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ARTICLE

Genetic Variability and Population Structure of Gulf Menhaden Compared with Yellowfin Menhaden

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Abstract

Gulf Menhaden *Brevoortia patronus* is one of the most intensively fished organisms in the Gulf of Mexico. Fishery managers and industry operators have historically worked towards a sustainable fishery and have cooperated on stock assessments to estimate feasible reference points for management. These stock assessments are necessarily rooted in a strong definition of the fishery stock, including the estimation of the number of populations that are exploited. Here, genetic population structure and variability were examined in *B. patronus*, using genetic markers specifically designed for the species. We observed genetic variability that indicates a relatively high effective population size for a marine finfish ($N_e \geq 1,200$), and two analytical approaches implied a single genetic population of *B. patronus*. We compared the latter finding with the population structure in the closely related Yellowfin Menhaden *B. smithi*, for which two distinct populations from Florida were identified using the same genetic loci ($F_{st} = 0.015$, $P = 0.027$). These contrasting patterns of population structure between sympatric congeneric species are likely driven by differences in distribution and census size and may relate to factors that originally drove speciation in North American *Brevoortia* species. The finding of a single Gulf-wide population of *B. patronus* suggests that there is an extensive migration throughout the species range and supports the notion of a single genetic stock.

The purse seine fishery for menhaden *Brevoortia* spp. (Clupeidae) is one of the oldest fisheries in United States (Smith 1991) and is supported primarily by two species: the Atlantic Menhaden *B. tyrannus* and the Gulf Menhaden *B. patronus*. Presently, *B. patronus* is the most intensively harvested finfish species in the Gulf of Mexico (“Gulf”) and supports one of the largest commercial fisheries in the United States (Vaughan et al. 2007). This species forms numerically immense, shallow, near-shore schools, and this tendency makes it easy to harvest in high numbers using purse boats deploying seines up to 365 m in length (Smith 1991). Despite this intensive harvest, landings of *B.*

patronus appear to be generally below the maximum sustainable yield, and the fishery is thought to be stable (Vaughan et al. 2000).

The maintenance of a long-term sustainable harvest of *B. patronus* is important given the prominence of this fishery in the Gulf. A critical element for achieving this goal is to develop stock assessments that are anchored by a strong definition of the unit stock (Begg and Waldman 1999). The definition of a fishery “stock” has evolved through the years from one that was primarily driven by distinct features of local fishing economies to one that is more rooted in the biology of the organism and includes assessments of reproduction, demo-

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graphics, and genetics (Carvalho and Hauser 1994; Begg and Waldman 1999; Waples et al. 2008). Modern stock concepts take into account practical fishery considerations as well as political concerns, but are also heavily rooted by the idea of a genetic stock (Carvalho and Hauser 1994). For *B. patronus*, the stock definition used in stock assessments has historically been defined by multiple data considerations, including an estimation of the number of genetically distinctive populations occurring in the fished area (SEDAR 2013). However, this component of the stock definition has been limited by the magnitude of genetic sampling in previous studies (Anderson 2006). An increase in the number of genetic markers used for eliciting discreet genetic stocks would greatly improve the stock definition by improving statistical power (SEDAR 2013; Anderson and Karel 2014).

The possibility of multiple genetically distinct stocks in *B. patronus* may be confounded by a potential for extensive gene flow across the range of the species based on aspects of reproductive biology at multiple life stages. The species undergoes an offshore migration beginning in October, and the fish are caught offshore at various depths in the northern Gulf (Roithmayr and Waller 1963). Based on tag return data, in addition to this offshore movement there also may be a tendency for adults of the species to move closer to the center of the species' range (Ahrenholz 1981; although see Pristas et al. 1976). The seasonal migratory period of adults also coincides with the spawning season (Raynie and Shaw 1994), and spawning in *B. patronus* occurs from near shore to at least 96 km offshore (Christmas and Waller 1975). The offshore pelagic egg–larval stage may last up to 10 weeks (Shaw et al. 1985; Deegan and Thompson 1987), whereby migration occurs passively and may rely on nearshore currents that potentially move individuals laterally along shorelines before they settle into estuaries. Thus, two stages of migration (adult seasonal active, larval passive) result in a potential for long-distance gene flow and homogenization of allele and genotypic frequencies across a broad scale. This would imply the potential for a single unit stock, although this hypothesis has not been rigorously examined with a large genetic data set.

An additional consideration with regard to management of *B. patronus* is whether intensive harvest or other processes have impacted the genetic variability or the effective population size (N_e) in Gulf stocks. There is a high estimated natural mortality in *B. patronus* (Smith 1991). In marine systems, variances in productivity and survivorship among demes and high estimates of natural mortality have historically been linked to wide disparities between population census size and genetic effective size (Turner et al. 2002). This phenomenon can be exacerbated in heavily harvested species; long-term harvest has been related to low N_e in other exploited species (Diaz et al. 2000; Hauser et al. 2002). A recent meta-analysis of marine populations indicated that genetic diversity is generally lower in overharvested populations than in stable ones (Pinsky and Palumbi 2014).

The commercial harvest of *B. patronus* is generally heaviest in an area between Cameron, Louisiana, and Moss Point,

Mississippi, which is roughly the center of the species' distribution. Estimates from landings data suggest that $4\text{--}11.5 \times 10^9$ fish are harvested each year (Smith 1991; Vaughan et al. 2007), but the stock is thought to be stable and not overfished (Vaughan et al. 2000). Thus, one question that might be answered with genetic data is whether there are observable negative impacts on genetic diversity in the center of the range of *B. patronus* due to fishing. Bowen and Avise (1990) previously estimated the effective size of the female population in menhaden (N_{fe}) to be $\sim 250,000$ individuals, suggesting very high genetic diversity. However, the method used in that study was based on nucleotide diversity at a single mitochondrial DNA (mtDNA) locus and relied upon rough estimation under a coalescent model. The estimate of Bowen and Avise (1990) thus may simply reflect long-term (evolutionary) effective size among mtDNA lineages only. In comparison, a short-term estimate of N_e would more accurately reflect the effective size of the contemporary population and would be more useful for estimating existing adaptive potential (Hare et al. 2011).

Here, the question of gene flow, population structure, and genetic variation of *B. patronus* was addressed by applying a suite of novel genetic markers (Anderson and Karel 2014) to Gulf-wide samples of *Brevoortia* species. The study design strategy involved sampling primarily juveniles from inshore areas, which also incidentally resulted in relatively large samples of Yellowfin Menhaden *B. smithi* from Florida. The population structure of *B. smithi* in Florida was compared with that of *B. patronus* Gulf-wide. This genetic examination of the unit stock in the *B. patronus* fishery can be used to improve the stock definition for future Gulf Menhaden stock assessments.

METHODS

Sample collection and laboratory methods.—Menhaden specimens were collected throughout the Gulf during two targeted sampling periods (Table 1; Figure 1). The first (“early”) period was during the years 2002–2003. These samples were collected during the course of a previous study (Anderson 2006), but DNA extracts from these specimens were reanalyzed in this study using additional genetic markers. Sampling methods for early sampling can be found in Anderson (2007).

The second (“late”) group of samples was collected in 2012–2014. These included five sampling locations in Texas, two locations in Louisiana, and three locations in Florida. When possible, sampling locations for late samples were geographically proximate to those for the early samples. Menhaden were collected using various gears, mainly bag seines and inshore trawls. In most cases, samples consisted of whole juvenile specimens (<100 mm TL). In a few cases, adults were sampled by excising a small portion of fin tissue from the caudal fin. Whole genomic DNA was extracted from caudal fin tissues of all specimens using a Gentra Puregene tissue isolation kit (Qiagen, Valencia, California) following the manufacturer's instructions.

TABLE 1. Sample sizes of *Brevoortia* for each sampling locality and each of two temporal (early versus late) sampling periods sorted from west to east longitude. Samples were broken down by species and hybrids using genetic species identification.

Sample	Year	<i>n</i> (total)	<i>B. patronus</i>	<i>B. gunteri</i>	<i>B. smithi</i>	Hybrids
Upper Laguna Madre, Texas	2002	35	34	1	0	0
Upper Laguna Madre, Texas	2013	43	42	1	0	0
Aransas Bay, Texas	2002	50	50	0	0	0
Aransas Bay, Texas	2012	39	39	0	0	0
San Antonio Bay, Texas	2002	53	53	0	0	0
San Antonio Bay, Texas	2013	60	60	0	0	0
Galveston Bay, Texas	2002	51	51	0	0	0
Galveston Bay, Texas	2013	46	46	0	0	0
Sabine Lake, Texas	2002	49	49	0	0	0
Sabine Lake, Texas	2012	50	50	0	0	0
Terrebonne Bay, Louisiana	2014	46	46	0	0	0
Empire, Louisiana	2002	46	46	0	0	0
Breton Basin, Louisiana	2014	50	50	0	0	0
Moss Point, Mississippi	2002	48	48	0	0	0
Apalachicola, Florida	2003	35	35	0	0	0
Apalachicola, Florida	2014	47	42	0	4	1
Cedar Key, Florida	2014	49	9	0	37	3
Tampa Bay, Florida	2014	50	0	0	50	0
Charlotte Harbor, Florida	2003	36	0	0	35	1
Total (all samples)		883	750	2	126	5

Fourteen microsatellite loci described in Anderson and Karel (2014) were used to develop multilocus genotypes for all specimens. Nine of these were novel loci developed specifically for the genus *Brevoortia*. The remaining five were developed for use in a closely related clupeid, American Shad *Alosa sapidissima*, and were included here for comparison with previous genetic examinations that used these loci in *Brevoortia* species (Anderson 2006, 2007). Each primer set was labeled with a WellRed fluorescent dye (primer manufacture and labeling by Sigma-Aldrich) and used to amplify approximately 100 ng of genomic DNA for each specimen through the use of PCR. Amplified products were multiplexed in four independent panels with a 400-bp size standard and separated with a CEQ 8000 sequencer (Beckman Coulter). Fragment analysis was performed using GeneMarker version 2.4 (SoftGenetics LLC, State College, Pennsylvania), and all allelic bins were established based on estimated allele size in base pairs.

Genetic species identification.—Samples consisted primarily of small juveniles, and it was anticipated that some individuals sampled in the western Gulf may actually have been Finescale Menhaden *B. gunteri* and that some individuals sampled in the eastern Gulf may actually have been *B. smithi*. The range of *B. patronus* overlaps the ranges of each of these species. Morphological identification in juvenile specimens is extremely difficult, so multilocus microsatellite genotypes were used to determine species in all specimens. This was accomplished by using the software Structure version 2.3.4 (Pritchard et al. 2000).

The analysis consisted of 10,000 burn-in iterations followed by 90,000 iterations for clustering. Populations were constrained to $K = 2$ to allow for the identification of two general categories: *B. patronus* and combined *B. gunteri*–*B. smithi* (the latter species likely share common lineage and generally cluster together in phylogenetic analyses: Anderson 2007). Genotypes were sorted into these categories, and if individuals shared a cluster lineage >0.20 with both inferred clusters, they were classified as hybrids (as in Anderson and Karel 2007). A second identical Structure analysis ($K = 2$, 10,000 burn-in iterations and 90,000 clustering iterations) was used to separate putative *B. gunteri* and *B. smithi*. Species identification was aided in this case by the knowledge that *B. gunteri* generally occur in the western Gulf west of the Mississippi River and *B. smithi* occur in the eastern Gulf (Dahlberg 1970). When the results of this second Structure analysis did not correspond to expectations based on sampling geography, an additional Structure run was completed with $K = 3$ to provide further resolution (see Results).

Microsatellite diversity and genetic structure.—The observed (H_o) and expected (H_e) heterozygosity of each marker locus was calculated for each species observed in the sample, and the within-group coancestry coefficient (F_{is}) was calculated over all samples (species specific) using the software Fstat version 2.9.3 (Goudet 1995). It was expected that H_e would generally exceed H_o and that most loci would demonstrate elevated F_{is} relative to the Hardy–Weinberg equilibrium expectations, as a result of combining two temporal sampling periods, with

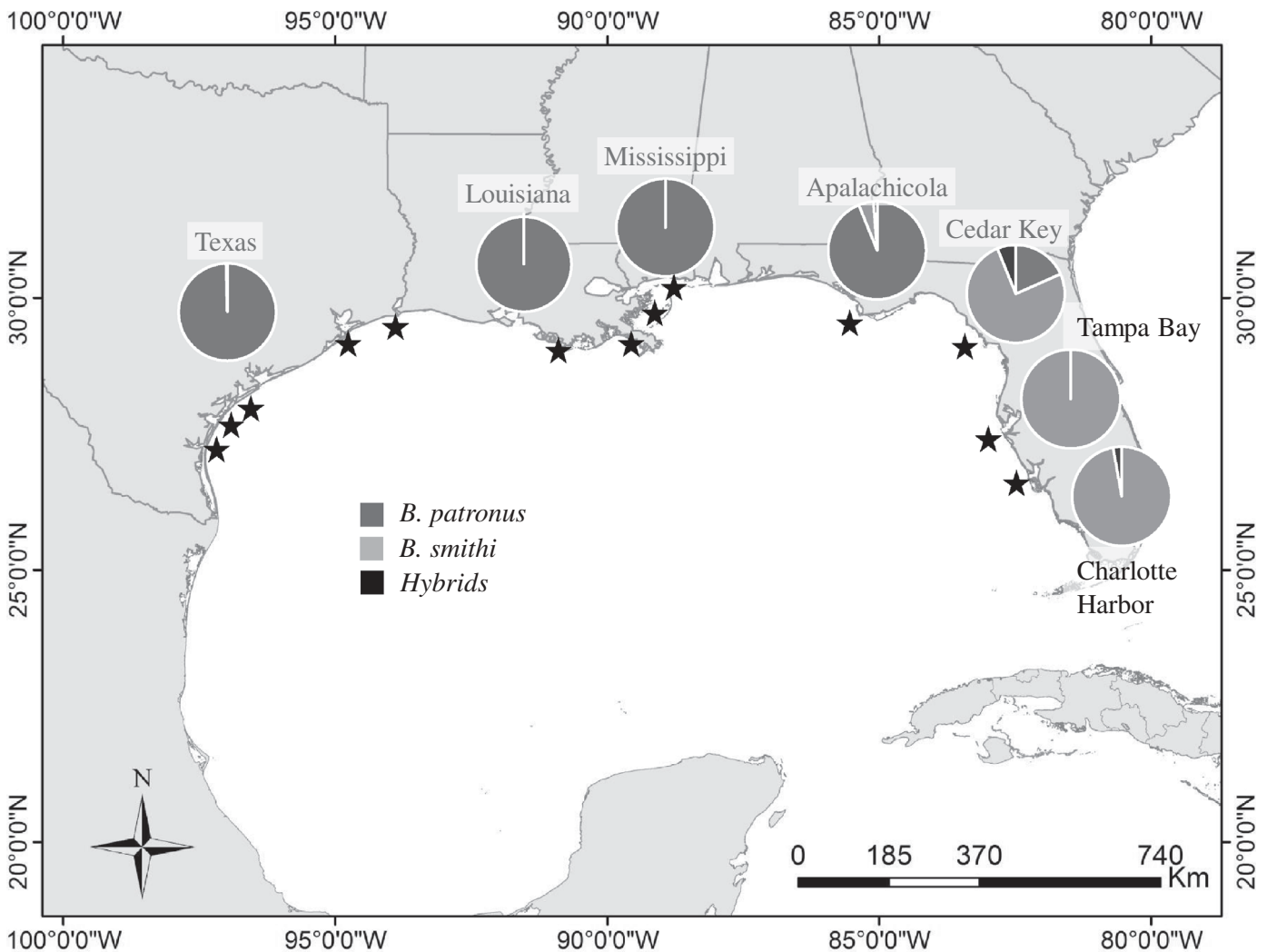


FIGURE 1. The sampling distribution for this study of *Brevoortia* spp. in the Gulf of Mexico. Approximate sample locations are indicated by stars. In some cases, multiple samples were collected over two time periods from the same location (these are listed in Table 1). Pie charts indicate the proportion of *B. patronus*, *B. smithi*, and hybrids observed in samples from each state. Note that samples taken in Florida have individual pie charts due to differences in the distribution of species among samples in the eastern Gulf of Mexico. Two individual *B. gunteri* were observed from Texas samples out of 474 combined specimens; these were the only specimens outside of Florida that were not *B. patronus*.

samples spread across the Gulf. The confidence interval for each locus-specific estimate of F_{is} (with all samples combined) was estimated in Fstat and used to determine which loci had elevated F_{is} relative to the overall mean F_{is} . Marker loci that deviated significantly from the group-wise estimate of F_{is} were included in genetic structure analysis but were excluded in comparisons of genetic diversity and effective population size (N_e , see below). It was expected that the genetic structure analysis would be fairly robust to violations of Hardy–Weinberg expectations but that estimates of genetic diversity and N_e would not.

Genetic structure was assessed in *B. patronus* and *B. smithi* independently using Structure version 2.3.4. All samples from across the sampling geography were combined, and three levels of potential group structure were assessed ($K = 1, 2,$

and 3) for each species. This analytical framework assumes that some level of population subdivision may be detected across the species range, but no assumptions were made as to the geographical arrangement of those populations (that is, geographic data were not used to influence a priori probability distributions). Previous assessments of genetic structure in *B. patronus* suggested no Gulf-wide genetic structure and high rates of migration across the Gulf (Anderson 2006; Anderson and Karel 2014). Therefore, the Structure analysis was completed assuming admixture among populations and also allele frequencies that were correlated among populations. Structure runs consisted of 10,000 sampling iterations followed by 90,000 cluster iterations; the number of iterations was determined after observing convergence of model parameters in

trial data runs. At each level of K , 10 replicate runs with identical run length and model parameters were used to determine whether results achieved consistency among runs. This analytical framework was repeated with samples of *B. smithi*.

Multiple marine taxa have demonstrated genetic structure across the northern Gulf, and in many cases this structure can be related to taxa occurring roughly east and west of the Mississippi River drainage (Dahlberg 1970; Portnoy and Gold 2012). A previous examination of the distribution of morphological characters among the North American forms of *Brevoortia* suggested that *B. gunteri* and *B. smithi* were sister taxa that had diverged according to this general pattern (Dahlberg 1970). Although no such pattern has yet been identified in populations of *B. patronus*, tagging data suggests that movement across the Mississippi Delta is infrequent (Pristas et al. 1976). The hypothesis that there may be divergent populations of *B. patronus* in the eastern and western Gulf was tested explicitly using the analysis of molecular variance (AMOVA; Excoffier et al. 1992). The AMOVA framework assessed genetic structure at three levels: among groups (western versus eastern Gulf), among samples, and within samples (among individuals). Divergence was assessed using F -statistics derived from the AMOVA sum of squares, and permutation tests ($n = 1,000$) were used to assess statistical significance of each F -statistic. A significantly positive F at any level was assumed to indicate significant genetic divergence. The AMOVA and all associated statistical testing were carried out using the software Arlequin version 3.5 (Excoffier et al. 2005).

Genetic diversity and N_e .—The central–marginal hypothesis was tested in *B. patronus* with the null hypothesis (H_0) that genetic variability was distributed evenly throughout the sampled range. The central–marginal hypothesis posits that, across a species range, genetic variability will be highest in populations at the center of the range due to larger effective population sizes and greater rates of migration than what is expected in marginal populations (Eckert et al. 2008). This mechanism may be even greater in *B. patronus* given that tagging data suggest that adults migrate towards the center of the species distribution over time (Ahrenholz 1981). Estimates of genetic variability were generated for each *B. patronus* sample using multilocus heterozygosity and the estimator θ_H (derived from Ohta and Kimura 1973), as implemented in Arlequin 3.5. Estimates were replicated using early (2002–2003) and late (2012–2014) data sets independently, and a t -test was used to determine whether mean values of θ_H from each sampling period differed significantly. Genetic variability, θ_H , was plotted against longitude to qualitatively assess whether θ_H was higher in middle longitudes or consistent across the species range. To quantitatively test for higher θ_H in the center of the species distribution, the Mississippi River was used to anchor the distribution of samples, and linear shoreline distance from the Mississippi River to each sample locale was approximated using the plot tool in Google Earth (Google, Mountain View, California). The relationship between this distance and θ_H was

tested for statistical significance using least-squares regression. The mean θ_H over all samples and all loci was used to indirectly estimate effective population size of *B. patronus* Gulf-wide, using the formula $\theta_H = 4N_e\mu$, where μ is the study-wide mutation rate of all microsatellite loci. Estimates of N_e were generated under fast ($\mu = 1 \times 10^{-3}$) and slow ($\mu = 1 \times 10^{-5}$) assumed mutation rates.

The effective population size (N_e) of *B. patronus* was also estimated directly from genotypic data using two methods: (1) the bias-corrected linkage disequilibrium (LD) method (Hill 1981; Waples 2006; Waples and Do 2010), and (2) the moment-based temporal method (Waples 1989). Initial estimates generated using the software NeEstimator 2.1 (Do et al. 2014) suggested that point estimates of N_e using the single-sample LD method usually were very large and had confidence intervals that overlapped infinity. Therefore, confidence intervals around each estimate were used to evaluate a minimum value of N_e . In particular, the lower bounds of estimates of N_e have value from a conservation standpoint and relate directly to the long-term adaptability of populations (Nelson and Soulé 1987). The methods used for N_e estimation are generally sensitive to low frequency alleles; in each case, estimates were generated while throwing out individuals with low-frequency alleles using three threshold frequencies: 0.01, 0.02, and 0.05.

Only the five Texas sampling locales that were identical between sampling time periods were used for N_e estimation. Estimates of N_e based on the single-sample LD method were generated independently for both the early and late samples; in each case, samples were pooled into a single population. Pooling samples was justified by a lack of genetic structure found among sample locales (see Results) and resulted in more robust sample sizes for N_e estimation. The temporal method required the use of both data sets simultaneously and an estimate of the number of generations that occurred between samples. Generation time was set at $t = 1.5$ years to account for the expected “knife-edge” maturity that occurs between ages 1 and 2 (Vaughan et al. 2007), as well as high estimated natural mortality and short life span of *B. patronus* (Ahrenholz 1981; Vaughan et al. 2000, 2007). The estimator F_s (Jorde and Ryman 2007) was used for the temporal estimate of N_e ; preliminary runs using NeEstimator suggested that this F_s was generally not as sensitive as other estimators to rare alleles in the Gulf Menhaden data set (e.g., Nei and Tajima 1981; Pollak 1983).

RESULTS

Genetic Species Identification

All three species of *Brevoortia* known to be present in the Gulf were observed in the data set based on genetic species identification (Table 1). The most commonly observed species was *B. patronus* (overall $n = 750$), which was present in 17 out of 19 samples. The second most commonly observed species was *B. smithi* ($n = 126$), occurring in four Florida samples. *Brevoortia gunteri* was only observed twice, once each in

early (2002) and late (2013) samples in the upper Laguna Madre, Texas. The contrasting distributions of *B. patronus* and *B. smithi* in Florida suggest that the latter species numerically replaces the former somewhere south of Apalachicola, Florida, although this finding is tenuous given the limited sampling in this study. Five hybrid genotypes were observed, all of which occurred in Florida samples. No attempt was made to resolve the genetic status of hybrids (F_1 versus late-generation back-cross), although the genetics of hybrids has been examined previously (Anderson and Karel 2007).

Interestingly, when the small-scaled menhaden, *B. gunteri* and *B. smithi*, were analyzed together using Structure, the model representing the most basic level of population structure ($K = 2$) did not conclusively resolve the species (Figure 2). Specifically, in the $K = 2$ model, the two individuals sampled in the upper Laguna Madre (presumed to be *B. gunteri*) were not conclusively assigned to

either population cluster based on admixture scores (individual admixture proportions for individual 1 = 0.40/0.60 and individual 2 = 0.32/0.68). This could be interpreted one of two ways: (1) the two individuals identified tentatively as *B. gunteri* based on location of catch (Laguna Madre) were actually *B. smithi* that had travelled farther west than what would be expected based on their species range, or (2) shallow lineage divergence between *B. gunteri* and *B. smithi* has resulted in multiple common alleles, resulting in difficulty in resolving these species using the given microsatellite markers. This latter interpretation is strengthened by the fact that the markers used in this study were heterologous (originally developed for *B. patronus* and *A. sapidissima*) and had low genetic variation in *B. gunteri* and *B. smithi* (Anderson and Karel 2014). An additional Structure run at $K = 3$, and using sampling location as a model prior, conformed to geographic expectations and easily distinguished the species.

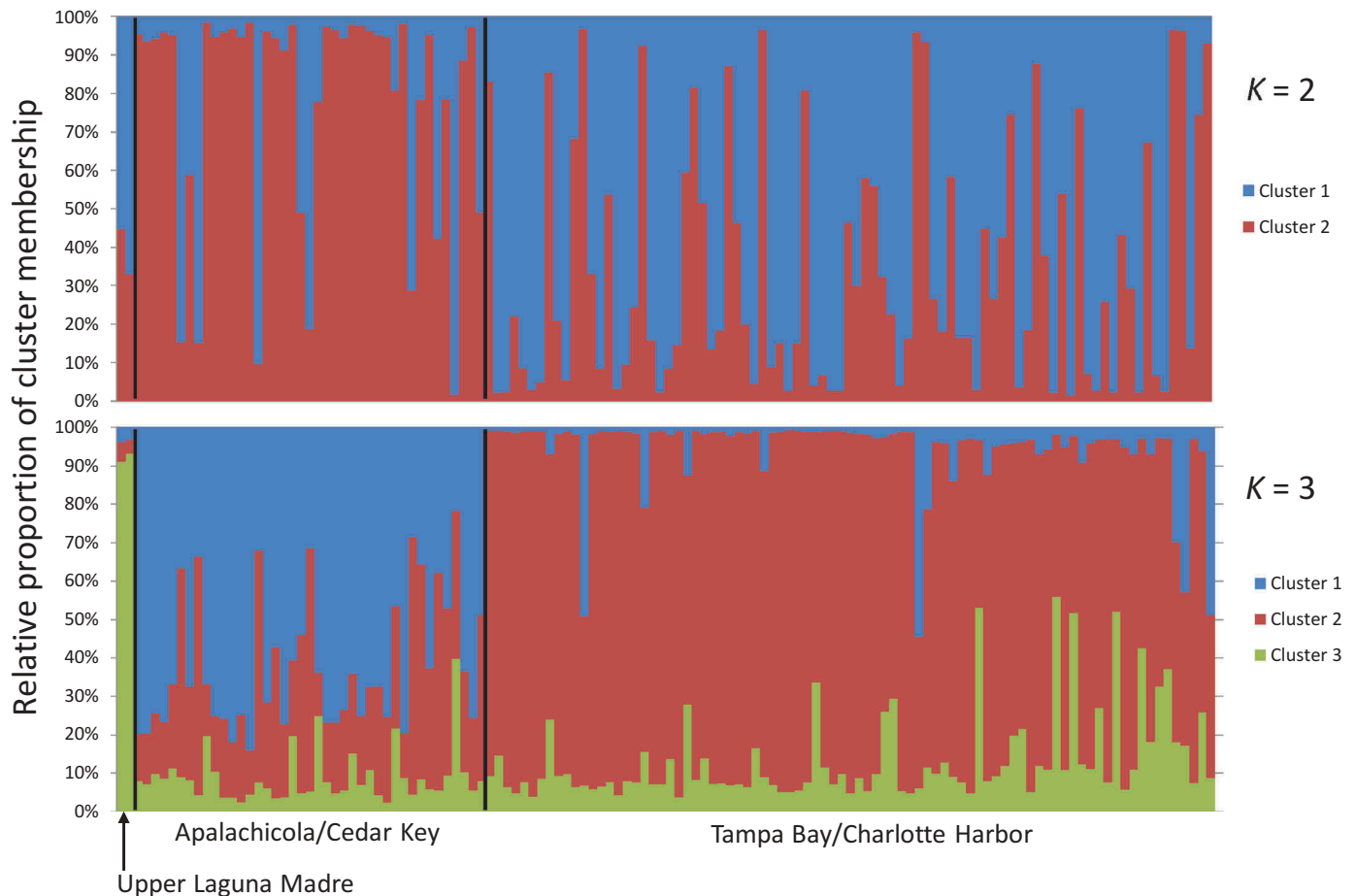


FIGURE 2. Bar graph of Structure analysis clustering results at two different levels of population structure ($K = 2, 3$) for composite small-scale menhadens (*Brevoortia gunteri* and *B. smithi*). Each bar on the graph represents a single individual; individuals are arranged in order of sample longitude from west to east. The first two individuals are presumed *B. gunteri*; all other individuals are presumed *B. smithi* (both presumptions are based on known species distributions). The proportion of membership in each genetic cluster is represented by relative proportions of bar colors.

Microsatellite Diversity and Genetic Structure

The expected heterozygosity H_e exceeded that of observed heterozygosity H_o at each genetic locus in *B. patronus* (Table 2). This resulted in positive F_{is} at all loci and an overall estimate of $F_{is} = 0.087 (\pm 0.029)$. Only a single locus (AF39660) had an exceptionally high estimate of F_{is} that exceeded the jackknifed estimate over all loci, indicating the possibility of null alleles. This locus was retained for genetic structure analysis but was excluded when estimating sample-wise genetic variability (θ_H) and effective population size (N_e) in *B. patronus*. In *B. smithi*, the overall F_{is} was higher than in *B. patronus* ($F_{is} = 0.116 \pm 0.055$), and five loci had significantly high estimates of F_{is} (BP003, BP039, BP221, BP500, AF39660). Low genetic diversity and deviation of these loci from Hardy–Weinberg expected genotypic frequencies in *B. smithi* has been demonstrated previously and likely is the result of using heterologous loci (Anderson and Karel 2014). As a result of high estimates of F_{is} , as well as limited sampling of *B. smithi*, sample genetic variability (θ_H) and effective population size (N_e) were not estimated in this species.

Structure analysis of *B. patronus* samples suggested a single genetic stock that comprised all samples. The single population ($K = 1$) model had the highest posterior probability of all tested models ($K = 1–3$) in each independent run. Models that assumed more complex population structure ($K = 2, 3$) resulted in admixture proportions for each individual that were approximately equal mixtures of each of the genetic clusters retained (equivocal

with respect to geography). The AMOVA that separated samples into eastern and western Gulf components also showed no evidence of genetic divergence between groups ($F_{ct} < 0.001$, $P = 0.403$), nor was there evidence for significant divergence among samples (within groups; $F_{sc} < 0.001$, $P = 0.844$). A post hoc examination of pairwise estimates of the genetic differentiation index, F_{st} , (among all samples) suggested no significant genetic divergence between any two samples after adjustment for multiple tests performed simultaneously.

In contrast to *B. patronus*, there was evidence for significant genetic structure among samples of *B. smithi*. Specifically, samples taken in Apalachicola and Cedar Key demonstrated a majority of genetic contribution (based on individual admixture scores) from cluster 1, which contrasted with Tampa Bay and Charlotte Harbor samples, which had a majority of contribution from cluster 2 (Table 3; Figure 2). This result suggests there is a “northern” and “southern” population of *B. smithi* in Florida. The AMOVA of *B. smithi* samples included the three largest samples (Cedar Key, Tampa Bay, and Charlotte Harbor) and also suggested significant divergence among samples ($F_{st} = 0.015$, $P = 0.027$). A post hoc examination of pairwise F_{st} scores suggested that divergence between Cedar Key (from the northern population) and the two southern samples drove the significant F_{st} in the AMOVA. The F_{st} between Cedar Key and Tampa Bay was $F_{st} = 0.019$, $P < 0.001$; the F_{st} between Cedar Key and Charlotte Harbor was $F_{st} = 0.016$, $P = 0.006$.

TABLE 2. Observed (H_o) and expected (H_e) heterozygosity, and estimated coancestry coefficient (F_{is}) of each genetic locus and overall among *Brevoortia patronus* and *B. smithi* samples observed in this study. The confidence intervals around individual locus estimates of F_{is} were compared with the mean estimate of F_{is} (jackknifed over loci) to determine which loci exceeded the overall estimate (statistically high F_{is} values are marked with an asterisk).

Locus	<i>Brevoortia patronus</i>			<i>Brevoortia smithi</i>		
	H_o	H_e	F_{is}	H_o	H_e	F_{is}
BP003	0.656	0.700	0.079	0.170	0.235	0.278*
BP039	0.619	0.646	0.043	0.000	0.013	1.000*
BP121	0.804	0.849	0.056	0.706	0.702	0.000
BP221	0.813	0.836	0.036	0.246	0.318	0.194*
BP230	0.825	0.908	0.087	0.679	0.657	0.000
BP473	0.572	0.578	0.002	0.400	0.452	0.133
BP489	0.144	0.156	0.076	0.007	0.007	0.002
BP500	0.886	0.894	0.018	0.047	0.115	0.559*
BP531	0.850	0.900	0.055	0.611	0.666	0.089
AF39657	0.646	0.680	0.050	0.111	0.106	0.000
AF39658	0.734	0.826	0.112	0.480	0.525	0.100
AF39660	0.228	0.532	0.586*	0.070	0.366	0.773*
AF39661	0.113	0.128	0.179	0.187	0.171	0.000
AF49462	0.816	0.899	0.102	0.610	0.636	0.046
Overall	0.622	0.681	0.087	0.309	0.355	0.116

Genetic diversity and N_e

The estimates of θ_H among all samples of *B. patronus* ranged from 1.77 (upper Laguna Madre, 2002) to 1.90 (Galveston Bay, 2013). There was no significant difference in θ_H between early and late samples ($t = 0.61$, $P = 0.55$). Qualitatively, θ_H appeared to be elevated in northern Gulf samples relative to the edges of the species range, and this effect was replicated across both sampling periods (Figure 3).

TABLE 3. Proportion of membership of each small-scaled menhaden (*Brevoortia smithi* and *B. gunteri*) sample in genetic clusters, generated by Structure analysis, and the samples size (n) used for each sample location. The upper Laguna Madre sample was presumed to be two *B. gunteri* individuals based on the known distribution of each species; all other samples were assumed to be composed of *B. smithi*.

Sample	Cluster 1	Cluster 2	Cluster 3	n
Upper Laguna Madre, Texas	0.04	0.04	0.92	2
Apalachicola, Florida	0.78	0.14	0.08	4
Cedar Key, Florida	0.60	0.30	0.09	37
Tampa Bay, Florida	0.03	0.87	0.10	50
Charlotte Harbor, Florida	0.10	0.71	0.19	35

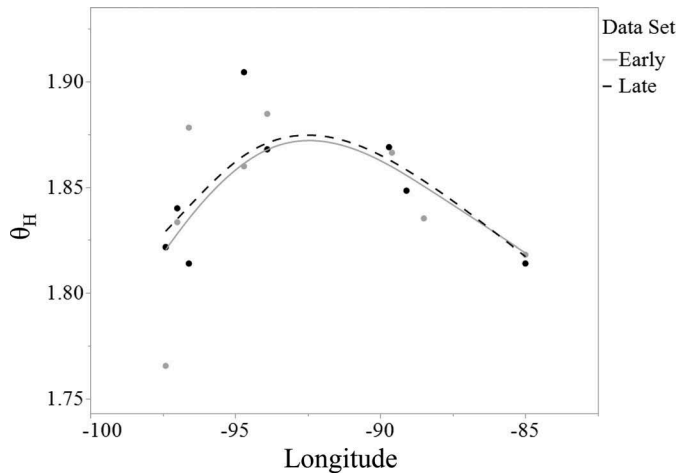


FIGURE 3. Plot of genetic variability (θ_H ; Ohta and Kimura 1973) by sample longitude. The two shades (light gray, black) represent independent data sets from early (2002–2003) and late (2012–2014) sampling periods. Data points are sample estimates of θ_H (derived from the stepwise mutation model of Ohta and Kimura 1973). The line fit is a spline smoother generated with a penalized least-squares approach and smoothing parameter ($\lambda = 0.25$).

However, the model of regression of θ_H against sample distance from the Mississippi River was not significant ($F = 0.07$, $P = 0.79$). The mean θ_H over all loci and all samples was 1.85, which equated to a range of N_e from 462 to 46,125 depending upon usage of slow versus fast mutation rates.

Point estimates of N_e using the LD method were unstable, ranging from 1,437 individuals to ∞ in the early samples (depending on low allele frequency threshold; Table 4). In late samples, point estimates at all cutoff levels were $\sim\infty$. The lowest bounds of the confidence interval for any allele

TABLE 4. Estimates of effective population size (N_e) in Texas samples of *Brevoortia patronus*. Estimates were generated using two methods: the single-sample linkage disequilibrium (LD) method and the two-sample temporal method. In the former case, estimates of N_e in early and late samples were generated independently. In all cases, N_e was estimated under three low-frequency allele thresholds (0.05, 0.02, and 0.01).

Sample	Confidence interval	Allele threshold		
		0.05	0.02	0.01
Early (LD method)	N_e estimate	1,437	∞	4,019
	Lower	611	2,309	1,422
	Upper	∞	∞	∞
Late (LD method)	N_e estimate	∞	∞	∞
	Lower	1,139	7,674	11,964
	Upper	∞	∞	∞
Both samples (temporal method)	N_e estimate	1,201	1,372	1,435
	Lower	819	1,018	1,094
	Upper	1,655	1,778	1,822

frequency cutoff threshold were 611 (in early samples) and 1,139 (in late samples). The temporal method yielded more stable estimates and confidence intervals of N_e . Point estimates using the temporal method ranged from 1,201 to 1,435, and the lower and upper bounds of confidence intervals ranged from 819 to 1,822.

DISCUSSION

The modern purse seine fishery for menhaden is centered between Cameron, Louisiana, and Moss Point, Mississippi, based on the location of reduction facilities (Smith 1991; SEDAR 2013). Nominal harvest takes place outside of this range, but close to 95% of the harvest takes place in this general geographic distribution (Smith 1991; SEDAR 2013). Previous assumptions about the taxonomic makeup of the menhaden catch have assumed <1% of the catch to be *B. gunteri* and *B. smithi* (Ahrenholz 1981). In this study, neither of the congeneric species observed (*B. gunteri*, *B. smithi*) were found in the range from Sabine Lake, Texas, to Moss Point, which roughly encompasses the fished area. These data generally support previous assumptions about the composition of the fishery and suggest that *B. patronus* likely constitute >99% of the menhaden catch in the targeted area. Harvest in areas outside of this range, particularly in Florida, may encounter other species on a more regular basis. However landings in Florida have historically accounted for <1% of the total annual catch of Gulf Menhaden (SEDAR 2013), and therefore the impact of harvest on *B. smithi* is likely to be minimal, assuming that sampling in this study is representative of species presence–absence.

With regard to population structure, previous studies of other marine clupeids have demonstrated gene flow over vast spatial scales, resulting in basinwide genetic homogeneity at neutral loci in several species (e.g. Ryman et al. 1984; Kinsey et al. 1994; Mariani et al. 2005; Lynch et al. 2010). The commonly observed pattern of basinwide homogeneity in marine fishes in general has generally been attributed to large population sizes and limited barriers to dispersal that typically characterize marine systems (Waples 1998). The biology of Gulf Menhaden, in particular the presence of an offshore egg–larval stage and the potential for vast migration at both the larval and adult life stages, suggests a predisposition for this species to conform to the expectation of genetic homogeneity over the range of the species. Two independent tests of this assumption here suggest a lack of localized genetic divergence in Gulf Menhaden across the sampled range. A naive Structure cluster analysis that made no assumptions about population boundaries suggested that all samples collected from Texas to Florida resided in a single genetic cluster. Additionally, a model-based AMOVA that assumed the Mississippi River as a boundary between independent populations was not supported by elevated levels of divergence. In fact, this latter analysis suggested that there was

not a single pairwise group of samples with elevated genetic divergence between them, confirming basinwide genetic homogeneity in the Gulf. This finding was also implied by results from previous studies (Anderson 2006; Anderson and Karel 2014) and strongly supports a single-stock hypothesis for *B. patronus* in the Gulf.

In contrast to the basinwide homogeneity observed in *B. patronus*, there was significant genetic divergence among localized populations of *B. smithi*. This result was elicited both by a Structure analysis that suggested two genetic clusters in *B. smithi* and also by elevated levels of a traditional genetic divergence metric (F_{st}) in comparisons between northern and southern Florida samples. Although the observed genetic divergence among samples was low ($F_{st} = 0.015$), it was statistically significant. Moreover, pairwise estimates of divergence among various samples of *B. smithi* resulted in a repeated pattern; specifically, two independent southern samples (Tampa Bay, Charlotte Harbor) displayed significantly elevated divergence with a single northern sample (Cedar Key). These results imply limited gene flow among localized groups of *B. smithi* in Florida and are similar to the result described in Anderson (2007), although in that case the divergence was higher ($F_{st} = 0.052$) and was observed between Gulf and Atlantic populations of *B. smithi* rather than two Gulf populations. The biology of *B. smithi* with respect to distribution may play a role in the establishment of genetically distinct populations in this species. For instance, *B. smithi* generally reside inshore and do not undertake the vast offshore migrations observed in *B. patronus* (Dahlberg 1970). Additionally, *B. smithi* may not form the vast schools observed in *B. patronus* (Hildebrand 1963) and likely has a smaller effective population size relative to *B. patronus* and *B. tyrannus* (Anderson 2007). Taken together, these factors (inshore distribution, limited migration, relatively low N_e) could be expected to result in elevated genetic divergence between localized populations. This interpretation is striking when viewed in the context of genetic homogeneity of the congeneric *B. patronus* across the entire Gulf, as well as *B. tyrannus* in the Atlantic Ocean (Lynch et al. 2010), and may hold implications towards the mechanisms that initially drove speciation in the various North American species of *Brevoortia*.

Estimates of N_e in *B. patronus* using the LD method were very high and sensitive to the presence of rare alleles. This could potentially be the result of effective population sizes that are higher than can be accurately estimated with the available data; a result that can be interpreted in two interrelated ways: (1) the N_e in Gulf Menhaden is significantly large that the LD method is only useful with extremely large sample sizes, and (2) the number of loci or individuals examined here was not high enough to detect the expected LD generated by a large but finite Gulfwide population. Whatever the reason for unstable estimates of N_e , the point estimates generated using both the historical and recent data sets generally approached infinity, and confidence intervals around point estimates using

a variety of estimators suggested an absolute lower bound of $N_e = 611$. Confidence intervals generated using the temporal method implied a lower bound of $N_e = 819$ but suggested more conservative point estimates ($1,201 < N_e < 1,435$). In addition to the instability in LD estimates, high gene flow and insufficient sampling could also create downward bias in estimates of N_e using the temporal method (Wang and Whitlock 2003; Waples and Do 2010; Hare et al. 2011). In general, these estimates that suggest $N_e > 1,000$ are supported by previous examinations of Gulf Menhaden reporting exceptionally high genetic variability based on mtDNA (Bowen and Avise 1990; Anderson 2007; Lynch et al. 2010) and microsatellite (Anderson 2007; Lynch et al. 2010) diversity.

With regard to the spatial distribution of genetic diversity, estimates of θ_H from Gulf-wide sampling of *B. patronus* suggest that genetic diversity is highest in the northern Gulf, towards the center of the species range. This result was repeated in both historical and contemporary sampling, suggesting repeatability in this pattern between two data sets. Based on tagging data, Ahrenholz (1981) suggested that adult menhaden migrate towards the center of the species range as they age. This migratory pattern would result in frequent immigrants into the center of the range, with immigrant frequency declining as distance from the center of the range decreases. Such a pattern could be expected to generate the spatial distribution of genetic diversity observed here. It would also potentially result in the level of gene flow that would be necessary to homogenize allele frequencies across the species range, resulting in a lack of discernible genetic structure. Our finding that no genetic structure was observed among spatial samples in this study may imply just such a mechanism.

If the finding that genetic variability is highest in the center of the species range is a biological reality, it implies that the fishery generally targets individuals in an area with the highest potential for genetic adaptability and that the fishery has likely had a minimal impact on this central–marginal pattern. The caveat to this point is that the data failed to show a statistical pattern with respect to this distribution of genetic variability, as θ_H was not significantly correlated with distance from the Mississippi River. Furthermore, the genetic variability statistic θ_H is influenced heavily by sample-level genetic heterogeneity and thus is heavily biased with respect to sampling error or allele-size calling error. Ideally, the central–marginal pattern in *B. patronus* could be tested using sample-based estimates of effective population size, although the current data suggest that estimates of N_e are unreliable even with very large sample sizes. Genomic sampling methods likely will be necessary to generate proper estimates of N_e and test the central–marginal pattern conclusively.

In conclusion, an inclusive data set of microsatellites designed specifically for use in *B. patronus* has repeated previous findings of genetic homogeneity in Gulf-wide samples and supports single-stock treatment of *B. patronus* in stock assessments. Given this repeated finding, it is unlikely that the

fishery might exploit rare or exceptional localized genetic diversity in areas where it operates. Because of logistical considerations, the fishery targets populations where the species is most commonly found, in the center of the species range (Smith 1991; SEDAR 2013). Based on estimates of genetic diversity, this area may be a genetic sink, where elevated rates of immigration result in high genetic diversity relative to the edges of the species range. Estimates of population-wide genetic diversity (θ_H) and effective population size (N_e) suggest relatively high genetic diversity for a marine finfish and reflect the presumed enormous census size projected from fishery-dependent estimates (Vaughn et al. 2000). Estimates of effective population size were similarly large between early and late samples, suggesting that long-term fishing pressure has not had an observable influence on N_e , although uncertainty associated with estimates of N_e makes this conclusion tenuous. In any event, using the temporal method point estimate of $N_e = 1,201$, the effective size of the *B. patronus* population in the Gulf appears to be higher than mean estimates generated for other exploited wild populations (Palstra and Ruzzante 2008), similar to or higher than estimates generated for other notable marine fish populations (Hauser et al. 2002; Turner et al. 2002; Hoarau et al. 2005), and above the conservation threshold ($N_e = 500$) that may be expected to result in long-term population adaptability (Nelson and Soulé 1987).

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REFERENCES

- Ahrenholz, D. W. 1981. Recruitment and exploitation of Gulf Menhaden. U.S. National Marine Fisheries Service Fishery Bulletin 53:3–19.
- Anderson, J. D. 2006. Conservation genetics of Gulf Menhaden (*Brevoortia patronus*): implications for the management of a critical forage component for Texas coastal gamefish ecology. Texas Parks and Wildlife Department, Federal Aid in Sport Fish Restoration Act, Project F-144-R, Final Report, Austin.
- Anderson, J. D. 2007. Systematics of the North American menhadens: molecular evolutionary reconstructions in the genus *Brevoortia* (Clupeiformes: Clupeidae). U.S. National Marine Fisheries Service Fishery Bulletin 205:368–378.
- Anderson, J. D., and W. J. Karel. 2007. Genetic evidence for asymmetric hybridization between menhadens (*Brevoortia* spp.) from peninsular Florida. *Journal of Fish Biology* 71:235–249.
- Anderson, J. D., and W. J. Karel. 2014. Limited genetic structure of Gulf Menhaden (*Brevoortia patronus*), as revealed by microsatellite markers developed for the genus *Brevoortia* (Clupeidae). U.S. National Marine Fisheries Service Fishery Bulletin 112:71–81.
- Begg, G. A., and J. R. Waldman. 1999. An holistic approach to fish stock identification. *Fisheries Research* 43:35–44.
- Bowen, B. W., and J. C. Avise. 1990. Genetic structure of Atlantic and Gulf of Mexico populations of sea bass, menhaden, and sturgeon: influence of zoogeographic factors and life-history patterns. *Marine Biology* 107:371–381.
- Carvalho, G. R., and L. Hauser. 1994. Molecular genetics and the stock concept in fisheries. *Reviews in Fish Biology and Fisheries* 4:326–350.
- Christmas, J. Y., and R. S. Waller. 1975. Location and time of menhaden spawning in the Gulf of Mexico. Gulf Coast Research Laboratory, Ocean Springs, Mississippi.
- Dahlberg, M. D. 1970. Atlantic and Gulf of Mexico menhadens, genus *Brevoortia* (Pisces: Clupeidae). *Bulletin of the Florida State Museum* 15:91–162.
- Deegan, L. A., and B. A. Thompson. 1987. Growth rate and early life history of young-of-the-year Gulf Menhaden as determined from otoliths. *Transactions of the American Fisheries Society* 116:663–667.
- Diaz, M., D. Wetthey, J. Bulak, and B. Ely. 2000. Effect of harvest and effective population size on genetic diversity in a Striped Bass population. *Transactions of the American Fisheries Society* 129:1367–1372.
- Do, C., R. S. Waples, D. Peel, G. M. Macbeth, B. J. Tillett, and J. R. Ovenden. 2014. NeEstimator V2: reimplementation of software for the estimation of contemporary effective population size (N_e) from genetic data. *Molecular Ecology Resources* 14:209–214.
- Eckert, C. G., K. E. Samis, and S. C. Loughheed. 2008. Genetic variation across species' geographical ranges: the central-marginal hypothesis and beyond. *Molecular Ecology* 17:1170–1188.
- Excoffier, L., G. Laval, and S. Schneider. 2005. Arlequin ver. 3.0: an integrated software package for population genetics data analysis. *Evolutionary Bioinformatics Online* [online serial] 1:47–50.
- Excoffier, L., P. Smouse, and J. Quattro. 1992. Analysis of molecular variance inferred from metric distances among DNA haplotypes: application to human mitochondrial DNA restriction data. *Genetics* 131:479–491.
- Goudet, J. 1995. FSTAT (version 1.2): a computer program to calculate F-statistics. *Journal of Heredity* 86:485–486.
- Hare, M. P., L. Nunney, M. K. Schwartz, D. E. Ruzzante, M. Burford, R. S. Waples, K. Ruegg, and F. Palstra. 2011. Understanding and estimating effective population size for practical application in marine species management. *Conservation Biology* 25:438–449.
- Hauser, L., G. J. Adcock, P. J. Smith, J. H. Bernal Ramirez, and G. R. Carvalho. 2002. Loss of microsatellite diversity and low effective population size in an overexploited population of New Zealand Snapper (*Pagrus auratus*). *Proceedings of the National Academy of Sciences of the USA* 99:11742–11747.
- Hildebrand, S. F. 1963. Family Clupeidae. Pages 257–454 in *Memoir 1: fishes of the western North Atlantic, part 3: soft-rayed bony fishes*. Sears Foundation for Marine Research, Yale University, New Haven, Connecticut.
- Hill, W. G. 1981. Estimation of effective population size from data on linkage disequilibrium. *Genetical Research* 38:209–216.
- Hoarau, G., E. Boon, D. N. Jongma, S. Ferber, J. Palsson, H. W. Van der Veer, A. D. Rignsdorp, W. T. Stam, and J. L. Olsson. 2005. Low effective population size and evidence for inbreeding in an overexploited flatfish, Plaice (*Pleuronectes platessa* L.). *Proceedings of the Royal Society of London B* 272:497–503.
- Jorde, P. E., and N. Ryman. 2007. Unbiased estimator for genetic drift and effective population size. *Genetics* 177:927–935.
- Kinsey, S. T., T. Orsoy, T. M. Bert, and B. Mahmoudi. 1994. Population structure of the Spanish Sardine *Sardinella aurita*: natural morphological variation in a genetically homogeneous population. *Marine Biology* 188:309–317.
- Lynch, A. J., J. R. McDowell, and J. E. Graves. 2010. A molecular genetic investigation of the population structure of Atlantic Menhaden (*Brevoortia tyrannus*). U.S. National Marine Fisheries Service Fishery Bulletin 108:87–97.

- Mariani, S., W. F. Hutchinson, E. M. C. Hatfield, D. E. Ruzzante, E. J. Simmonds, T. G. Dahlgren, C. Andre, J. Brigham, E. Torstensen, and G. R. Carvalho. 2005. North Sea herring population structure revealed by microsatellite analysis. *Marine Ecology Progress Series* 303:245–257.
- Nei, M., and F. Tajima. 1981. Genetic drift and estimation of effective population size. *Genetics* 1981:625–640.
- Nelson, K., and M. Soulé. 1987. Genetical conservation of exploited fishes. Pages 345–368 in N. Ryman and E. Utter, editor. *Population genetics and fisheries management*. University of Washington Press, Seattle.
- Ohta, T., and M. Kimura. 1973. A model of mutation appropriate to estimate the number of electrophoretically detectable alleles in a finite population. *Genetical Research* 22:201–204.
- Palstra, F. P., and D. E. Ruzzante. 2008. Genetic estimates of contemporary effective population size: what can they tell us about the importance of genetic stochasticity for wild population persistence? *Molecular Ecology* 17:3428–3447.
- Pinsky, M. L., and S. R. Palumbi. 2014. Meta-analysis reveals lower genetic diversity in overfished populations. *Molecular Ecology* 23:29–39.
- Pollak, E. 1983. A new method for estimating the effective population size from allele frequency changes. *Genetics* 104:531–548.
- Portnoy, D. S., and J. R. Gold. 2012. Evidence of multiple vicariance in a marine suture-zone in the Gulf of Mexico. *Journal of Biogeography* 39:1499–1507.
- Pristas, P. J., E. J. Levi, and R. L. Dryfoos. 1976. Analysis of returns of tagged Gulf Menhaden. U.S. National Marine Fisheries Service Fishery Bulletin 74:112–117.
- Pritchard, J. K., M. Stevens, and P. Donnelly. 2000. Inference of population structure using multilocus genotype data. *Genetics* 155:945–95.
- Raynie, R. C., and R. F. Shaw. 1994. A comparison of larval and postlarval Gulf Menhaden, *Brevoortia patronus*, growth rates between an offshore spawning ground and an estuarine nursery. U.S. National Marine Fisheries Service Fishery Bulletin 92:890–894.
- Roithmayr, C. M., and R. A. Waller. 1963. Seasonal occurrence of *Brevoortia patronus* in the northern Gulf of Mexico. *Transactions of the American Fisheries Society* 92:301–302.
- Ryman, N., U. Lagercrantz, L. Andersson, R. Chakraborty, and R. Rosenberg. 1984. Lack of correspondence between genetic and morphologic variability patterns in Atlantic Herring (*Clupea harengus*). *Heredity* 53:687–704.
- SEDAR (SouthEast Data, Assessment, and Review). 2013. 32A stock assessment report: Gulf of Mexico Menhaden. SEDAR, North Charleston, South Carolina.
- Shaw, R. F., B. D. Rogers, J. H. Cowan Jr., and T. L. Tillman. 1985. Distribution and density of *Brevoortia patronus* (Gulf Menhaden) eggs and larvae in the continental shelf off western Louisiana. *Bulletin of Marine Science* 36:96–103.
- Smith, J. 1991. The Atlantic and Gulf menhaden purse seine fisheries: origins, harvesting technologies, biostatistical monitoring, recent trends in fisheries statistics, and forecasting. *Marine Fisheries Review* 53:28–41.
- Turner, T. F., J. P. Wares, and J. R. Gold. 2002. Genetic effective size is three orders of magnitude smaller than adult census size in an abundant, estuarine-dependent marine fish (*Sciaenops ocellatus*). *Genetics* 162:1329–1339.
- Vaughan, D. S., K. W. Shertzer, and J. W. Smith. 2007. Gulf Menhaden (*Brevoortia patronus*) in the U.S. Gulf of Mexico: fishery characteristics and biological reference points for management. *Fisheries Research* 83:263–275.
- Vaughan, D. S., J. W. Smith, and M. H. Prager. 2000. Population characteristics of Gulf Menhaden, *Brevoortia patronus*. NOAA Technical Report NMFS 149.
- Wang, J., and M. C. Whitlock. 2003. Estimating effective population size and migration rates from genetic samples over space and time. *Genetics* 163:429–446.
- Waples, R. S. 1989. A generalized approach for estimating effective population size from temporal changes in allele frequency. *Genetics* 121:379–391.
- Waples, R. S. 1998. Separating the wheat from the chaff: patterns of genetic differentiation in high gene flow species. *Genetics* 89:438–450.
- Waples, R. S. 2006. A bias correction for estimates of effective population size based on linkage disequilibrium at unlinked gene loci. *Conservation Genetics* 7:167–184.
- Waples R. S., and C. Do. 2010. Linkage disequilibrium estimates of contemporary N_e using highly variable genetic markers: a largely untapped resource for applied conservation and evolution. *Evolutionary Applications* 3:244–262.
- Waples, R. S., A. E. Punt, and J. M. Cope. 2008. Integrating genetic data into management of marine resources: how can we do it better? *Fish and Fisheries* 9:423–449.