Dec 2008 | TL | Exponential | 3.97 | 0.01570 | 0.007 | 0.295 | 23 |
| Jan 2009 | TL | Exponential | 8.47 | 0.00471 | 0.227 | 0.232 | 8 |
| Feb 2009 | TL | Exponential | 7.64 | 0.00767 | 0.012 | 0.198 | 31 |
| Mar 2009 | TL | Exponential | 5.40 | 0.01140 | 0.106 | 0.176 | 16 |
| Jan 2010 | TL | Exponential | 5.64 | 0.01100 | $<0.001$ | 0.344 | 140 |
| Feb 2010 | TL | Exponential | 5.61 | 0.01290 | $<0.001$ | 0.427 | 36 |
| Mar 2010 | TL | Exponential | 7.58 | 0.00510 | 0.576 | 0.021 | 17 |

${ }^{\text {a }}$ All observations for this data set are from March 2008.

## Annual Fecundity and Spawning Potential Ratio

In 2008-2009, estimated $f_{a}$ that incorporated the sizedependent $n_{b}$ based on the negative relationship between $\mathrm{POF}_{0}$ presence and TL was slightly concave-downward relative to TW. This contrasted with the estimated $f_{a}$ that incorporated sizeindependent $n_{b}$, which demonstrated an exponential increase with TW and was about $200 \%$ higher for the largest females (Figure 7A). The SPR was higher when calculated using the estimated $f_{a}$ based on a size-dependent $n_{b}$ than when $n_{b}$ was assumed to be independent of fish size (Figure 8A).

In 2009-2010, estimates of $f_{a}$ that incorporated sizedependent $n_{b}$ based on the positive relationship between SC, $\mathrm{POF}_{0}$ presence, or HO presence and TL all showed positive exponential relationships with TW. The estimate of $f_{a}$ that incorporated size-independent $n_{b}$ was also positively related to TW but was $20-30 \%$ lower for the largest females (Figure 7B). The SPR was lower when calculated with $f_{a}$ estimates based on a size-dependent $n_{b}$ than when no size dependence of $n_{b}$ was assumed (Figure 8B).

TABLE 6. Results from ANCOVA modeling of the $\log _{e}\left(f_{b}\right)$ of Red Porgy as a function of year or month, with TL $(\mathrm{mm})$ as the covariate $\left(P_{\text {Slopes }}=P\right.$-value for the test of slope homogeneity; $P_{\text {Group }}=P$-value for the test of adjusted means).

| Data set | Group | $P_{\text {Slopes }}$ | $P_{\text {Group }}$ | $n$ |
| :--- | :--- | :---: | ---: | ---: |
| All years | Year | 0.915 | $<0.001$ | 285 |
| 2008-2009 and 2009-2010 | Year | 0.711 | 0.002 | 276 |
| 2008-2009 (Nov-Mar) | Month | 0.445 | $<0.001$ | 82 |
| $2009-2010$ | Month | 0.542 | $<0.001$ | 193 |

## DISCUSSION

## Red Porgy Reproductive Output

This study represents the first investigation in which the size at maturity for the SAB Red Porgy stock was based on large sample sizes collected throughout the spawning season. Our


FIGURE 5. Scatter plot of the natural logarithm (ln) of Red Porgy batch fecundity versus TL for each sampling year.


FIGURE 6. Box plots of $\log _{e}\left(f_{b}\right) \mathrm{TL}^{-1}$ for Red Porgy in each sampling month during (A) 2008-2009 and (B) 2009-2010. Box color corresponds to month. The line within each box represents the median, the lower and upper box ends represent the 1st and 3rd quartiles, and the end of each whisker extends to the most extreme observation within 1.5 times the interquartile range below or above the box. Open circles represent observations outside the whiskers.
findings show that females spawn at smaller body sizes than previous estimates suggest. Prior to this study, the $\mathrm{TL}_{50}$ for mature females was estimated separately at 289 mm (Daniel 2003) and 272 mm (SEDAR 2006); however, all of the females that we sampled were mature, including fish as small as 241 mm TL. Since we did not encounter any immature Red Porgy, we could not determine $\mathrm{TL}_{50}$ for mature females, but our estimate of $\mathrm{TL}_{50}$ for SC females was 255 mm based on both complete years combined. The size when spawning capability is attained should represent an upper estimate of $\mathrm{TL}_{50}$ for mature females since some mature fish may not yet have entered their spawning periods. During each year, we also observed a period in which all of the females that we encountered were SC (see Figure 3A-C), suggesting that $100 \%$ of females above the minimum TL during those periods spawned. It is important to note that encountering all fish in SC condition also indicates that mature females were unlikely to have skipped spawning.

Since we conducted sampling in and around the spawning season, we were more able to determine maturity than previous studies in which samples were mainly collected after the spawning season, when most mature females were in a regenerating state (Harris and McGovern 1997; Daniel 2003; SEDAR 2006). As was noted by Hunter and Macewicz (2003) in a review of methodologies, even histology cannot always distinguish between regenerating and immature females. In contrast, maturity is often unmistakable during the spawning season, when mature ovaries contain more developed oocytes. Aside from the technical difficulty in distinguishing between regenerating and immature females, the validation of characteristics that are used to make this distinction is strongly lacking. Maturity staging of Red Porgy in this study and other recent
studies (Daniel 2003; SEDAR 2006) was largely based on the key developed by Harris and McGovern (1997). However, none of the sources cited in the development of this key actually demonstrated that the characteristics used to distinguish regenerating females from immature females were indicative of maturity. The most relevant of these sources provided a description of gonads from known-age Red Seabream Pagrus major (Matsuyama et al. 1988b), but that study neither demonstrated nor suggested that characteristics observed among inactive females indicated maturity. Given this, we suggest that the accuracy of the immature versus regenerating distinction remains in question; furthermore, considering that our data are based primarily on active females, past studies may have overestimated the size at maturity and thus underestimated female spawning stock size.

In our study, female Red Porgy were often found to contain cohorts of both HOs and $\mathrm{POF}_{0}$. In addition, all females within a sample often contained a cohort of the same reproductive indicator ( HOs or $\mathrm{POF}_{0}$ ). Together, these observations indicate that Red Porgy were spawning frequently, sometimes daily, with $p$ estimated to be as high as 0.88 , yielding one batch every 1.13 d during the spawning season. Published estimates of $p$ exist for nine other snapper-grouper species in the SAB (Collins et al. 1996, 1998; Cuellar et al. 1996; Harris et al. 2002, 2004, 2007; Daniel 2003; Burgos et al. 2007; Danson 2009), with a median $p$ of 0.24 for both $\mathrm{POF}_{0}$ and HO reproductive indicators. Our findings imply that $p$ in Red Porgy may be much higher than estimates reported for other members of the SAB snapper-grouper complex. Previously, $p$ in Red Porgy has been estimated at 0.37 and 0.40 based on POFs and HOs, respectively (Daniel 2003), although uncertainty in POF


FIGURE 7. Predicted annual fecundity $\left(f_{a}\right)$ versus total weight (g) for Red Porgy sampled during (A) 2008-2009 and (B) 2009-2010. The $f_{a}$ was calculated by using a size-independent batch number or by incorporating observed size-dependent relationships based on the proportion of spawning-capable (SC) females, females with day-0 postovulatory follicles $\left(\mathrm{POF}_{0}\right)$, or females with hydrated oocytes (HO). During 2008-2009, only $\mathrm{POF}_{0}$ presence demonstrated a significant relationship with body size.
duration could have caused a negative bias. Other sparids that exhibit daily spawning include the Bluespotted Seabream Pagrus caeruleostictus (Stepkina 1973), Gilthead Bream Sparus auratus (Zohar and Gordin 1979), Red Seabream (Matsuyama et al. 1988a), Squirefish Chrysophrys auratus (Scott et al. 1993), and Yellow Seabream Dentex hypselosomus (Yoda and Yoneda 2009); however, none of these studies explicitly reported estimates of $p$.

We found that $f_{b}$-size relationships were generally weak when data were pooled across all time periods, whereas interannual and intra-annual relationships were often stronger. Higher levels of $f_{b}$ were observed in 2008-2009 than in 20092010, but size-independent $n_{b}$ was higher in 2009-2010, leading to similar levels of size-dependent $f_{a}$ for each year (see Figure 7A, B). These findings confirm that $n_{b}$ and $f_{b}$ can vary separately and possibly independently, meaning that temporal differences in $f_{b}$ do not necessarily imply that differences in $f_{a}$ will be of similar magnitude. In addition, fluctuations in $f_{a}$ may be driven solely by year-to-year changes in $n_{b}$. For example, $f_{b}$ of Red Snapper Lutjanus campechanus did not vary among three years, but $n_{b}$ ranged from 20 to 35 (Collins et al. 1996). Life history theory recognizes eggs per batch and the number of batches as two separate traits, which have been found to be negatively correlated for marine and freshwater fish species within several orders (Wine-


FIGURE 8. Spawning potential ratio (SPR) at different levels of fishing mortality for Red Porgy in (A) 2008-2009 and (B) 2009-2010. The SPR was calculated from annual fecundity estimated using a size-independent batch number or by incorporating observed size-dependent relationships based on the proportion of spawning-capable (SC) females, females with day-0 postovulatory follicles $\left(\mathrm{POF}_{0}\right)$, or females with hydrated oocytes $(\mathrm{HO})$; the SPR calculated using only the biomass of mature females is shown for reference.
miller 1989; Winemiller and Rose 1992). Our findings provide additional evidence that $f_{b}$ and $n_{b}$ probably also vary separately among years within a given species; thus, in many cases it may be inappropriate to assume that $n_{b}$ is temporally constant or varies in a manner similar to $f_{b}$.

During each of our study years, $f_{b}$ was significantly different among months, being lower both early and late in the spawning season. A similar trend was observed by Daniel (2003) but was not statistically significant. Black Sea Bass Centropristis striata have also demonstrated higher $f_{b}$ earlier in the spawning season (Danson 2009), although no appreciable intra-annual variation has been noted for several other SAB snapper-grouper species, including the Vermilion Snapper Rhomboplites aurorubens (Cuellar et al. 1996), Scamp Mycteroperca phenax (Harris et al. 2002), and Blueline Tilefish Caulolatilus microps (Harris et al. 2004). Within the snapper-grouper complex, $f_{b}$ appears to exhibit three general intra-annual relationships that were previously described by Conover (1985) as concave-down, decreasing, or constant. Conover (1985) posited that the concave-down relationship, as observed for Red Porgy in this study, should maximize reproductive success when environmental conditions vary predictably and are optimal in the middle of the season. Sea surface temperature profiles in our study area are reliably concave-up when Red Porgy are reproductively active (November-April; NOAA National Data Buoy Center,
stations 41035 and 41036; www.ndbc.noaa.gov), so perhaps egg survival is highest at colder water temperatures. In a study of the Dwarf Perch Micrometrus minimus, a viviparous temperate marine species, Schultz (1993) found that females born in the middle of the spawning season experienced the highest survival to first reproduction. Female Dwarf Perch that were born in early spring suffered high mortality immediately after birth, whereas females that were born very late suffered high overwinter mortality, probably related to small body size (Schultz 1993). If recently spawned Red Porgy also experience variable seasonal mortality, then increasing egg production in the middle of the season may lead to higher offspring survival rates on average.

## Size Dependence of Batch Number

Significant relationships between the presence of reproductive indicators and Red Porgy body size demonstrated that $n_{b}$ varied with body size and that these relationships differed between years. In 2008-2009, $p$ based on spawning capability or HO presence did not vary with TL, but $p$ based on $\mathrm{POF}_{0}$ decreased with TL. Since HOs precede POFs during oocyte development, $p$ based on HOs should also have decreased with TL; however, we may have failed to detect HOs because they last less than 24 h and are not always detected. In 2009-2010, $p$ increased with TL for all reproductive indicators (SC, $\mathrm{POF}_{0}$ presence, and HO presence), although $p$ was less dependent on TL when using HOs than when using $\mathrm{POF}_{0}$ (see Figure 4B). Our observations for Red Porgy during year two are aligned with a common pattern among batch-spawning species. Studies of the Yellowfin Tuna Thunnus albacares (Schaefer 1998), Baltic Sprat Sprattus sprattus balticus (Kraus and Köster 2004), Southern Blue Whiting Micromesistius australis (Macchi et al. 2005), and Spotted Seatrout Cynoscion nebulosus (LowerreBarbieri et al. 2009) each observed an increase in $p$ with TL, while studies of the Gag Mycteroperca microlepis (Collins et al. 1998) and Ballyhoo Hemiramphus brasiliensis (McBride and Thurman 2003) found that $p$ increased with age. Temporal variation in this pattern, as was observed in our study, may also be common given that (1) Gags only showed a $p-$ TL relationship during one of three study years (Collins et al. 1998) and (2) the pattern was evident for Baltic Sprat during 3 of 4 months (Kraus and Köster 2004). Furthermore, $p$ and $d$ may only sometimes have the same relationship with body size. For the Ballyhoo and Spotted Seatrout, $d$ also increased with age and TL, respectively (McBride and Thurman 2003; Lowerre-Barbieri et al. 2009); however, for Queenfish Seriphus politus, $d$ increased with TL, whereas $p$ did not (DeMartini and Fountain 1981). In a laboratory study of Atlantic Cod (Trippel 1998), $d$ and $n_{b}$ were each higher among second-time spawners than among first-time spawners, although $p$ was not different.

Life history theory suggests that reproductive effort should generally increase with fish age (Roff 1984), and empirical work has shown that across fish species, both potential $f_{a}$ and reproductive investment (i.e., the product of potential $f_{a}$ and egg weight) increase with fish length (Duarte and Alcaraz 1989).

Most of the research on batch spawners has found that $f_{b}$ also increases with fish size and that if $n_{b}$ is constant or increasing, then $f_{a}$ will also increase. A decrease in $n_{b}$ with size, as we observed during 2008-2009 for Red Porgy, is somewhat unexpected since it will tend to cause $f_{a}$ to decrease. One possible explanation for a decrease in $n_{b}$ with increasing size would be if larger females are more likely to skip spawning individual batches while small females skip less often because they are less likely to survive until the next batch (Peterson and Wroblewski 1984). Jonsson et al. (1991) found that the proportion of females that skipped a year between spawning periods increased with body size for Atlantic Salmon Salmo salar. Rideout and Tomkiewicz (2011) suggested that for indeterminate batch spawners, batch skipping is probably a more common tactic than skipping entire years. If local environmental conditions are unfavorable for offspring survival on a particular date, a female with a high probability of survival should benefit from postponing a batch until conditions improve. Because females must provision a limited supply of yolk among their eggs (Lloyd and Gosselin 2007) and because initial offspring size can affect growth rate and survival (Marteinsdottir and Steinarsson 1998), skipping a current batch may also allow for the production of a future batch of larger, more successful offspring. By the end of the spawning season, fewer batches may have been spawned, but higher success per offspring and energy retained for future spawning seasons may increase lifetime reproductive success (Stearns 1992).

Among protogynous species, large females may have another incentive to divert surplus energy from egg production to growth. According to Warner's model (Warner 1984; Munday et al. 2006), the reproductive value of a protogynous fish increases dramatically when it reaches a large body size and can transition to a dominant male; this idea is also supported by empirical research (Warner and Hoffman 1980). Warner's model suggests that as protogynous fish grow, the ratio of female to male reproductive value increases up to a certain size and then decreases, and sex change is predicted to occur when the ratio declines to $1: 1$. Thus, as females approach the size at which sex transition is favored, the value of investment in eggs should decrease relative to the value of investment in processes (e.g., growth) that would be expected to improve male reproductive success. Sex-specific reproductive values are rarely fixed and tend to be highly variable among individuals based on local demographics (e.g., size distribution, sex ratio, and density) that impact social behavior (Munday et al. 2006). Thus, the reproductive benefit to females of either maximizing egg production or diverting energy to improve future success as a male is likely to be largely dependent on local demographics.

Although the causal mechanisms are not clear, our findings for Red Porgy support the idea that $n_{b}$-size relationships vary temporally. Considering that other studies-albeit few in number-have noted similar variation (Collins et al. 1998; Kraus and Köster 2004), temporal variation in the effect of body size on $n_{b}$ may be common among batch-spawning fishes. The potential for the timing of sex transition to contribute to
variability in the batch spawning pattern remains speculative but is generally supported by existing theory. Given the potentially large impact of $n_{b}$ on estimates of $f_{a}$, the relationship between $n_{b}$ and body size deserves further empirical and theoretical study among both gonochoristic and hermaphroditic species.

## Effects of Variable Reproductive Indicators on Estimates of Egg Production and Management Reference Points

Incorporating size-based relationships for reproductive indicators into calculations of $f_{a}$ and SPR produced notable differences compared to models that assumed $n_{b}$ to be independent of size. The decrease in $\mathrm{POF}_{0}$ presence at larger body sizes during 2008-2009 resulted in lower $f_{a}$ in large females, whereas increases in the presence of all reproductive indicators at larger sizes during 2009-2010 resulted in higher $f_{a}$ for large females (see Figure 7A, B). In turn, size-selective fishing mortality was predicted to have less impact on SPR in 2008-2009 than in 2009-2010. When $n_{b}$ was assumed to be independent of size, the estimated SPR was generally similar to calculations that were based solely on mature female biomass (see Figure 8A, B).

It is clear that $n_{b}$ is size dependent, but the relationships can differ depending on the choice of reproductive indicator. We conclude that the presence of $\mathrm{POF}_{0}$ is the most reliable reproductive indicator and likely produces the most accurate estimates of egg production and SPR. Hydrated oocytes are more readily identifiable when using histological techniques, but their short duration means that they may not be encountered during all times of day. Egg production estimates based on spawning capability are somewhat indirect because they assume that the duration of the SC season is proportional to $n_{b}$. However, this is an implicit assumption of standard calculations in which $n_{b}$ is assumed to be independent of body size. In any regard, spawning capability-size relationships can be useful proxies for $n_{b}$-size relationships because (1) spawning capability is easier to assess than $\mathrm{POF}_{0}$ presence and can be determined macroscopically and (2) vitellogenic oocytes have much longer durations that are not temperature dependent like those of HOs or $\mathrm{POF}_{0}$ (Fitzhugh and Hettler 1995; Kurita et al. 2011). Thus, for long-term surveys investigating the size dependence of $n_{b}$, an assessment of spawning capability may be more feasible than assessing $\mathrm{POF}_{0}$ or HO presence.

Our results indicate that Red Porgy SPR could be lowered sufficiently (relative to current projections) to impact management if $n_{b}$ relates positively to body size. If size dependence of $p$ is mostly positive, as is supported by evidence for other species, and if such size dependence is widespread among indeterminate reef fishes, this could have important implications for management of the SAB snapper-grouper complex. The use of nontraditional management strategies (e.g., marine protected areas and maximum size limits) within the offshore fishery has already been discussed or introduced in attempts to protect males from harvest. This is because model simulations have predicted that decreasing fertilization rates due to a loss of male biomass will cause female SSB to be a poor predictor of recruitment in protogynous hermaphrodites (Brooks et al. 2008). Management
strategies that are designed to protect larger individuals should also help to prevent the removal of larger females that may be generating a greater number of egg batches each year. Increased biomass of males may also contribute to a reversal in the declining trend in size at sex transition (observed by Harris and McGovern 1997), which could translate to larger female sizes in the stock and thus a potential for greater contrast in the $n_{b}-\mathrm{TL}$ relationship.

Spawning stock biomass is often considered a poor proxy for egg production in determinate-spawning species (Marshall et al. 1998, 2003; Cardinale and Arrhenius 2000), and we have shown that this is also likely true for an indeterminate species. Our work represents one of the more detailed studies of fecundity in an indeterminate protogynous reef fish in terms of the number of reproductively active females that were sampled per year. The level of sampling enabled us to identify size-dependent relationships for both $f_{b}$ and $n_{b}$ with sufficient temporal resolution to detect variability between years. Although sampling demands can make it difficult to conduct detailed studies of reproductive dynamics in larger, less-abundant species, the patterns observed for Red Porgy outline a range of potential patterns that may be widespread among similar species. Due to the relatively small body size and accessibility of the Red Porgy, it may prove to be a valuable model species for studying the complex patterns of egg production among indeterminate, batch-spawning, protogynous hermaphrodites.

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## REFERENCES

Alekseev, F. E. 1982. Hermaphroditism in sparid fishes (Perciformes, Sparidae): 1. protogyny in porgies, Pagrus pagrus, P. orphus, P. ehrenbergi and P. auriga from West Africa. Journal of Ichthyology 22(5):85-94.
Armstrong, M. J., and P. R. Witthames. 2012. Developments in understanding of fecundity of fish stocks in relation to egg production methods for estimating spawning stock biomass. Fisheries Research 117/118:35-47.
Ball, A. O., G. R. Sedberry, J. H. Wessel III, and R. W. Chapman. 2003. Largescale genetic differentiation of Pagrus pagrus in the Atlantic. Journal of Fish Biology 62:1232-1237.
Brooks, E. N., K. W. Shertzer, T. Gedamke, and D. S. Vaughan. 2008. Stock assessment of protogynous fish: evaluating measures of spawning biomass used to estimate biological reference points. U.S. National Marine Fisheries Service Fishery Bulletin 106:12-23.

Brown-Peterson, N. J., D. M. Wyanski, F. Saborido-Rey, B. J. Macewicz, and S. K. Lowerre-Barbieri. 2011. A standardized terminology for describing reproductive development in fishes. Marine and Coastal Fisheries: Dynamics, Management, and Ecosystem Science 3:52-70.
Burgos, J. M., G. R. Sedberry, D. M. Wyanski, and P. J. Harris. 2007. Life history of Red Grouper (Epinephelus morio) off the coasts of North Carolina and South Carolina. Bulletin of Marine Science 80:45-65.
Cardinale, M., and F. Arrhenius. 2000. The relationship between stock and recruitment: are the assumptions valid? Marine Ecology Progress Series 196:305-309.
Collins, L. A., A. G. Johnson, and C. P. Keim. 1996. Spawning and annual fecundity of Red Snapper (Lutjanus campechanus) from the northeastern Gulf of Mexico. Pages 174-188 in F. Arreguín-Sánchez, J. L. Munro, M. C. Balgos, and D. Pauly, editors. Biology, fisheries and culture of tropical groupers and snappers. International Center for Living Aquatic Resources Management, Manila.
Collins, L. A., A. G. Johnson, C. C. Koenig, and M. S. Baker Jr. 1998. Reproductive patterns, sex ratio, and fecundity in Gag, Mycteroperca microlepis (Serranidae), a protogynous grouper from the northeastern Gulf of Mexico. U.S. National Marine Fisheries Service Fishery Bulletin 96:415-427.

Conover, D. O. 1985. Field and laboratory assessment of patterns in fecundity of a multiple spawning fish: the Atlantic Silverside Menidia menidia. U.S. National Marine Fisheries Service Fishery Bulletin 83:331-341.
Cuellar, N., G. R. Sedberry, and D. M. Wyanski. 1996. Reproductive seasonality, maturation, fecundity, and spawning frequency of the Vermilion Snapper, Rhomboplites aurorubens, off the southeastern United States. U.S. National Marine Fisheries Service Fishery Bulletin 94:635-653.
Dalgaard, P. 2008. Introductory statistics with R. Springer, New York.
Daniel, E. A. 2003. Sexual maturity, spawning dynamics, and fecundity of Red Porgy, Pagrus pagrus, off the southeastern United States. Master's thesis. University of Charleston, Charleston, South Carolina.
Danson, B. L. 2009. Estimating reef fish reproductive productivity on artificial and natural reefs off the Atlantic coast of the southeastern United States. Master's thesis. College of Charleston, Charleston, South Carolina.
DeMartini, E. E., and R. K. Fountain. 1981. Ovarian cycling frequency and batch fecundity in the Queenfish, Seriphus politus: attributes representative of serial spawning fishes. U.S. National Marine Fisheries Service Fishery Bulletin 79:547-560.
Duarte, C. M., and M. Alcaraz. 1989. To produce many small or few large eggs: a size-independent reproductive tactic of fish. Oecologia 80:401-404.
Fitzhugh, G. R., and W. F. Hettler. 1995. Temperature influence on postovulatory follicle degeneration in Atlantic Menhaden, Brevoortia tyrannus. U.S. National Marine Fisheries Service Fishery Bulletin 93:568-572.
Fitzhugh, G. R., K. W. Shertzer, G. T. Kellison, and D. M. Wyanski. 2012. Review of size- and age-dependence in batch spawning: implications for stock assessment of fish species exhibiting indeterminate fecundity. U.S. National Marine Fisheries Service Fishery Bulletin 110:413-425.
Gabriel, W. L., M. P. Sissenwine, and W. J. Overholtz. 1989. Analysis of spawning stock biomass per recruit: an example for Georges Bank Haddock. North American Journal of Fisheries Management 9:383-391.
Ganias, K., S. Somarakis, A. Machias, and A. Theodorou. 2004. Pattern of oocyte development and batch fecundity in the Mediterranean sardine. Fisheries Research 67:13-23.
Goodyear, C. P. 1993. Spawning stock biomass per recruit in fisheries management: foundation and current use. Pages 67-81 in S. J. Smith, J. J. Hunt, and D. Rivard, editors. Risk evaluation and biological reference points for fisheries management. Canadian Special Publication of Fisheries and Aquatic Sciences 120.
Harris, P. J., and J. C. McGovern. 1997. Changes in the life history of Red Porgy, Pagrus pagrus, from the southeastern United States, 1972-1994. U.S. National Marine Fisheries Service Fishery Bulletin 95: 732-747.
Harris, P. J., D. M. Wyanski, and P. T. Powers Mikell. 2004. Age, growth, and reproductive biology of Blueline Tilefish along the southeastern coast of the

United States, 1982-1999. Transactions of the American Fisheries Society 133:1190-1204.
Harris, P. J., D. M. Wyanski, D. B. White, P. P. Mikell, and P. B. Eyo. 2007. Age, growth, and reproduction of Greater Amberjack off the southeastern U.S. Atlantic coast. Transactions of the American Fisheries Society 136:15341545.

Harris, P. J., D. M. Wyanski, D. B. White, and J. L. Moore. 2002. Age, growth, and reproduction of Scamp, Mycteroperca phenax, in the southwestern North Atlantic, 1979-1997. Bulletin of Marine Science 70:113-132.
Hilborn, R., and C. J. Walters. 1992. Quantitative fisheries stock assessment: choice, dynamics and uncertainty. Chapman and Hall, New York.
Hosmer, D. W., and S. Lemeshow. 1989. Applied logistic regression. Wiley, New York.
Hunter, J. R., and S. R. Goldberg. 1980. Spawning incidence and batch fecundity in Northern Anchovy, Engraulis mordax. U.S. National Marine Fisheries Service Fishery Bulletin 77:641-652.
Hunter, J. R., and R. Leong. 1981. The spawning energetics of female Northern Anchovy, Engraulis mordax. U.S. National Marine Fisheries Service Fishery Bulletin 79:215-230.
Hunter, J. R., N. C. H. Lo, and R. J. H. Leong. 1985. Batch fecundity in multiple spawning fishes. NOAA Technical Report NMFS 36:67-77.
Hunter, J. R., and B. J. Macewicz. 1985. Measurement of spawning frequency in multiple spawning fishes. NOAA Technical Report NMFS 36:79-94.
Hunter, J. R., and B. J. Macewicz. 2003. Improving the accuracy and precision of reproductive information used in fisheries. Pages 57-68 in O. S. Kjesbu, J. R. Hunter, and P. R. Witthames, editors. Report of the working group on modern approaches to assess maturity and fecundity of warm- and cold-water fish and squids. Institute of Marine Science, Bergen, Norway.
Jennings, S., M. J. Kaiser, and J. D. Reynolds. 2001. Marine fisheries ecology. Blackwell Scientific Publications, Oxford, UK.
Jonsson, N., L. P. Hansen, and B. Jonsson. 1991. Variation in age, size and repeat spawning of adult Atlantic Salmon in relation to river discharge. Journal of Animal Ecology 60:937-947.
Kjesbu, O. S. 2009. Applied fish reproductive biology: contribution of individual reproductive potential to recruitment and fisheries management. Pages 293332 in T. Jakobsen, M. Fogarty, B. A. Megrey, and E. Moksness, editors. Fish reproductive biology: implications for assessment and management. Blackwell Scientific Publications, Chichester, UK.
Kjesbu, O. S., P. R. Witthames, P. Solemdal, and M. Greer Walker. 1998. Temporal variations in the fecundity of Arcto-Norwegian Cod (Gadus morhua) in response to natural changes in food and temperature. Journal of Sea Research 40:303-321.
Klibansky, N. 2006. Differences in fecundity and egg size of cod on Georges Bank and in the Gulf of Maine. Master's thesis. University of Massachusetts, Amherst.
Klibansky, N., and F. Juanes. 2008. Procedures for efficiently producing highquality fecundity data on a small budget. Fisheries Research 89:84-89.
Kraus, G., and F. W. Köster. 2004. Estimating Baltic sprat (Sprattus sprattus balticus S.) population sizes from egg production. Fisheries Research 69:313329.

Kraus, G., A. Müller, K. Trella, and F. W. Köster. 2000. Fecundity of Baltic cod: temporal and spatial variation. Journal of Fish Biology 56:1327-1341.
Kurita, Y., Y. Fujinami, and M. Amano. 2011. The effect of temperature on the duration of spawning markers-migratory-nucleus and hydrated oocytes and postovulatory follicles-in the multiple-batch spawner Japanese Flounder (Paralichthys olivaceus). U.S. National Marine Fisheries Service Fishery Bulletin 109:79-89.
Lambert, Y., N. A. Yaragina, G. Kraus, G. Marteinsdottir, and P. J. Wright. 2003. Using environmental and biological indices as proxies for egg and larval production of marine fish. Journal of Northwest Atlantic Fishery Science 33:115-159.
Lloyd, M. J., and L. A. Gosselin. 2007. Role of maternal provisioning in controlling interpopulation variation in hatching size in the marine snail Nucella ostrina. Biological Bulletin 213:316-324.

Lowerre-Barbieri, S. K., N. Henderson, J. Llopiz, S. Walters, J. Bickford, and R. Muller. 2009. Defining a spawning population (Spotted Seatrout Cynoscion nebulosus) over temporal, spatial, and demographic scales. Marine Ecology Progress Series 394:231-245.
Macchi, G. J., M. Pájaro, O. C. Wöhler, M. J. Acevedo, R. L. Centurión, and D. G. Urteaga. 2005. Batch fecundity and spawning frequency of Southern Blue Whiting (Micromesistius australis) in the southwest Atlantic Ocean. New Zealand Journal of Marine and Freshwater Research 39:993-1000.
Manooch, C. S., III. 1976. Reproductive cycle, fecundity, and sex ratios of the Red Porgy, Pagrus pagrus (Pisces: Sparidae) in North Carolina. U.S. National Marine Fisheries Service Fishery Bulletin 74:775-781.
Marshall, C. T., O. S. Kjesbu, N. A. Yaragina, P. Solemdal, and Ø. Ulltang. 1998. Is spawner biomass a sensitive measure of the reproductive and recruitment potential of northeast Arctic cod? Canadian Journal of Fisheries and Aquatic Sciences 55:1766-1783.
Marshall, C. T., C. L. Needle, A. Thorsen, O. S. Kjesbu, and N. A. Yaragina. 2006. Systematic bias in estimates of reproductive potential of an Atlantic Cod (Gadus morhua) stock: implications for stock-recruit theory and management. Canadian Journal of Fisheries and Aquatic Sciences 63:980-994.
Marshall, C. T., L. O'Brien, J. Tomkiewicz, F. W. Köster, G. Kraus, G. Marteinsdottir, M. J. Morgan, F. Saborido-Rey, J. L. Blanchard, D. H. Secor, P. J. Wright, N. V. Mukhima, and H. Björnsson. 2003. Developing alternative indices of reproductive potential for use in fisheries management: case studies for stocks spanning an information gradient. Journal of Northwest Atlantic Fishery Science 33:161-190.
Marteinsdottir, G., and A. Steinarsson. 1998. Maternal influence on the size and viability of Iceland cod Gadus morhua eggs and larvae. Journal of Fish Biology 52:1241-1258.
Matsuyama, M., S. Adachi, Y. Nagahama, and S. Matsuura. 1988a. Diurnal rhythm of oocyte development and plasma steroid hormone levels in the female Red Sea Bream, Pagrus major, during the spawning season. Aquaculture 73:357-372.
Matsuyama, M., R. Torres Lara, and S. Matsuura. 1988b. Juvenile bisexuality in the Red Sea Bream, Pagrus major. Environmental Biology of Fishes 21: 27-36.
McBride, R. S., and P. E. Thurman. 2003. Reproductive biology of Hemiramphus brasiliensis and H. balao (Hemiramphidae): maturation, spawning frequency, and fecundity. Biological Bulletin 204:57-67.
McIntyre, T. M., and J. A. Hutchings. 2003. Small-scale temporal and spatial variation in Atlantic Cod (Gadus morhua) life history. Canadian Journal of Fisheries and Aquatic Sciences 60:1111-1121.
Munday, P. L., P. M. Buston, and R. R. Warner. 2006. Diversity and flexibility of sex-change strategies in animals. Trends in Ecology and Evolution 21:89-95.
Murua, H., G. Kraus, F. Saborido-Rey, P. R. Witthames, A. Thorsen, and S. Junquera. 2003. Procedures to estimate fecundity of marine fish species in relation to their reproductive strategy. Journal of Northwest Atlantic Fishery Science 33:33-54.
Peterson, I., and J. S. Wroblewski. 1984. Mortality rate of fishes in the pelagic ecosystem. Canadian Journal of Fisheries and Aquatic Sciences 41: 1117-1120.
Rideout, R. M., and J. Tomkiewicz. 2011. Skipped spawning in fishes: more common than you might think. Marine and Coastal Fisheries: Dynamics, Management, and Ecosystem Science 3:176-189.

Roff, D. A. 1984. The evolution of life history parameters in teleosts. Canadian Journal of Fisheries and Aquatic Sciences 41:989-1000.
Schaefer, K. M. 1998. Reproductive biology of Yellowfin Tuna (Thunnus albacares) in the eastern Pacific Ocean. Inter-American Tropical Tuna Commission Bulletin 21:205-272.
Schultz, E. T. 1993. The effect of birth date on fitness of female Dwarf Perch, Micrometrus minimus (Perciformes: Embiotocidae). Evolution 47:520539.

Scott, S. G., J. R. Zeldis, and N. W. Pankhurst. 1993. Evidence of daily spawning in natural populations of the New Zealand Snapper Pagrus auratus (Sparidae). Environmental Biology of Fishes 36:149-156.
SEDAR (SouthEast Data, Assessment, and Review). 2006. Stock assessment of Red Porgy off the southeastern United States: SEDAR update assessment. SEDAR, Southeast Fisheries Science Center, Beaufort, North Carolina.
Stearns, S. C. 1992. The evolution of life histories. Oxford University Press, New York.
Stepkina, M. V. 1973. Some biological characteristics of Pagrus ehrenbergii Val. Journal of Ichthyology 13:641-649.
Tomkiewicz, J., M. J. Morgan, J. Burnett, and F. Saborido-Rey. 2003. Available information for estimating reproductive potential of northwest Atlantic groundfish stocks. Journal of Northwest Atlantic Fishery Science 33: 1-21.
Trippel, E. A. 1998. Egg size and viability and seasonal offspring production of young Atlantic Cod. Transactions of the American Fisheries Society 127: 339-359.
Warner, R. R. 1984. Mating behavior and hermaphroditism in coral reef fishes. American Scientist 72:128-136.
Warner, R. R., and S. G. Hoffman. 1980. Population density and the economics of territorial defense in a coral reef fish. Ecology 61:772-780.
Winemiller, K. O. 1989. Patterns of variation in life history among South American fishes in seasonal environments. Oecologia 81:225-241.
Winemiller, K. O., and K. A. Rose. 1992. Patterns of life-history diversification in North American fishes: implications for population regulation. Canadian Journal of Fisheries and Aquatic Sciences 49:2196-2218.
Witthames, P. R., A. Thorsen, H. Murua, F. Saborido-Rey, L. N. Greenwood, R. Dominguez, M. Korta, and O. S. Kjesbu. 2009. Advances in methods for determining fecundity: application of the new methods to some marine fishes. U.S. National Marine Fisheries Service Fishery Bulletin 107: 148-164.
Wootton, R. J., and D. A. Fletcher. 2009. Effect of spawning number and ration on reproductive performance of the batch-spawning Three-Spined Stickleback Gasterosteus aculeatus. Journal of Fish Biology 75:618-629.
Yoda, M., and M. Yoneda. 2009. Assessment of reproductive potential in multiple-spawning fish with indeterminate fecundity: a case study of Yellow Sea Bream Dentex hypselosomus in the East China Sea. Journal of Fish Biology 74:2338-2354.
Yoneda, M., and P. J. Wright. 2004. Temporal and spatial variation in reproductive investment of Atlantic Cod Gadus morhua in the northern North Sea and Scottish west coast. Marine Ecology Progress Series 276:237248.

Zohar, Y., and H. Gordin. 1979. Spawning kinetics in the Gilthead Sea-Bream, Sparus aurata L. after low doses of human chronic gonadotropin. Journal of Fish Biology 15:665-670.


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