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Abstract

Background and Research Aims: Morphological and physiological effects of anthropogenic metals have been the focus of several studies in the Sierra de Huautla Biosphere Reserve (REBIOSH, Mexico) but little is known about how they affect anurans in the area. We evaluated metal bioaccumulation in anurans from streams in the REBIOSH and examined anuran communities in five sites at different positions relative to abandoned mine tailings.

Methods: Three and two sites were located upstream and downstream from mine tailings, respectively. We collected anuran community information and obtained sediment, water and liver samples for metal quantification from each site.

Results: Concentrations of Cu, Pb, Zn and Mn in water, and Zn, Mn, Fe and Cr in sediment, were significantly different among sites. We found bioaccumulation of Pb, Zn and Fe in livers of *Smilisca baudinii*, *Lithobates spectabilis*, *L. zweifeli* and *Rhinella horribilis*, species with different life traits. We found similar among-site species richness and diversity. Communities were dominated by *L. zweifeli*, *R. horribilis* and *S. baudinii*. Dominance was highest in sites distant from mine tailings. *Agalychnis dacnicolor*, *Tlalocohyla smithii* and *Hypopachus variolosus* were only present in sites located closest to mine tailings.

Conclusion: We did not detect an effect of metal bioaccumulation on anuran communities, but our findings suggest a potential effect on certain species in the protected area. Our results form a baseline for future explorations of the impacts of metals in the region.

Implications for Conservation: Toxic metal bioaccumulation has potential for reducing anuran genetic variability and altering sexual proportions and fecundity, potentially leading to anuran extirpations. We provide first evidence for metal bioaccumulation in anurans in the protected area, and one of few studies on its anuran communities. Our evidence can serve as guidance to ameliorate effects of legacy metal mining in this biodiversity hotspot.

Keywords

frog, pollution, toxicology, mining, lead, iron, toad

Introduction

Metal mining activities have an essential role in the economic development of modern societies but often have significant negative long-term impacts on natural ecosystems (Chen et al., 2019; Xiao et al., 2020) due to the generation of mine tailings rich in heavy metals and metalloids that are not biodegradable and accumulate in the environment. Metals bioaccumulate in different organisms and incorporate into trophic chains (Mussali-Galante et al., 2013a). Thus, not only are metals a risk for organisms in direct contact with a source (i.e., via inhalation or by skin contact), but also to others who

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are indirectly exposed via their food sources (Soliman et al., 2022).

Mexico has a long history of mining activities; mining accounts for approximately 4% of national gross domestic product (SE, 2020). Unfortunately, mining in Mexico has led to environmental deterioration throughout the country. For example, the state of Morelos in south-central Mexico has a long history of metal mining (e.g., silver [Ag], copper [Cu], iron [Fe], lead [Pb]). While mining activity has declined in the state (SGM, 2020), abandoned mine tailings continue to threaten natural ecosystems. Old mining operations have left a legacy of numerous solid mine tailings from which metals leach out into streams and groundwater or are dispersed by wind (CONANP, 2005).

Huautla, a small town in southern Morelos located inside the MAB-UNESCO Sierra de Huautla Biosphere Reserve (REBIOSH) is surrounded by approximately 780,000 tons of silver, lead, and zinc (Zn) mine solid tailings that have gone without proper environmental treatment (Márquez-Huitzil et al. 2022). Metals are responsible for numerous documented negative effects on local biota and humans (Mussali-Galante et al., 2013a; Mussali-Galante et al., 2013b; Tovar-Sánchez et al., 2016; Muro-González et al. 2020, Santoyo-Martínez et al., 2020; De la Cruz-Guameros et al., 2021) (Figure 1).

Anurans are particularly sensitive to metal pollution (Egusquiza-Morínigo, 2016; Kiesecker et al., 2001; Venne et al., 2006). They incorporate metals into their bodies primarily via respiration, ingestion and skin absorption (Hopkins, 2007; Ilizaliturri, 2010). Exposure to metals is one of many anthropogenic stressors threatening amphibians in tropical environments and has led to extirpations (Hopkins, 2007; Jofré et al., 2012) and alteration of community structure (Ficken & Byrne, 2013; Calderon et al., 2019). While a growing body of literature has reported on anthropogenic effects on anurans in Mexico (Hernández-Ordóñez et al., 2015, 2019), few have directly documented the effects resulting from metal exposure and bioaccumulation on their community structure (but see Suárez et al., 2016).

At least 38 species of anurans have been described in the state of Morelos (Castro-Franco et al., 2006; Ramírez-Bautista et al. 2023). Tropical dry forests in the southern portion of the state are home to at least 29 species (Castro-Franco et al., 2006). While most anurans require freshwater environments in early stages of their ontogeny, as adults they adopt life strategies that lead to differential use of areas throughout a stream ecotone. Species such as Van Vliet's Frog (*Smilisca baudinii*), Mexican Leaf Tree Frog (*Agalychnis dacnicolor*) and Dwarf Mexican Treefrog (*Tlalocohyla smithii*) are tree-living species; Zweifel's Frog (*Lithobates zweifeli*) and Showy Leopard Frog (*L. spectabilis*) live in areas surrounding permanent waterbodies; and Giant Toad (*Rhinella horribilis*) and Confusing Toad (*Incilius perplexus*) live in terrestrial environments but do not move far away from bodies of

water (Cortés-Suárez, 2017). Differential habitat use has been suggested to lead to varying potential metal bioaccumulation among species (Stolyar et al., 2008; Zhou et al., 2008; Jofré et al., 2012; Severtsova & Aguillón-Gutiérrez, 2013; Severtsova et al., 2013; Gastelum et al., 2019; Thanomsangad et al., 2019). Similarly, some evidence exists that anuran community structure can be affected by mining activities; species sensitive to metal bioaccumulation may disappear from communities close to mining operations (Ficken & Byrne, 2013; De Lucca et al., 2018; Calderon et al., 2019).

Here we document metal concentrations in water and sediment from stream habitats, and report metal bioaccumulation in *Smilisca baudinii*, *Rhinella horribilis*, *Lithobates zweifeli* and *L. spectabilis*, from sites located along a stream near Huautla. Sites were located at different positions upstream and downstream from mine tailings; we expected higher metal concentrations and bioaccumulation in areas closer to these structures. Further, we report on anuran community structure in these sites; where we expected differences in composition and diversity related to the proximity to mine tailings. This is the first report of metal bioaccumulation in anurans of the state and region, and one of few accounts of the anuran fauna in the REBIOSH.

Methods

Study Sites

The Sierra de Huautla Biosphere Reserve (REBIOSH) is a natural protected area in the southern State of Morelos (Figure. 1). REBIOSH was established in 1996, but mining (for Ag, Pb, Zn) activity had been present in the area since 1774 (Ruíz de Velasco, 1890; Volke and Velasco, 2005) and until 1991 (CONANP, 2005), when it ceased. Named mine tailings *Huautla* and *Las Presas*, located approximately at 18° 26' N-99° 01' W and 18° 27' N-99° 01' W (at an altitude of 995 m above sea level), remained at less than 500 m from streams once mining activity ceased (Mussali-Galante et al., 2013b; Márquez-Huitzil et al., 2020). Other unnamed tailings were also left close to streams (Figure 1). Much is unknown about the attributes of mine tailings; those from the *Las Presas* tailings have a pH = 8.2, a predominant particle size <45 µm, and a cation exchange capacity of 30.1 cmol (+) kg (Hernández-Plata et al., 2020). Solís (2016) reported generally high concentrations of cadmium (Cd) (48.6 mg/kg), Cu (214.8 mg/kg), Fe (28296.7 mg/kg), manganese (Mn) (605.4 mg/kg), Pb (5265.9 mg/kg) and Zn (3778.3 mg/kg) from these mine tailings.

The Huautla and Ajuchitlán streams are small (maximum width = 10.5 m), intermittent, medium gradient systems that drain a semi-arid mountainous area of REBIOSH dominated by dry tropical forest. Once the Ajuchitlán joins the Huautla, the stream runs S and E to join the Amacuzac river (Balsac

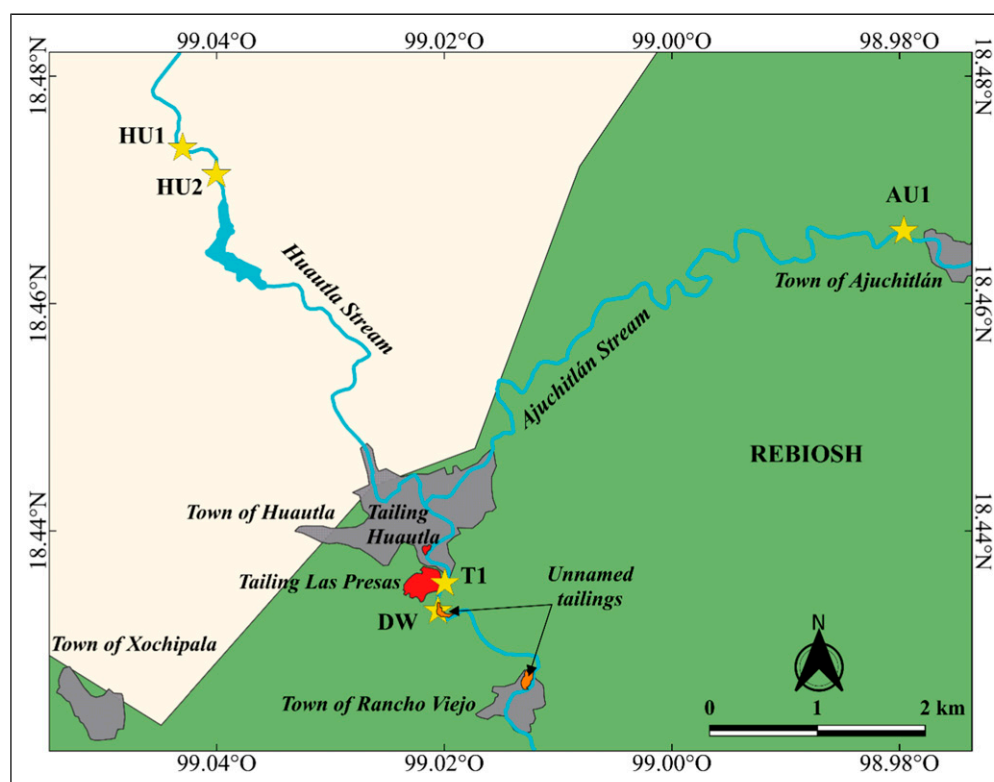


Figure 1. Study sites in the Huautla and Ajuchitlán streams in the southern portion of the State of Morelos, Mexico where anurans were collected. Sites (AU1, HU1, HU2, T1 and DW), towns and the approximate location of mine tailings are shown. The grey shading indicates the area inside the Sierra de Huautla Biosphere Reserve (REBIOSH). The orange shading indicates the location of unnamed tailings within the Las Presas tailings, according to data from [Mussali-Galante et al. \(2013b\)](#) and [Márquez-Huitzil et al. \(2022\)](#).

Basin). The study sites were located along the Huautla and Ajuchitlán streams. The Huautla runs generally N to S from the town of Quilamula, feeds the Lorenzo Vázquez (also known as Cruz Pintada) reservoir ($\geq 300,000 \text{ m}^3$) ([CEAGUA, 2017](#)), continues S towards Huautla where it meets Ajuchitlán Stream and crosses the town of Huautla, to continue south adjacent to the tailings. The Ajuchitlán runs generally NE to SW for about 8 km after crossing the town of Ajuchitlán and entering a relatively small gorge where a paved road transiently crosses the stream before it joins the Huautla ([Figure 2](#)). The Ajuchitlán is also dammed upstream from the town. In the dry season (November-June) both streams typically lose longitudinal connectivity with pools remaining in some areas; during the rainy season (July-October), both streams regain connectivity. Both systems have a relatively quick response in discharge after precipitation events and erosion from surrounding mountains and tailings is common. Prevailing winds in the area are from the North ([Ortíz, 2019](#)).

For this study, we sampled five sites located on Ajuchitlán ($n = 1$, site AU1, approximately 5.5 km upstream from mine tailing Huautla) and Huautla streams ($n = 4$). Sites in the Huautla were located 8.4 (HU1) and 6.2 (HU2) km upstream from the tailing Las Presas. Two other sites were located 0.4 (T1) and 1.6 (DW) km downstream from tailing Las Presas

([Figure 1](#)). Sites were chosen given their ease of access and their position relative to the mine tailings.

Sample Collection

We sampled water and sediment from each site in January 2019. Three water and sediment samples were obtained in each site. Three - 500 mL water samples were obtained from each site following methods in NMX-AA-051-2016 ([NMX, 2016](#)). All samples were preserved in the field by adding suprapure nitric acid (HNO_3) to attain $\text{pH} < 2$. Water samples were transported to the laboratory in clean polypropylene bottles and refrigerated at $4.0 \text{ }^\circ\text{C} (\pm 2.0 \text{ }^\circ\text{C})$ until further processing. Three superficial sediment samples were obtained following procedures in NMX-AA-112-SCFI-2017 ([NMX, 2017](#)) and NMX-AA-014-1980 ([NMX 1980](#)) from pools in each stream. Sediment samples were obtained with a shovel and kept in heavy duty polyethylene 500 mL bags (Whirl-Pak®) which were then transported to the laboratory at 4°C . In all, 30 (15 water and 15 sediment) samples were obtained and processed.

We sampled the anuran community and obtained liver samples for metal concentration measurements at different times of the year in 2018-2019. The anuran community was sampled in sites HU1, HU2, T1 and DW once monthly between June and

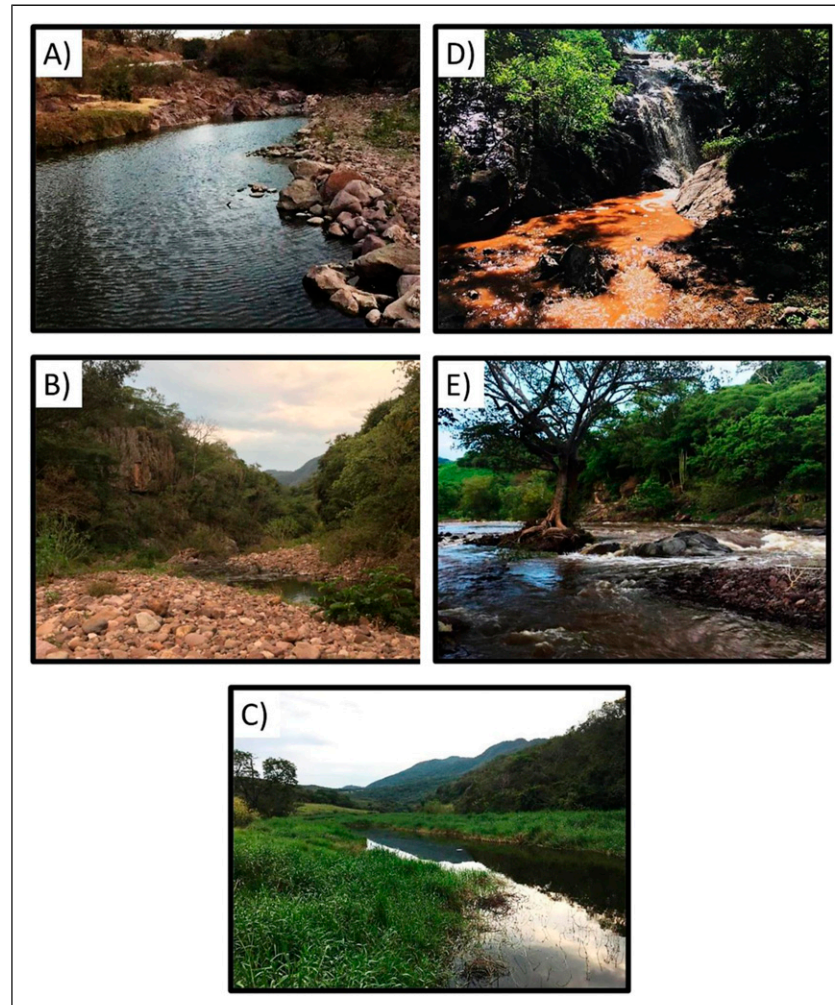


Figure 2. Sampling sites. The physical structure of the study sites is shown. The letters indicate: A) AU1, B) HU1, C) HU2, D) T1 and E) DW. All photos by Eduardo Aarón Chávez Ramírez.

September 2018; sites AU1, HU1, HU2, T1 and DW (Figure 1) were sampled once monthly in June and July 2019. Sampling periods were restricted by budgetary and logistical constraints. Six 20 x 2 m (total area = 20m²) transects were established within each site alongside the wetted portion of the stream (Scott & Woodward, 2001). We sampled for anurans within these transects. To avoid double counting of individuals, transects were separated by at least 50 m. All transects (total transects = 156) were surveyed once during each field trip by three observers at nighttime (20:00-24:00). Within each transect we carried out visual encounter surveys (VES) (Crump & Scott, 2001), in which we slowly walked along the transect looking into all possible microhabitats (to a height of 2 m above ground) where anurans could be located. Detected anurans were captured manually using nitrile gloves; each individual was placed in its own clean plastic bag until it was identified (Casas-Andreu et al., 1991).

Individuals were identified following Canseco-Márquez & Gutiérrez-Mayén (2010), Flores-Villela et al. (1995) and Frost (2021). Once identified and counted, most individuals

were released in the vicinity of the collection site, but some were kept for further processing (see below). All individuals were collected under permit SGPA/DGVS/008320/18 granted to EACR (author).

Individuals of four anuran species were selected for metal processing: *Rhinella horribilis*, *Lithobates spectabilis*, *L. zweifeli* and *Smilisca baudinii*. Selection was based on these species not being “red listed” by NOM-059-SEMARNAT-2010 (Semarnat, 2010) or the International Union for Conservation of Nature (IUCN, 2023), their ease of capture, abundance in the area, relatively large size and relatively well-known biology, including their habitat preference. *Rhinella horribilis* prefers terrestrial habitats with high humidity, including savannahs, open forests, degraded areas such as low grasslands or near human settlements (Cortés-Suárez, 2017). *Lithobates spectabilis* and *L. zweifeli* typically inhabit in proximity to permanent water bodies surrounded by deciduous tropical forest (Chávez-Ramírez, 2017). *Smilisca baudinii* typically inhabits ponds, streams and crevices with

leaves and branches (Santos-Barrera et al., 2010). Individuals captured in each site (see Table 1) were transported to the laboratory, where they were sacrificed with a local anesthetic following McDiarmid (2001). Individuals were then dissected, and their liver extracted for further processing. Individuals processed for metal concentration measurements were then fixed and deposited in the Alfonso Herrera Zoological Museum of the Universidad Nacional Autónoma de México (MZFC-HE).

Concentration of Metals in Water, Sediments and Anurans

Water and sediment samples were processed to quantify metal (Fe, Mn, Zn, Cu, chromium [Cr], Cd and Pb) concentrations.

Prior to metal concentration analyses, water samples were filtered with a 45- μ m membrane prewashed with nitric acid (1%) (see NMX-AA-051-SCFI-2001 [NMX, 2001]).

A total of 58 livers (7, 20, 14 and 17 for *R. horribilis*, *L. zweifeli*, *L. spectabilis*, and *S. baudinii*, respectively) were analyzed to determine the concentration of metals (Cd, Cr, Cu, Fe, Mn, Pb and Zn) following previous studies in the area (Mussali-Galante et al., 2013a; De la Cruz-Guarneros et al., 2021). Samples were individually dried (electric stove at 70°C, until they reached a constant weight) and sieved following the method established by the Mexican standard NMX-AA-132-SCFI-2006 (NMX, 2006). Each dehydrated liver was placed in a polyethylene terephthalate (PET) container previously washed with a 10% nitric acid solution. For metal analysis, a 0.25 g (dry weight) porcelain-pulverized portion of each liver was used. Samples were subjected to

Table 1. Mean (\pm standard error, SE) concentration of metals in water (mg/L), sediments (mg/kg), and anuran liver (mg/kg) samples from specimens captured in five sites (S) of the Huautla and Ajuchitlán streams in the State of Morelos, Mexico. Only metals with significant differences (PERMANOVA) among sites are shown. Values for sediment and water samples from nine samples obtained per site. Number of individuals examined per species are included (*R. horribilis*, n = 7; *L. zweifeli*, n = 20; *L. spectabilis*, n = 14 and *S. baudinii*, n = 17); three samples from each individual were examined for metal quantification (n = 58). # denotes the number of water, sediment and liver samples analyzed. Letters accompanying concentration values indicate significance; different letters for a metal denote significant differences for metal concentration. **UPS**) up stream; **TAL**) Tailings; **DWS**) Downstream; **nd**) not detected.

Type	Metal	UPS			TAL	DWS
		AUI Mean \pm SE	HUI Mean \pm SE	HU2 Mean \pm SE	TI Mean \pm SE	DW Mean \pm SE
Water	Pb	2.926 \pm 0.053ac	2.628 \pm 0.030a	3.365 \pm 0.042a	4.383 \pm 0.052bc	5.078 \pm 0.059b
	Cd	0.093 \pm 0.003a	0.105 \pm 0.002a	0.081 \pm 0.001a	0.008 \pm 0.002a	0.108 \pm 0.003a
	Cu	1.262 \pm 0.024a	1.464 \pm 0.018 ab	1.496 \pm 0.007 ab	1.679 \pm 0.007 ab	1.716 \pm 0.017b
	Mn	0.126 \pm 0.010a	nd	0.096 \pm 0.016ab	0.155 \pm 0.014a	0.424 \pm 0.007ac
	Zn	0.112 \pm 0.001a	0.038 \pm 0.006ab	0.120 \pm 0.003a	0.158 \pm 0.002ac	0.124 \pm 0.002a
	#	3	3	3	3	3
Sediment	Mn	1.240 \pm 0.016a	0.534 \pm 0.013b	0.479 \pm 0.037b	0.815 \pm 0.051b	1.617 \pm 0.023a
	Zn	4.123 \pm 0.339a	1.737 \pm 0.126bc	3.216 \pm 0.429ac	5.980 \pm 0.463bd	3.683 \pm 0.324a
	Cr	1.409 \pm 0.071a	1.926 \pm 0.007a	1.966 \pm 0.014b	1.675 \pm 0.046ab	1.002 \pm 0.011c
	Fe	162.050 \pm 2.058a	234.189 \pm 3.095ac	143.013 \pm 3.604bd	126.473 \pm 6.603b	198.156 \pm 3.448bc
	#	3	3	3	3	3
<i>L. zweifeli</i>	Pb	0.082 \pm 0.001a	0.005 \pm 0.00b	0.188 \pm 0.005bc	0.035 \pm 0.019ac	0.017 \pm 0.001bc
	Zn	4.346 \pm 0.192a	6.689 \pm 0.569a	0.810 \pm 0.001b	0.446 \pm 0.006ab	0.595 \pm 0.035ab
	#	6	5	1	5	5
<i>L. spectabilis</i>	Pb	nd	3.833 \pm 0.202a	2.557 \pm 0.087ab	4.007 \pm 0.085a	0.131 \pm 0.004ac
	Zn	nd	0.319 \pm 0.06a	0.570 \pm 0.012b	0.481 \pm 0.010a	7.464 \pm 0.417c
	Fe	nd	1.638 \pm 0.065a	8.107 \pm 0.784b	2.463 \pm 0.075a	25.958 \pm 0.684b
	Cu	nd	0.134 \pm 0.004a	0.428 \pm 0.008b	0.286 \pm 0.003a	0.311 \pm 0.018ab
	#	-	1	18	3	18
<i>S. baudinii</i>	Zn	nd	0.224 \pm 0.005a	6.816 \pm 0.339b	nd	1.135 \pm 0.058ab
	Fe	nd	1.436 \pm 0.080a	19.873 \pm 0.629b	nd	nd
	Cu	nd	0.214 \pm 0.007a	0.293 \pm 0.008b	nd	0.312 \pm 0.005b
	#	-	1	24	-	21
<i>R. horribilis</i>	Pb	nd	nd	nd	4.062 \pm 0.048a	2.869 \pm 0.042b
	Zn	nd	nd	nd	0.412 \pm 0.007a	0.719 \pm 0.008b
	Mn	nd	nd	nd	0.212 \pm 0.006a	0.153 \pm 0.009b
	Fe	nd	nd	nd	2.035 \pm 0.023a	6.559 \pm 0.119b
	Cu	nd	nd	nd	0.412 \pm 0.011a	1.392 \pm 0.021b
	#	-	-	-	18	3

acid digestion using an accelerated reaction system microwave (CEM®MARS-5), using 10 mL of 70% HNO₃ in closed Teflon pumps. Samples were dissolved to a final volume of 50 mL for analysis. A sample without tissue was processed concurrently and was utilized as a control. Metals were then analyzed by atomic absorption spectrophotometry (GBC-908-AA, Scientific Equipment Ltd.), a technique that determines the concentration of a metallic element in a sample, calibrating the spectrophotometer with standard solutions containing known concentrations of each analyzed item. The standard calibration curves showed correlation coefficients (r^2) between 0.99 and 1. For each metal analyzed the average value of three repeated quantifications for a sample is reported. Concentration values for water are reported in milligrams/liter. Concentration values for sediment and liver samples are reported in milligrams/kilogram (mg/kg) dry weight. The minimum detection limits (mg/L) according to the manufacturer of the spectrophotometer were 0.0004 for Cd, 0.003 for Cr, 0.001 for Cu, 0.005 for Fe, 0.0015 for Mn, 0.01 for Pb and 0.0005 for Zn. When metal concentrations were below detection limits, we used 0.5 of detection limit for statistical analyses.

Statistical Analyses

We calculated the average and standard deviation for each metal concentration for each species per site. We compared metal concentration (for each metal) among sites and per species among sites using Bray-Curtis distances from a triangular matrix using permutational multivariate analysis of variance PERMANOVA (Anderson et al., 2008). Metal concentration data used in PERMANOVA were fourth root-transformed based on type 1 sequential sums of squares (Legendre & Gallagher, 2001). In these analyses, species or sites were used as fixed factors. Each was analyzed using pairwise analyses via PERMANOVA t with 9999 permutations. P -values were calculated using Monte Carlo simulations following Anderson et al. (2008). PERMANOVA draws statistical inferences in a distribution-free process based upon geometric partitioning of distance measures. We used a principal component analysis (PCA) to explore metal concentration variability among sites (for water, sediment, and liver samples) and species. All analyses were carried out using Primer 7.0.13 (Clarke & Gorley, 2015).

We explored species richness, capture rates (number of specimens captured per sampling hour), total and relative abundance, sampling completeness and species diversity from anuran community data. Only data from replicated transects (AU1 $n = 12$; HU1-DW $n = 36$) in each site were used for these calculations and comparisons. Since temporal differences in diversity were not significant ($t_{\text{Hutchinson}} = 0.585$, $df = 4212.9$, $P = 0.558$), data from transects in each site taken on different dates were grouped for analysis. We analyzed sample completeness in all sites to assess richness variation in relation to Chao 1 and Bootstrap

estimators (Chao & Jost, 2012; Gotelli & Colwell, 2011; Moreno et al., 2011). We estimated three diversity dimensions based on Hill's numbers: Species richness or Chao 1 (q^0), the number of common species or Shannon-Wiener's exponential (q^1), and the number of dominant species or inverse Simpson (q^2) (Chao, 1984; Jost, 2006; Moreno et al., 2011). For each measure of diversity, we calculated confidence intervals (CI) at 95% which were then used to estimate among-site differences. These analyses were carried out in R (iNEXT package, V 2.0.17) (Hsieh et al., 2016). Comparisons among diversity indexes were based on Euclidean distance matrices via PERMANOVA (Anderson, 2001). We also evaluated anuran assemblage similarity using Sørensen and Jaccard indexes (Magurran, 1988; Moreno, 2001). Rank-abundance curves were generated for the assemblages in each site. These curves depict the abundance of each taxon relative to the total number of individuals in the assemblage (Whittaker, 1965).

Results

We found significant differences in metal concentrations among sites for water ($F_{\text{pseudo}} = 3.6725$, $P_{\text{perm}} = 0.0005$) and sediment ($F_{\text{pseudo}} = 172.27$, $P_{\text{perm}} = 0.0001$). The first two axes of the PCA on metals in water samples explained 79.1 % of variability, driven mainly by Pb, Cu and Zn concentrations, with Mn and Cd being relatively less important drivers (Figure 3A). The similarity percentage analysis showed differences (at $P < 0.05$) between AU1, HU1 and HU2 with T1 and DW, but not between AU1, HU1 and HU2 or between T1 and DW. Cu, Pb, Zn and Mn were mostly responsible for these differences. T1 and DW had higher Cu, Pb and Mn, compared to AU1, HU1 and HU2 (Table 1, Figure 3A).

The first two axes of the PCA analysis on metal concentrations in sediment explained 98.2% of variability among sites (Figure 3B). This variability was driven by Fe, Cr and Mn concentrations, with Zn being relatively less important. Similarity percentage analysis on metals in sediment concentrations showed differences (at $P < 0.05$ and $P < 0.001$) among all sites, with Fe, Zn, Mn, or Cr being important in explaining among site similarity depending on the sites being contrasted; no consistent differences were found among sites located close or far from mine tailings (Table 1, Figure 3B).

Metal Concentrations in Anurans

Pb, Zn, Cu, Mn, Fe and Cd bioaccumulated in livers of the examined species (Table 1). Detectable Zn and Cu concentrations were found on specimens from all sites and species. Cr was not detected in any sample. Pb was not detected in *L. zweifeli* from sites HU1 or DW. Mn was not detected in *S. baudinii* from DW, *L. spectabilis* from DW or in any *L. zweifeli*. Fe was not detected in *S. baudinii* from DW. Cd was not detected in *S. baudinii* from HU1, on *L. spectabilis* from HU1 or T1, or any *R. horribilis* (Table 1).

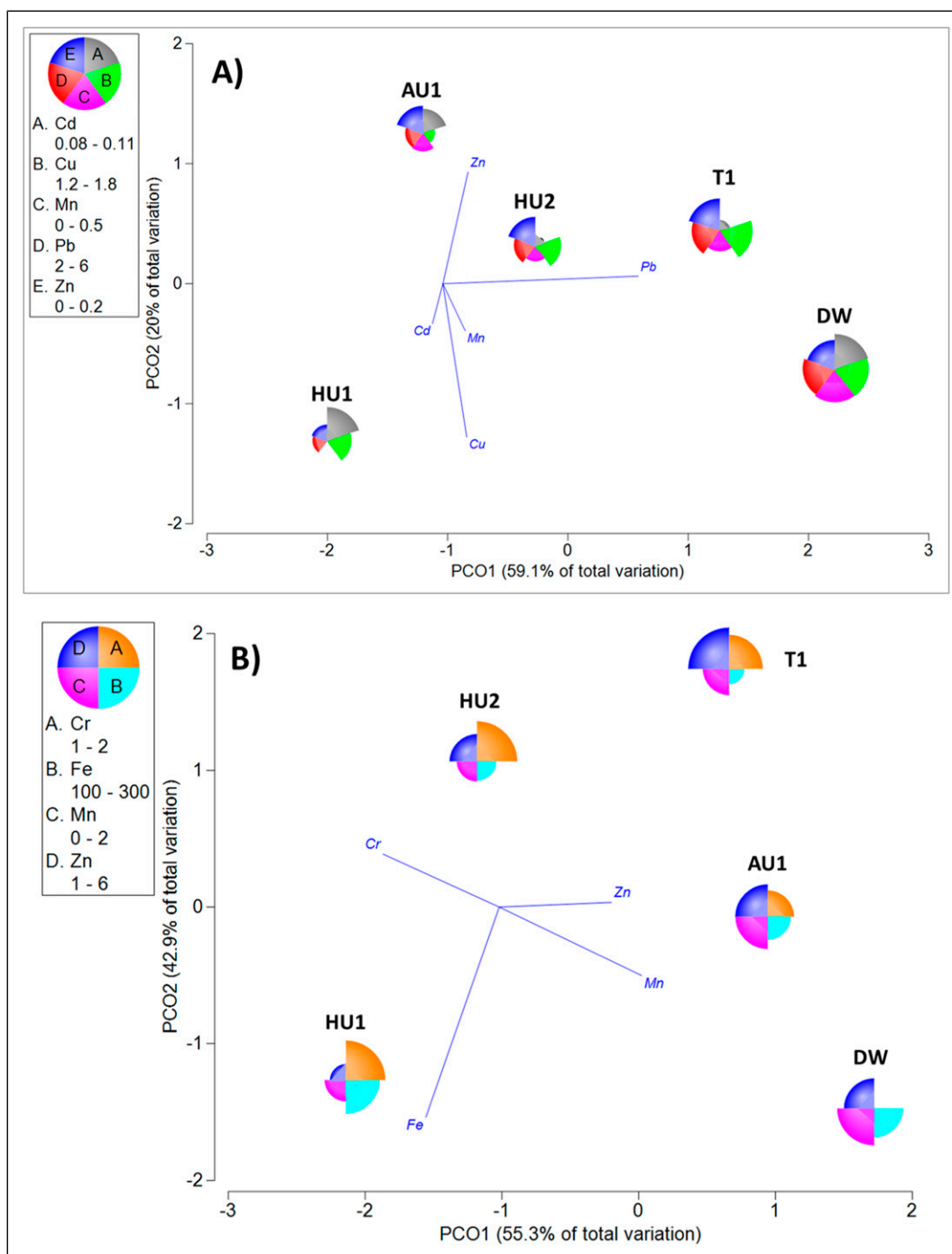


Figure 3. PCO triplot of variability in the levels of metals in A) water and B) sediment from five sites in the Huautla and Ajuchitlán streams in Morelos, Mexico. The centroids of the groups are indicated by the site label and the contribution of each metal in a site is indicated by the relative size of the pie section.

For liver we found significant differences among sites ($F_{\text{pseudo}} = 4.1874$, $P_{\text{perm}} = 0.0002$), species ($F_{\text{pseudo}} = 6.6511$, $P_{\text{perm}} = 0.0003$) and their interaction ($F_{\text{pseudo}} = 4.3124$, $P_{\text{perm}} = 0.0002$). For *L. zweifeli*, among site Fe, Cu and Cd concentrations were similar ($F = 2.07$, $df = 4$, $P\text{-perm} = 0.12$; $F = 1.68$, $df = 4$, $P\text{-perm} = 0.18$; and $F = 2.38$, $df = 4$,

$P\text{-perm} = 0.096$; respectively). Pb concentrations in *L. zweifeli* were significantly higher in site AU1 than in sites HU1, HU2, T1 and DW ($F = 8.44$, $df = 4$, $P\text{-perm} = 0.002$). Zn concentrations for this species were significantly higher in site AU1 than in site HU2 and site T1 ($F = 3.09$, $df = 4$, $P\text{-perm} = 0.018$). For *L. spectabilis*, among-site Pb

concentrations were similar ($F = 2.87$, $df = 3$, $P\text{-perm} = 0.06$); but Zn concentrations were significantly higher in sites HU2 and DW, than in sites HU1 and T1 ($F = 7.90$, $df = 3$, $P\text{-perm} = 0.006$). In *L. spectabilis*, Mn concentrations were significantly higher in site HU2 than in sites HU1, T1 and DW ($F = 4.03$, $df = 3$, $P\text{-perm} = 0.03$); concentrations from site DW were also higher than those in sites HU1 and T1 ($F = 4.03$, $df = 3$, $P\text{-perm} = 0.03$). Concentrations of Fe in *L. spectabilis* were much higher in site DW than in site HU2 and sites HU1 and T1, respectively ($F = 6.58$, $df = 3$, $P\text{-perm} = 0.007$). Cu concentrations in *L. spectabilis* were higher in site HU2 than in sites DW and sites HU1 and T1 ($F = 5.10$, $df = 3$, $P\text{-perm} = 0.01$). For *S. baudinii* Pb and Mn concentrations were similar among sites ($F = 0.92$, $df = 2$, $P\text{-perm} = 0.42$ and $F = 0.55$, $df = 2$, $P\text{-perm} = 0.612$, respectively). For *R. horribilis* median Pb ($F = 392.27$, $df = 1$, $P\text{-perm} = 0.0001$) and Mn ($F = 54.31$, $df = 1$, $P\text{-perm} = 0.0004$) concentrations were significantly higher in site T1 than site DW. In *R. horribilis*, median Zn ($F = 317.92$, $df = 1$, $P\text{-perm} = 0.002$), Fe ($F = 868.13$, $df = 1$, $P\text{-perm} = 0.001$) and Cu ($F = 76.30$, $df = 1$, $P\text{-perm} = 0.003$) concentrations were significantly higher in site DW than in site T1.

Considering metal concentrations in liver (irrespective of species), the first and second axes of the PCA analysis explained 70.1% of variability among sites. Variability was driven by Pb, Zn, Cu and Fe, with Mn and Cd being relatively less important (Figure 4A). Sites AU1, HU1, HU2 and T1 were significantly different (at $p < 0.05$ or $p < 0.001$) from DW, but no differences were found among AU1, HU1 or HU2. AU1 was significantly different from T1 ($p < 0.001$) (Table 1). These differences were mostly driven by Cu and Pb concentrations, with Mn, Zn and Fe also having some contribution.

Lithobates spectabilis, *L. zweifeli* and *S. baudinii* differed in the concentration of metals in liver, irrespective of site (Figure 4B). First and second components of a PCA on this data explained all among-species variability, and this was driven by changes in Pb, Fe and Zn, with other metals playing a relatively smaller role. Nearly all metals were found at higher concentrations in *L. spectabilis*, except Cd (higher in *L. zweifeli*) and Pb (higher in *S. baudinii*).

Generally, site \times species interaction on similarity percentages analyses for *L. spectabilis*, *L. zweifeli* and *S. baudinii* indicated significant differences in liver concentrations between sites far from (AU1, HU1 and HU2) and close to the mine tailings (T1 and DW). A PCA of such interaction explained close to 70% variability on first and second axes (Figure 4C), and was most influenced by Fe, Pb and Zn concentrations, with other metals playing a relatively smaller role. For *L. spectabilis*, significant differences were found between HU1, HU2 and T1 with DW, but not with T1. These changes were primarily driven by Pb, Mn and Cd (Table 1). For *L. zweifeli*, significant differences were found between AU1, HU1 and HU2 with T1, but not DW (Table 1); Zn and Cu concentrations were responsible for these differences. For

S. baudinii, significant differences were found between HU2 and T1 with DW and between T1 and DW; other sites did not show differences. Differences in concentrations of Pb, Fe, Zn, Cu and Mn guided these differences (Table 1).

Anuran Communities

We captured 199 anurans (nine species, eight genera, four families, Table 2) in 260 hours of sampling in wet season sampling events in 2018 ($n = 99$) and 2019 ($n = 100$). Overall average rate of capture was 0.77 ind./hr. Considering both sampling years, capture rates in site AU1 (1.18 ind./hr.) were higher than those in T1 (1.04), DW (0.83), HU2 (0.59), and HU1 (0.40). Species richness was similar among sites (Table 2). Sample completeness in each site was between 83-100 % and 75 – 100% according to Chao 1 (AU1, HU1, HU2 and T1 100%; HU1 = 83%) and Bootstrap (AU1 = 75%; HU1-HU2 = 83%; T1 = 100; DW = 89%) procedures, respectively.

DW ($n = 55$) and T1 ($n = 53$) had the highest anuran abundance and richness ($S = 8$ and $S = 7$, respectively) (Table 2). HU1 and HU2 had intermediate richness (both with $S = 5$) and AU1 had the lowest species richness ($S = 3$). The number of common and equally abundant species were significantly lower in AU1 than in HU1, HU2, T1 and DW ($F = 6.383$; $df = 4$, $P\text{-perm} = 0.0001$). Paired comparisons PERMANOVA showed differences between AU1 and HU2, T1 and DW ($P\text{-perm} < 0.01$); HU1 and HU2, T1 ($P\text{-perm} < 0.01$); HU2 and AU1-DW ($P\text{-perm} < 0.01$); T1 and AU1, HU1, HU2 ($P\text{-perm} < 0.01$); and DW and AU1, HU2 ($P\text{-perm} < 0.01$).

The most abundant and prevalent species was *Lithobates zweifeli* ($n = 74$, 5 sites) followed by *Rhinella marina* (33, 5) and *Smilisca baudinii* (31, 4); least abundant species were *Hypopachus variolosus* (2, 1) and *Agalychnis dacnicolor* (2,1) (Table 2, Figure 5). Considering all sampling events, in AU1 the most and least abundant species were *L. zweifeli* ($n = 36$) and *D. arenicolor* ($n = 1$) respectively (Table 2). Most and least abundant species in HU1 were *L. zweifeli* ($n = 15$) and *R. horribilis* ($n = 1$) (Figure 5). *Lithobates spectabilis* was the most abundant species in HU2 ($n = 13$) while the least abundant species ($n = 2$) were *L. zweifeli* and *R. horribilis*. The most abundant species in T1 were *R. horribilis* ($n = 21$) and *L. zweifeli* ($n = 11$) and the least abundant species was *H. variolosus* ($n = 2$). *Smilisca baudinii* and *L. zweifeli* were the most abundant species ($n = 15$ and $n = 10$, respectively) in DW; *D. arenicolor* was the least abundant ($n = 1$) species (Table 2, Figure 5).

Community composition was most similar (using Jaccard's index) between sites AU1 and HU1, and T1 and HU2 (Table 2). Sites HU2 and HU1, and HU2 and T1 were also similar; and HU2 and AU1 were most dissimilar (Table 2). HU2 and T1 were most similar based on Sorensen's index and HU2 and AU1 were again, most dissimilar. Generally

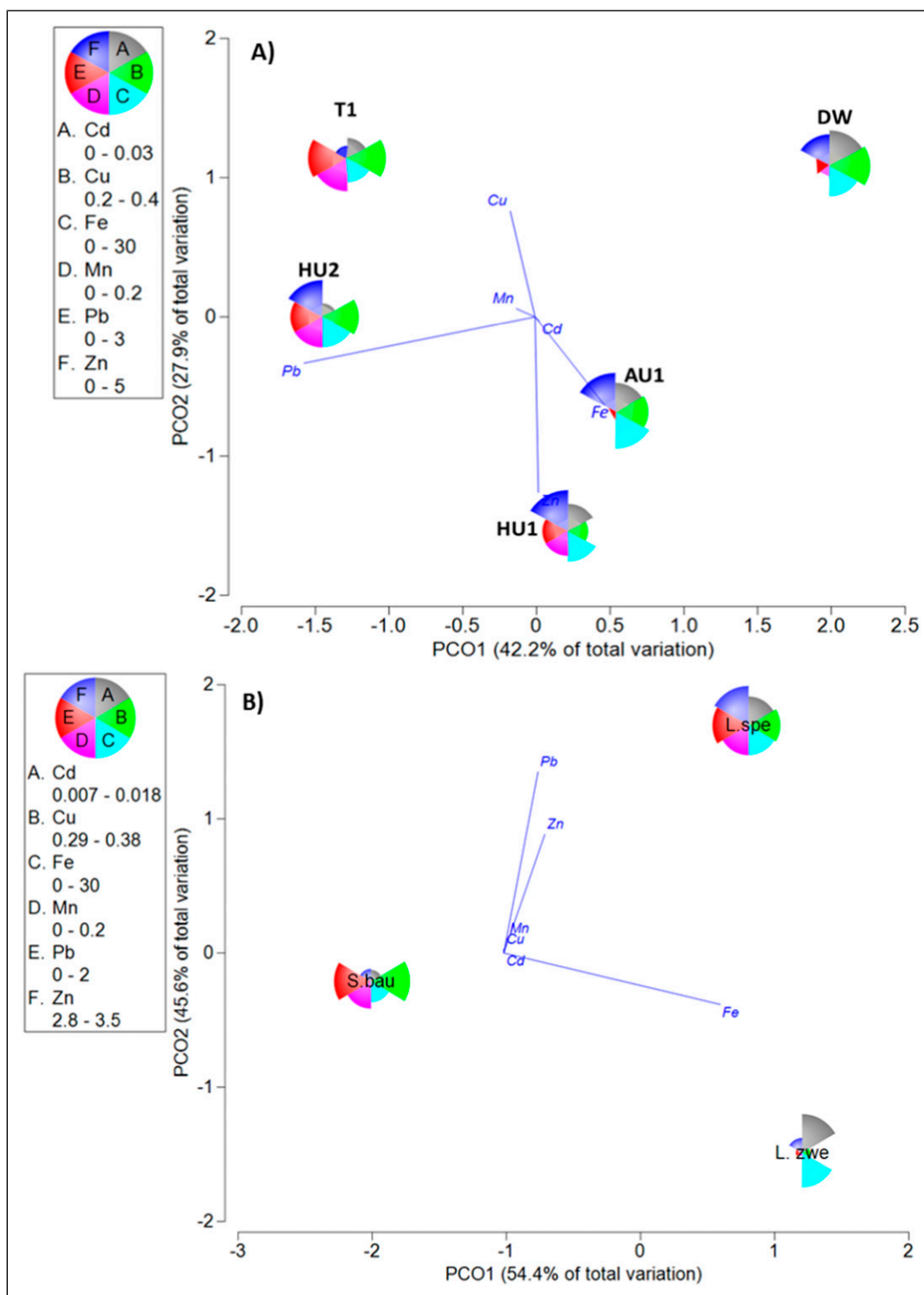


Figure 4. PCO triplot of variability in the levels of metals in amphibian livers from five sites in the Huautla and Ajuchitlán streams in Morelos, Mexico. A) Includes the contribution of each metal in amphibian’s livers by site. B) Includes the contribution of each metal in amphibian livers for a species. C) Includes the contribution of each metal in amphibian livers in a species x site interaction. Centroids of the groups (either site and/or species) are presented.

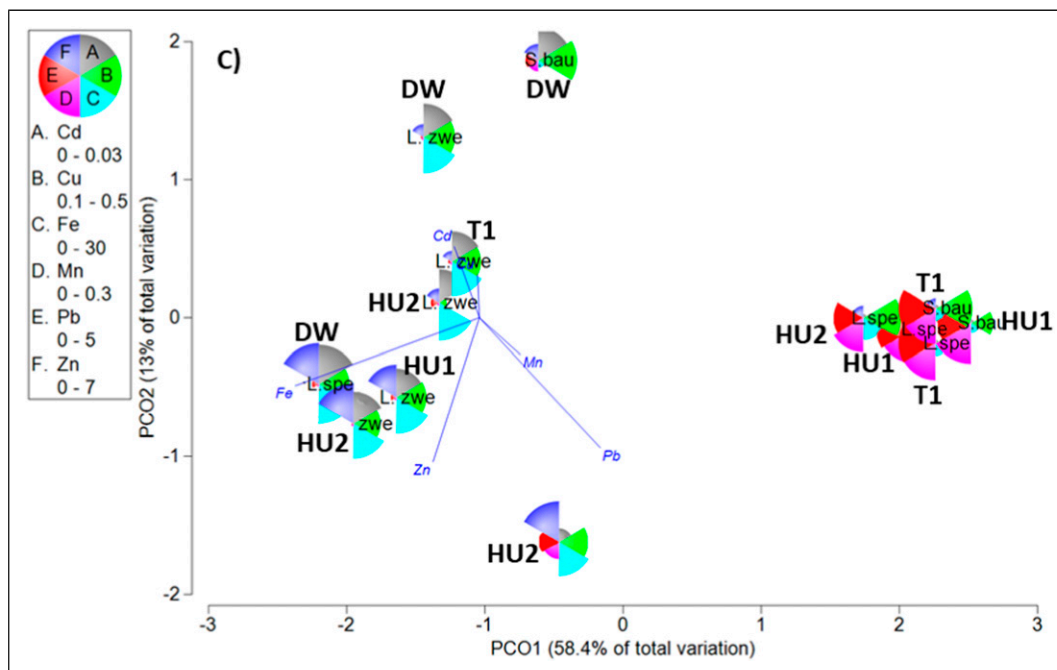


Figure 4. Continued.

speaking, AU1 was most dissimilar to other sites using either similarity index (Table 2) and species composition (Figure 3).

Discussion

Metals are known to have important consequences for the individual health of anurans and are a known factor for the decline of several anuran populations worldwide (Lee, 2000; Hernández et al., 2013; Severtsova & Aguillón-Gutiérrez, 2013; Aguillón-Gutiérrez & Ramírez-Bautista, 2015; Suárez et al., 2016). However, metal effects on anurans in the wild and at higher levels of biological organization have been poorly studied in Mexico (Ilizaliturri, 2010; Aguillón-Gutiérrez & Ramírez-Bautista, 2015). In addition to presenting updated information for the anuran community at the REBIOSH (Herrera-Balcázar, 2014; Roa-Mata, 2017), our work constitutes the first record of metal bioaccumulation for anurans in the protected area and the State of Morelos. Our findings should alert regional managers on potential threats on the anuran communities from metal exposure.

Metal concentrations in water differed significantly among all samples and sites closest to mine tailings (T1, DW) typically had the highest values. Cu, Zn and Pb in site T1, and Cd and Mn in DW, had the highest concentrations among sites. Generally, mean metal concentrations in water from the Huautla Stream were higher for Pb than Cu, Mn, Zn and Cd, in decreasing order. Metal concentrations in sediments on the other hand, showed among-site significant differences for Fe, Zn, Mn and Cr, with Mn, Fe and Zn relatively higher in T1 and DW, and Cr higher in HU2 and HU1. Zn and Mn were higher in site AU1 than in sites HU2 y HU1; Mn was also

more concentrated in DW than in AU1. Sediment metal concentrations in the Huautla Stream were higher for Fe than Zn, Cr and Mn, in decreasing order. We suggest elevated metal concentrations in both matrices for T1 and DW are due to their relative proximity to the Las Presas mine tailings. Sediment samples taken from different soil levels in these mine tailings (Solís, 2016) show higher concentrations for Fe than Zn, Pb, Mn, Cu, and Cd, in decreasing order; although all were at higher concentrations than we found in our samples. Additionally, tailings have low stability and are highly susceptible to wind and water erosion (Volke and Velesco, 2005), which could lead to metal bioaccumulation via lixiviation (Márquez-Huitzil et al., 2022) in sites T1 and DW. Pb, Cd, Cu and Zn concentrations in water samples from the Huautla Stream (0.082, 0.0018, 0.009 and 0.118 mg/L, respectively) exceeded toxicity reference values (TRV) suggested by EPA (2019). This renders water from the Huautla not recommended for human consumption. However, we note that our samples were taken during the dry season, when metal concentrations usually are higher than those in the wet season given lack of dilution of potential contaminants (Islam et al., 2017; Pandey et al., 2019; Proshad et al., 2019; 2020). Water hardness and its specific ionic properties, as well as chemical composition of substrates can lead to among-matrix metal concentration variability and bioavailability and should be studied further in the area (Miranda et al., 2022).

Contrary to our expectations, upstream sites showed elevated metal concentrations. While relatively distant from studied mine tailings, we recognize that the entire area could be subject to relatively elevated natural metal concentrations. What today is the REBIOSH was for decades one of the most

important mining districts in Morelos state and many mines and other tailings exist in the area (Volke and Velesco, 2005; Mussali-Galante et al., 2013b; Márquez-Huitzil et al. 2022). As such, the area's mineralogy is potentially elevated in metals throughout. This, in addition to geologic weathering, wind dispersal, atmospheric precipitation, erosion, bioturbation and human activity have likely accelerated metal inputs into soil and water (Proshad et al., 2017, 2019, 2020; Barats et al., 2020; Kabir et al., 2020). Finally, human waste deposition, fertilizer and pesticide use are pervasive throughout the area; these could be additional sources of metals into the region's soils and water resources (Rinklebe et al., 2019; Timofeev et al., 2018; Varol et al., 2020).

We found Pb, Zn, Cu, Mn, Fe and Cd concentrations in the livers from analyzed anurans. While some of these can be

considered essential (e.g., Zn and Fe) and were expected in livers, others (e.g., Pb and Cd) are not and their presence suggests exposure to anthropogenic contaminants. Our findings agree with other studies that report liver as one (but not the only) important target organ for metal bioaccumulation in several organisms, including anurans (de la Cruz et al., 2021; Curi et al., 2021). Diet and potential exposure to metals in water and sediment could explain this finding (EPA, 2019; Esteller et al., 2015; Rodríguez-Rodríguez, 2020; Solís, 2016). Lepidopterans, Coleopterans, Hymenopterans and even larval anurans are known food items for *R. horribilis*, *L. spectabilis*, *L. zweifeli* and *S. baudinii* (Barrios-Damián, 2006; Mendoza-Estrada et al., 2008). Many of those are primary consumers that may incorporate metals adhered to organic or inorganic material and

Table 2. Anuran community composition, community parameters and similarity indexes from five sites (AUI, HUI, HU2, TI and DW) sampled in the Huaatla and Ajuchitlán streams in the southern portion of the state of Morelos, Mexico during 2018-2019. For species data we present the number of individuals captured in each site; the number not in parenthesis is the total number of individuals captured, the number in parenthesis is relative abundance (%) for the species in a site across both years of study. For each species, next to its name we present its conservation status according to IUCN (2023). Community parameters and similarity indexes are as described in methods section.

		Collection Data					
Family	Code	Site					Total
		AUI	HUI	HU2	TI	DW	
Bufonidae							
<i>Incilus perplexus</i> <small>LC</small>	Ip	0	0	3(12)	7(13)	6(11)	16(8)
<i>Rhinella horribilis</i> <small>LC</small>	Rh	2(5)	1(4)	2(8)	21(40)	7(13)	33(16)
Hylidae							
<i>Agalychnis dacnicolor</i> <small>LC</small>	Ad	0	0	0	0	2(4)	2(1)
<i>Smilisca baudinii</i> <small>LC</small>	Sb	0	7(27)	6(23)	3(6)	15(27)	31(15)
<i>Tlalocohyla smithii</i> <small>LC</small>	Ts	0	0	0	0	9(16)	9(4)
<i>Dryophytes arenicolor</i> <small>LC</small>	Da	1(3)	2(8)	0	6(11)	1(2)	10(5)
Microhylidae							
<i>Hypopachus variolosus</i> <small>LC</small>	Hv	0	0	0	2(4)	0	2(1)
Ranidae							
<i>Lithobates spectabilis</i> <small>LC</small>	Ls	0	1(4)	13(50)	3(6)	5(9)	22(11)
<i>Lithobates zweifeli</i> <small>LC</small>	Lz	36(92)	15(58)	2(8)	11(21)	10(18)	74(37)
Community parameters							
	Sample Coverage	0.97	0.92	1.00	1.00	0.98	
	q ⁰	3	5	5	7	8	10
	q ¹	1.3	3.0	3.7	5.2	6.5	
	q ²	1.6	2.4	3.0	4.1	5.8	
	Abundance	39	26	26	53	55	199
Similarity indexes							
Sørensen		AUI	HUI	HU2	TI	DW	
	HUI	0.75	*	*	*	*	
	HU2	0.50	0.80	*	*	*	
	TI	0.60	0.67	0.83	*	*	
	DW	0.55	0.77	0.77	0.80	*	
Jaccard		AUI	HUI	HU2	TI	DW	
	HUI	0.60	*	*	*	*	
	HU2	0.33	0.67	*	*	*	
	TI	0.43	0.50	0.71	*	*	
	DW	0.38	0.63	0.63	0.67	*	

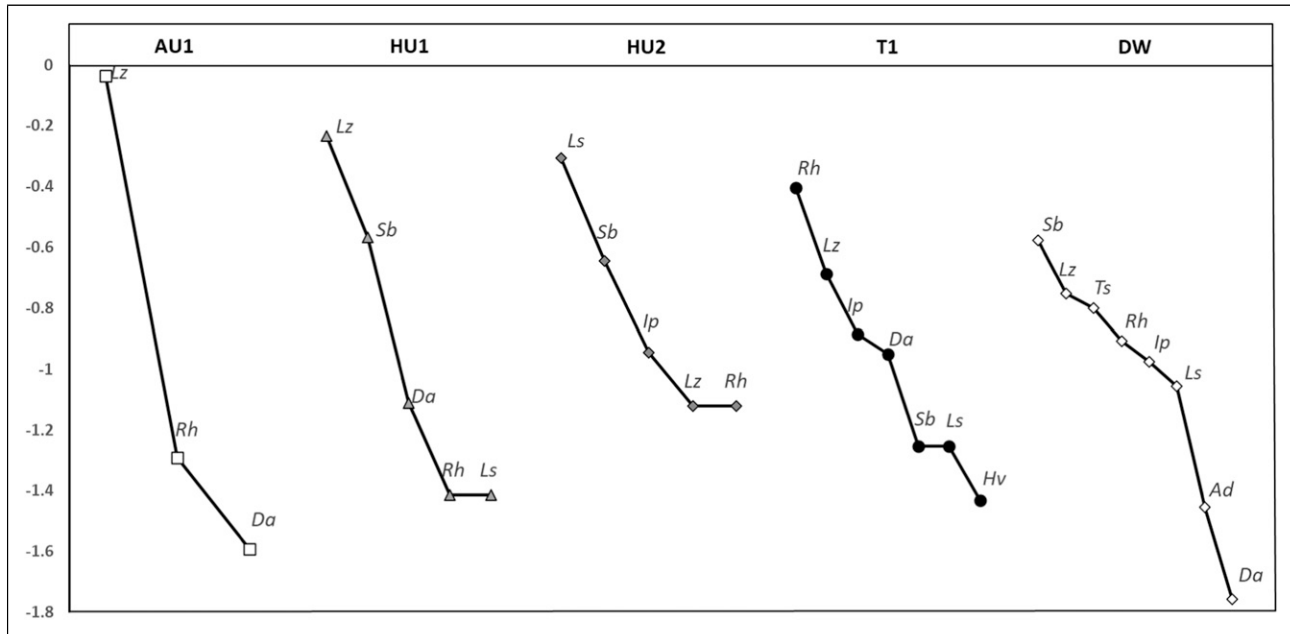


Figure 5. Rank abundance curves for anurans from five sites in the Huautla and Ajuchitlán streams, Morelos. Curves are presented independently for each site (AU1, HU1, HU2, T1 and DW). The nine species that occurred at the sampled sites are shown with codes: lp) *I. perplexus*; Rh) *R. horribilis*; Ad) *A. dacnicolor*; Sb) *S. baudinii*; Ts) *T. smithii*; Da) *D. arenicolor*; H. variolosus; Ls) *L. spectabilis*; Lz) *L. zweifeli*.

sediment in streams. Once bioavailable, metals are known to bioaccumulate and biomagnify along the food chain (Penicaud et al., 2017). An additional route for metals exposure in anurans, includes direct contact with contaminated water (Calderon et al., 2019; Jofré et al., 2012). Highly permeable skin, typically involved in respiration, is a well-known route for metal bioaccumulation in amphibians (Sparling, 2017).

While our focus was not to detect anomalies or other effects of metal accumulations in Huautla anurans, we note that Pb, Fe, Cd, Mn, Zn and Cu are known to cause effects at the individual, population and community levels in anurans. Elevated concentrations of Pb and Fe can cause morphological abnormalities in developing anurans (Severtsova et al., 2013; Severtsova & Aguillón-Gutiérrez, 2013; Aguillón-Gutiérrez & Ramírez-Bautista, 2015; Aguillón-Gutiérrez, 2018) which could lead to declines in recruitment to adult populations. Cd accumulation can lead to decrease in larval anuran survival, especially for species highly dependent on plant material (Gastelum et al., 2019), but another effect can be skin detachment (Severtsova et al., 2013; Aguillón-Gutiérrez & Ramírez-Bautista, 2015). Mn, Zn and Cu are essential micronutrients that at high concentrations can be highly toxic, as they accumulate in different organs in adult anurans and induce oxidative stress (via oxygen reactive species [ROS]). This in turn can alter the antioxidant defense system (AOS) (Londoño-Franco et al., 2016; Prokić et al., 2016; Stolyar et al., 2008) and ultimately cause genotoxic damage, which has been related to

detrimental effects at higher levels of biological organization (Mussali-Galante et al., 2014).

We found several metals accumulating in livers of anurans in the Huautla river, but the concentrations they attained are in general lower than those found in similar studies. For example, concentrations of 0.09 mg/kg for Cd, 2.78 mg/kg for Cr, 19.70 mg/kg for Zn and 46.40 mg/kg for Cu, have been reported in muscle and liver from anurans (Thanomsangad et al., 2019; Zhelev et al., 2020). A lower concentration in the Huautla versus other systems could result from slow but steady mine tailing leaching into the environment as mining activities ceased more than 30 years ago.

Anuran metal bioaccumulation was not consistent among species or sites. Given the location of sites relative to the position of mine tailings, we expected T1 and DW to have higher concentrations of all metals examined. However, some metals were found at higher concentrations in sites located at greater distance from mine tailings. Almost all metals analyzed, except for Cd, were found at higher concentrations in *L. spectabilis*. Cd was found at higher concentrations in *L. zweifeli* and Pb in *S. baudinii*. These three species have a close relationship to water bodies for reproduction, feeding and thermoregulation. They primarily feed upon aquatic invertebrates and are also cannibalistic. These could be the reasons for having elevated metal concentrations in liver versus other species. However, lack of consistency in results among species suggest multifactorial influences in metal bioaccumulation for anurans in the region. High variability in metal concentrations in anurans may stem from differential

tissue bioaccumulation rates for each metal, differential exposure and excretion rates at the individual or species level and generally differences in metal toxicokinetics and toxicodynamics (Thanomsangad et al., 2019).

Anuran Communities

In our study we found approximately 45% of all species reported for the REBIOSH (Herrera-Balcazar, 2014) and 75 % of all species previously reported for the study area (Roa-Mata, 2017). The highest richness, diversity and abundance of anurans was obtained from T1 and DW (sites also closest to existing mine tailings). Anuran species distribution and abundance in a landscape depend on a variety of factors influencing individuals, populations and communities (Álvarez-Grzybowska et al., 2020; Muths et al., 2020). Among these, anurans are highly susceptible to humidity conditions. Tropical dry forests are subject to seasonally dry conditions which can be stressful for anurans; many streams in such areas can dry up for extended periods. Under such conditions, we hypothesize that a richer community can be found in areas near mine tailings given increased water retention capacity. Areas with inactive mine tailings (i.e., areas where not new materials are being dumped) tend to have highly porous soils which can retain more water, leading to increased vegetation cover and organic matter (Ramos-Arroyo & Siebe-Grabach, 2006; Ryzhenko & Bokhonov, 2020). This in turn could provide increased humidity conditions for anurans and facilitate their burrowing-in during the dry season (Mora et al., 2012).

Anuran communities in the study area were dominated by species in the Hylidae family, which is common for neotropical regions (Both et al., 2008; Pansonato et al., 2011). Families Bufonidae and Ranidae had two species each and Microhylidae only one (Table 2). That semi-aquatic *L. zweifeli* and *L. spectabilis* were the most abundant species may follow their close association with riparian areas (Canseco-Márquez & Gutiérrez-Mayén, 2010) throughout the year, even during dry seasons (Chávez-Ramírez, 2017). *Rhinella horribilis*, a species with relatively high tolerance to desiccation given a thick and impermeable skin (Cruz-Piedrahita et al., 2018) was also abundant, especially in T1. Similar abundance was observed for *S. baudinii*, a species inhabiting trees but with aquatic larvae. The least abundant species were *Hypopachus variolosus* and *Agalychnis dacnicolor*. This last species, larger than other hylids in the region and sometimes considered a generalist, can often be found in areas with anthropogenic activity (Mendenhall et al., 2014; Soto-Sandoval et al., 2017).

While our study provides important information on community composition and potential threats from mining pollution in the REBIOSH, we acknowledge that our period of study was short, that we were not able to

explore communities during the dry season, that our study area and sample number for some species was small and that we assumed no anuran movement among sites. We studied the anuran community only during the wet season as this is when previous studies (Herrera-Balcazar, 2014; Roa-Mata, 2017) had identified the highest abundance and diversity for the group. Prior to this study we carried out pilot sampling trips during the dry season (when only isolated pools were present in the streams) and we were unable to capture any anurans (applying a similar sampling effort). Our study only encompassed three small streams in the REBIOSH; several other lotic systems in the REBIOSH exist which remain relatively unexplored (i.e., the riparian area of the Amacuzac River) and where the anuran communities require further study. However, our assumption that individuals cannot move among sites is generally supported by existing literature, as generally anurans do not move >1km away from their breeding site (Jeliakov et al., 2019; Lemckert 2004). For metal analyses in some sites we were only able to obtain and process one individual of a given species. We caution that metal concentrations from such samples, absent measures of variability, should be used simply as a reference to compare against other species but not considered representative of the entire species in the site. We set our study area to explore if metals from inactive mine tailings were present in anuran communities. Having found some pollutants in livers for anurans in the Huautla, we propose that similar studies are needed in the more than ten active or dormant mining areas in Morelos (SGM, 2020).

Much remains unknown about the biology, life history, population dynamics and community structure of anurans in southern central Mexico. Descriptive studies such as the one we present here are an initial but important step toward a better understanding of anuran biology and distribution. Our work joins several other documents from the area (Castro-Franco et al., 2006; Herrera-Balcazar, 2014; Roa-Mata, 2017) to aid in our understanding of the distribution of the various anuran species in this biologically diverse area of the country. While all species we encountered are regarded of least concern by IUCN (IUCN, 2023), we caution that other species or evolutionary significant units for some species could be facing greater threats from a variety of stressors than acknowledged in large-scale conservation assessments.

Implications for Conservation

Mining will continue to be an important economic activity worldwide (SE, 2020). It will also continue to be an important environmental problem as it contributes contaminants and dangerous residues (Xiao et al., 2020) and alters entire ecosystems (Ali & Khan, 2018; Chen et al., 2019). The effects from metal pollution will persist at all levels of biological organization (Mussali-Galante et al., 2013b) as they

move via food webs (Fuentes et al., 2020). While the heaviest mining activity in the State of Morelos has ceased for the moment (Télez-Ramírez & Sánchez-Salazar, 2018), its polluting legacy remains a threat for numerous socio-ecosystems in this area.

Anurans play an important role in riverine food webs and link various components in riparian ecotones (Hecnar and M'Closkey et al., 1996; Cortés-Gomez et al., 2015). Given their exposure to metals in the study area, they could serve as a route of exposure for fish, birds and mammals in the region (Ilizaliturri, 2010; Hernández-Plata et al., 2020). While our results did not show an impact of metals on anuran richness, abundance, or diversity among the sampled sites, we cannot discard possible effects at lower levels of organization. It is possible that future studies might find DNA adducts, oxidative damage or metallothionein induction at the molecular level (Hu et al., 2021). In addition, sex proportion and age structure alterations, low reproductive success, loss of genetic diversity, lowered fitness (Sasaki et al., 2016; Giroto et al., 2020), or population declines should be explored. These and higher level effects, such as alterations in energy and nutrient cycles and food webs at the ecosystem level need to also be addressed (Mussali-Galante et al., 2013a, 2014). Identifying these effects will require level-specific biomarkers able to detect biological changes that might be obscured by natural or other anthropogenic influences. Additionally, future studies shall consider phenotypic plasticity in anurans facing stressors such as metal exposure and include other potential target tissues (i.e., muscle, bone).

Despite the difficulty in carrying out above mentioned studies, they are key to conservation efforts in the REBIOSH. Here we have presented the first lines of evidence to understand the potential effects of metals to anurans in the area. It is upon building blocks like this study that better conservation strategies for protected areas can be achieved.

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Supplemental Material

Supplemental material for this article is available online.

References

- Aguillón-Gutiérrez, D. R. (2018). Anomalías macroscópicas en larvas de anfibios anuros. *Revista latinoamericana de herpetología*, 01(01), 8–21. <https://doi.org/10.22201/fc.25942158e.2018.1.12>
- Aguillón-Gutiérrez, D.R., & Ramírez-Bautista, A. (2015). Anomalías frecuentes en una población de *Hyla plicata* (Anura: Hylidae) expuesta a plomo y hierro durante el desarrollo postembrionario. *BIOCYT Biología Ciencia y Tecnología*, 8(29), 515–529. <https://doi.org/10.22201/fesi.20072082e.2015.8>
- Ali, H., & Khan, E. (2018). Trophic transfer, bioaccumulation, and biomagnification of non-essential hazardous heavy metals and metalloids in food chains/webs—Concepts and implications for wildlife and human health. *Human and Ecological Risk Assessment*, 25(6), 1353–1376. <https://doi.org/10.1080/10807039.2018.1469398>
- Álvarez-Grzybowska, E., Urbina-Cardona, J.N., Córdova-Tapia, F., & García, A. (2020). Amphibian communities in two contrasting ecosystems: functional diversity and environmental filters. *Biodiversity and Conservation*, 29(8), 2457–2485. <https://doi.org/10.1007/s10531-020-01984-w>
- Anderson, M. J. (2001). Permutation tests for univariate or multivariate analysis of variance and regression. *Canadian Journal of Fisheries and Aquatic Sciences*, 58(3), 626–639. <https://doi.org/10.1139/f01-004>
- Anderson, M., Gorley, R., & Clarke, K. (2008). “*PERMANOVA+ for PRIMER: Guide to software and statistical methods*,” in *PRIMER-E* (Plymouth: PRIMER-E).
- Barats, A., Renac, C., Orani, A. M., Durrieu, G., Martin, H. S., Esteller, M. V., & Hoyos, S. E. G. (2020). Tracing source and mobility of arsenic and trace elements in a hydrosystem impacted by past mining activities (Morelos state, Mexico). *Science of The Total Environment*, 712, 135565. <https://doi.org/10.1016/j.scitotenv.2019.135565>
- Barrios-Damián, M. (2006). *Hábitos alimentarios de Rana spectabilis Hillis y Frost, 1985 (Anura: Ranidae) en la reserva estatal Sierra de Montenegro, Morelos, México* [Bachelor of

- Science in Biology. Universidad Autónoma del Estado de Morelos].
- Both, C., Kaefer, I. L., Santos, T. G. D., & Cechin, S. (2008). An Austral anuran assemblage in the Neotropics: seasonal occurrence correlated with photoperiod. *Journal of Natural History*, 42(3-4), 205–222. <https://doi.org/10.1080/00222930701847923>
- Calderon, M. R., Almeida, C. A., González, P., & Jofré, M. B. (2019). Influence of water quality and habitat conditions on amphibian community metrics in rivers affected by urban activity. *Urban Ecosystems*, 22(4), 743–755. <https://doi.org/10.1007/s11252-019-00862-w>
- Canseco-Márquez, L., & Gutiérrez-Mayén, M. G. (2010). Anfibios y reptiles del Valle de Tehuacán-Cuicatlán. México. Comisión Nacional para el Conocimiento y Uso de la Biodiversidad, 302. México. <https://doi.org/10.1111/poms.12085>
- Casas-Andreu, G., Valenzuela-López, G., & Ramírez-Bautista, A. (1991). Como hacer una colección de Anfibios y Reptiles. Instituto de Biología, UNAM, Cuadernos (Vol. 10), México.
- Castro-Franco, R., García, G. G. V., Zagal, M. G. B., & Arizmendi, W. M. (2006). Diversidad y distribución de anfibios del estado de Morelos, México. *Acta zoológica mexicana*, 22(1), 103–117. <https://doi.org/10.21829/azm.2006.2211964>
- CEAGUA (Comisión Estatal del Agua). (2017). Estadísticas del Agua en el Estado de Morelos. *Comisión Nacional del Agua*. Comisión Estatal del Agua. Ciudad de México. P. 196.
- Chao, A., & Jost, L. (2012). Coverage-based rarefaction and extrapolation: standardizing samples by completeness rather than size. *Ecology*, 93(12), 2533–2547. <https://doi.org/10.1890/11-1952.1>
- Chao, A. (1984). Nonparametric estimation of the number of classes in a population. *Scandinavian Journal of statistics*, 11(4), 265–270.
- Chávez-Ramírez, E. A. (2017). Diversidad y ocupación de nicho espacial y temporal por anuros en cuatro tipos de vegetación en el ejido Real del Puente, Xochitepec, Morelos. [Bachelor of Science in Biology, Universidad Autónoma del Estado de Morelos. México].
- Chen, J., McIlroy, S. E., Archana, A., Baker, D. M., & Panagiotou, G. (2019). A pollution gradient contributes to the taxonomic, functional, and resistome diversity of microbial communities in marine sediments. *Microbiome*, 7(1), 1–12. <https://doi.org/10.1186/s40168-019-0714-6>
- Clarke, K. R., & Gorley, R. N. (2015). *Getting started with PRIMER v7*. 20. Ltd, Plymouth: Plymouth Marine Laboratory, UK.
- CONANP (Comisión Nacional de Áreas Naturales Protegidas). (2005). *Programa de conservación y manejo de la Reserva de la Biosfera Sierra de Huautla, México*. O. Dorado, B. Maldonado, D.M. Arias, V. Sorani, R. Ramirez, E. Leyva, & y D. Valenzuela (eds). México, D.F.
- Cortés-Gómez, A. M., Ruiz-Agudelo, C. A. R., Valencia-Aguilar, A., & Ladle, R. J. (2015). Ecological functions of neotropical amphibians and reptiles: A review. *Universitas Scientiarum*, 20(2), 229–245. <https://doi.org/10.11144/Javeriana.SC20-2.efna>
- Cortés-Suárez, J. E. (2017). Uso de microhábitat por parte del sapo gigante *Rhinella horribilis* en pastizales en el municipio de Villa de Leyva, Boyacá, Colombia. *Biodiversidad Neotropical*, 7(4), 253–257. <https://doi.org/10.18636/bioneotropical.v7i4.701>
- Crump, M. L., & Scott, N. Y. (2001). Relevantamientos por encuentros visuales, p. 80–93. In: *Medición y Monitoreo de la Diversidad Biológica. Métodos estandarizados para anfibios*. W. M. Heyer, A. Donnelly, R. W. McDiarmid, L. A. Hayek, & M. C. Foster (eds.). Editorial Universitaria de la Patagonia, Chubut, República Argentina.
- Cruz-Piedrahita, C., Navas, C. A., & Crawford, A. J. (2018). Life on the edge: A comparative study of ecophysiological adaptations of frogs to tropical semiarid environments. *Physiological and Biochemical Zoology*, 91(1), 740–756. <https://doi.org/10.1086/695705>
- Curi, L. M., Peltzer, P. M., Attademo, M. A., & Lajmanovich, R. C. (2021). Alterations in gonads and liver tissue in two neotropical anuran species commonly occurring in rice fields crops. *Water Air and Soil Pollution*, 232(5). <https://doi.org/10.1007/s11270-021-05164-6>
- De La Cruz-Guameros, N., Tovar-Sánchez, E., & Mussali-Galante, P. (2021). Assessing effects of chronic heavy metal exposure through a multibiomarker approach: the case of *Liomys irroratus* (Rodentia: Heteromyidae). *Environmental Science and Pollution Research*, 28(39), 55373–55387. <https://doi.org/10.1007/s11356-021-14855-w>
- De Lucca, G. S., Barros, F. A. P., Oliveira, J. V., Dal Magro, J., & Lucas, E. M. (2018). The role of environmental factors in the composition of anuran species in several ponds under the influence of coal mining in southern Brazil. *Wetlands Ecology and Management*, 26(3), 285–297. <https://doi.org/10.1007/s11273-017-9573-8>
- Egusquiza-Morínigo, G. M. (2016). Ecotoxicología en anfibios: un análisis cuantitativo. [Bachelor of Science, Universidad Federal de la Integración Latinoamericana, Foz de Iguazú, Paraná]. <https://dspace.unila.edu.br/123456789/669>
- EPA. (2019). *Exposure Assessment Tools by Routes*. United States Environmental Protection Agency. <https://www.epa.gov/expobox/exposure-assessment-tools-routes>. Accessed 29 August 2023.
- Esteller, M.V., Domínguez-Mariani, E., & Garrido, S.E. (2015). Groundwater pollution by arsenic and other toxic elements in an abandoned silver mine, Mexico. *Environmental Earth Sciences*. 74, 2893–2906 (2015). <https://doi.org/10.1007/s12665-015-4315-9>
- Ficken, K. L. G., & Byrne, P. G. (2013). Heavy metal pollution negatively correlates with anuran species richness and distribution in south-eastern Australia. *Austral Ecology*, 38(5), 523–533. <https://doi.org/10.1111/j.1442-9993.2012.02443.x>
- Flores-Villela, O., Mendoza-Quijano, F., González-Porter, G., et al. (1995). Recopilación de claves para la identificación de anfibios y reptiles de México. In *En Publicaciones especiales del museo de zoología* (Vol. 10). UNAM.

- Frost, D. R. (2021). Amphibian Species of the World: an Online Reference. Version 6.1. Electronic Database accessible at 2023-07-29; American Museum of Natural History, New York, USA. <https://doi.org/10.5531/db.vz.0001>
- Fuentes, I., Márquez-Ferrando, R., Pleguezuelos, J. M., Sanpera, C., & Santos, X. (2020). Long-term trace element assessment after a mine spill: Pollution persistence and bioaccumulation in the trophic web. *Environmental Pollution*, 267, 115406. <https://doi.org/10.1016/j.envpol.2020.115406>
- Gastelum, A., Aquino, A., & Aldama, L. (2019). Efecto del cloruro de cadmio durante el desarrollo larvario de la rana toro *Lithobates catesbeianus* (Shaw, 1802). *Acta Universitaria*, 29, 1–11. <https://doi.org/10.15174/au.2019.1902>
- Giroto, L., Espíndola, E. L. G., Gebara, R. C., & Freitas, J. S. (2020). Acute and chronic effects on tadpoles (*Lithobates catesbeianus*) exposed to mining tailings from the dam rupture in Mariana, MG (Brazil). *Water, Air, & Soil Pollution*, 231(7). <https://doi.org/10.1007/s11270-020-04691-y>
- Gotelli, N.J., & Colwell, R.K. (2011). Estimating species richness. In *Biological diversity frontiers in measurement and assessment* (eds A.E. Magurran, & B.J. McGill), pp. 39–54. Oxford University Press, Oxford, UK.
- Hecnar, S. J., & M'Closkey, R. T. (1996). Amphibian species richness and distribution in relation to pond water chemistry in south-western Ontario, Canada. *Freshwater Biology*, 36(1), 7–15. <https://doi.org/10.1046/j.1365-2427.1996.00054.x>
- Hernández, O. D. H., Castro-Herrera, F., & Melo, M.P. (2013). Bioacumulación de mercurio en larvas de anuros en la zona afectada por la minería de oro en el río Dagua, buenaventura, Valle del Cauca, Colombia. *Acta Biológica Colombiana*, 18(2), 341–348.
- Hernández-Ordóñez, O., Santos, B. A., Pyron, R. A., Arroyo-Rodríguez, V., Urbina-Cardona, J. N., Martínez-Ramos, M., Parra-Olea, G., & Reynoso, V. H. (2019). Species sorting and mass effect along forest succession: Evidence from taxonomic, functional, and phylogenetic diversity of amphibian communities. *Ecology and Evolution*, 9(9), 5206–5218. <https://doi.org/10.1002/ece3.5110>
- Hernández-Ordóñez, O., Urbina-Cardona, N., & Martínez-Ramos, M. (2015). Recovery of amphibian and reptile assemblages during old-field succession of tropical rain forests. *Biotropica*, 47(3), 377–388. <https://doi.org/10.1111/btp.12207>
- Hernández-Plata, I., Rodríguez, V. M., Tovar-Sánchez, E., Carrizalez, L., Villalobos, P., Mendoza-Trejo, M. S., & Mussali-Galante, P. (2020). Metal brain bioaccumulation and neurobehavioral effects on the wild rodent *Liomys irroratus* inhabiting mine tailing areas. *Environmental Science and Pollution Research*. 27(29), 36330–36349. <https://doi.org/10.1007/s11356-020-09451-3>
- Herrera-Balcázar, A. F. (2014). Herpetofauna de Cerro frío, Reserva De La Biosfera Sierra De Huautla, Morelos, México. [Bachelor of Science in Biology, Universidad Nacional Autónoma de México]. <https://ru.dgb.unam.mx/handle/20.500.14330/TES01000719286>
- Hopkins, W. A. (2007). Amphibians as models for studying environmental change. *ILAR Journal*, 48(3), 270–277. <https://doi.org/10.1093/ilar.48.3.270>
- Hsieh, T. C., Ma, K. H., & Chao, A. (2016). iNEXT: an R package for rarefaction and extrapolation of species diversity (Hill numbers). *Methods in Ecology and Evolution*, 7(12), 1451–1456. <https://doi.org/10.1111/2041-210X.12613>
- Hu, J., Wang, J., Lv, X., Yu, L., Li, J., Lan, S., & Yang, Y. (2021). In situ assessment of genetic and epigenetic alterations in frog *Rana plancyi* and *Rana limnocharis* inhabiting aquatic ecosystems associated with PB/ZN/CU mining. *Science of The Total Environment*, 779, 146139. <https://doi.org/10.1016/j.scitotenv.2021.146139>
- Ilizaliturri, C. A. (2010). *Aplicación de una metodología de evaluación de riesgo ecológico en el sistema de humedales de Coatzacoalcos*, Veracruz. [Doctor of Environmental Science, Universidad Autónoma de San Luis Potosí]. <https://repositorioinstitucional.uaslp.mx/xmlui/handle/i/3584>
- Islam, M. S., Proshad, R., & Ahmed, M. S. (2017). Ecological risk of heavy metals in sediment of an urban river in Bangladesh. *Human and Ecological Risk Assessment*, 24(3), 699–720. <https://doi.org/10.1080/10807039.2017.1397499>
- IUCN (International union for conservation of nature). (2023). The IUCN Red List of Threatened Species. <https://www.iucnredlist.org>. Downloaded on 29/07/23.
- Jeliazkov, A., Lorrillière, R., Besnard, A., Garnier, J., Silvestre, M., & Chiron, F. (2019). Cross-scale effects of structural and functional connectivity in pond networks on amphibian distribution in agricultural landscapes. *Freshwater Biology*, 64(5), 997–1014. <https://doi.org/10.1111/fwb.13281>
- Jofré, M. B., Antón, R. I., & Caviedes-Vidal, E. (2012). Lead and cadmium accumulation in anuran amphibians of a permanent water body in arid Midwestern Argentina. *Environmental Science and Pollution Research*, 19(7), 2889–2897. <https://doi.org/10.1007/s11356-012-0795-2>
- Jost, L. (2006). Entropy and diversity. *Oikos*, 113(2), 363–375. <https://doi.org/10.1111/j.2006.0030-1299.14714.x>
- Kabir, M. H., Islam, M. S., Hoq, M. E., Tusher, T. R., & Islam, M. S. (2020). Appraisal of heavy metal contamination in sediments of the Shitalakhya River in Bangladesh using pollution indices, geo-spatial and multivariate statistical analysis. *Arabian Journal of GeoSciences*, 13(21), 1135. <https://doi.org/10.1007/s12517-020-06072-5>
- Kiesecker, J. M., Blaustein, A. R., & Belden, L. K. (2001). Complex causes of amphibian population declines. *Nature*, 410(6829), 681–684. <https://doi.org/10.1038/35070552>
- Lee, J. C. (2000). Patterns of distribution of amphibians: A global perspective. *Copeia*, 2000(4), 1138–1140. [https://doi.org/10.1643/0045-8511\(2000\)000](https://doi.org/10.1643/0045-8511(2000)000)
- Legendre, P., & Gallagher, E. D. (2001). Ecologically meaningful transformations for ordination of species data. *Oecologia*, 129(2), 271–280. <https://doi.org/10.1007/s004420100716>
- Lemckert, F.L. (2004). Variations in anuran movements and habitat use: Implications for conservation. *Applied*

- Herpetology*, 1(3-4), 165-181. <https://doi.org/10.1163/157075403323012179>
- Londoño-Franco, L. F., Londoño Muñoz, P. T., & Muñoz García, F. G. (2016). Los riesgos de los metales pesados en la salud humana y animal. *Biotecnología en el Sector Agropecuario y Agroindustrial*, 14(2), 145. [https://doi.org/10.18684/BSAA\(14\)145-153](https://doi.org/10.18684/BSAA(14)145-153)
- Magurran, A. E. (1988). *Ecological Diversity and Its Measurement*. Springer eBooks. <https://doi.org/10.1007/978-94-015-7358-0>
- Márquez-Huitzil, R., Martínez-Garza, C., Hinojosa-Espinoza, S., & Sorani, V. (2022). Identificación de minas y jales en Huautla, Morelos, México. *Investigaciones Geográficas*, (109). <https://doi.org/10.14350/ig.60633>
- McDiarmid, W.R. (2001). Preparación de anfibios como especímenes de científicos. In: W. M. Heyer, A. Donnelly, R. W. McDiarmid, L. A. Hayek, & M. C. Foster (eds.), *Medición y Monitoreo de la Diversidad Biológica. Métodos estandarizados para anfibios* (pp. 279–289). Editorial Universitaria de la Patagonia, Chubut, República Argentina.
- Mendenhall, C. D., Frishkoff, L. O., Santos-Barrera, G., Pacheco, J., Mesfun, E., Quijano, F. M., Ehrlich, P. R., Ceballos, G., Daily, G. C., & Pringle, R. M. (2014). Countryside biogeography of Neotropical reptiles and amphibians. *Ecology*, 95(4), 856–870. <https://doi.org/10.1890/12-2017.1>
- Mendoza-Estrada, L. J., Lara-López, R., & Castro-Franco, R. (2008). Dieta de *Lithobates zweifeli* Hillis, Frost y Webb 1984 (Anura: Ranidae) en un río estacional del Centro de México. *Acta Zoológica Mexicana*, 24(1), 169–197. <https://doi.org/10.21829/azm.2008.241630>
- Miranda, L. S., Ayoko, G. A., Egodawatta, P., & Goonetilleke, A. (2022). Adsorption-desorption behavior of heavy metals in aquatic environments: Influence of sediment, water and metal ionic properties. *Journal of Hazardous Materials* 421(12674). <https://doi.org/10.1016/j.jhazmat.2021.126743>
- Mora, G. G., Arias, J. C., Reyes, A., Jiménez, A., Padilla, S. E., Gómez-Mestre, I., & Sasa, M. (2012). Fenología reproductiva de anuros en humedales del Bosque Tropical Seco de Costa Rica. *Ciencias Ambientales*, 43(1), 29. <https://doi.org/10.15359/rca.43-1.3>
- Moreno, C. E. (2001). *Métodos para medir la biodiversidad. M&T-Manuales y Tesis SEA*, 1, 84.
- Moreno, C. E., Barragán, F., Pineda, E., & Pavón, N. P. (2011). Reanálisis de la diversidad alfa: Alternativas para interpretar y comparar información sobre comunidades ecológicas. *Revista Mexicana de Biodiversidad*, 82(4), 1249–1261. <https://doi.org/10.22201/ib.20078706e.2011.4.745>
- Muro-González, D. A., Mussali-Galante, P., Valencia-Cuevas, L., Flores-Trujillo, K., & Tovar-Sánchez, E. (2020). Morphological, physiological, and genotoxic effects of heavy metal bioaccumulation in *Prosopis laevigata* reveal its potential for phytoremediation. *Environmental Science and Pollution Research*, 27(32), 40187–40204. <https://doi.org/10.1007/s11356-020-10026-5>
- Mussali-Galante, P., Tovar-Sánchez, E., Valverde, M., & Del Castillo, E. R. (2013a). Biomarkers of exposure for assessing environmental metal pollution: from molecules to ecosystems. *Revista Internacional De Contaminación Ambiental*, 29(1), 117-140.
- Mussali-Galante, P., Tovar-Sánchez, E., Valverde, M., & Rojas, E. (2014). Genetic Structure and Diversity of Animal Populations Exposed to Metal Pollution. In: D. Whitacre (eds), *Reviews of Environmental Contamination and Toxicology*, vol 227, pp. 70-106. Springer, Cham. https://doi.org/10.1007/978-3-319-01327-5_3
- Mussali-Galante, P., Tovar-Sánchez, E., Valverde, M., Valencia-Cuevas, L., & Rojas, E. (2013b). Evidence of population genetic effects in *Peromyscus melanophrys* chronically exposed to mine tailings in Morelos, Mexico. *Environmental Science and Pollution Research*, 20(11), 7666–7679. <https://doi.org/10.1007/s11356-012-1263-8>
- Muths, E., Hossack, B. R., Campbell Grant, E. H., Pilliod, D. S., & Mosher, B. A. (2020). Effects of snowpack, temperature, and disease on demography in a wild population of amphibians. *Herpetologica*, 76(2), 132–143. <https://doi.org/10.1655/0018-0831-76.2.132>
- NMX. (1980). Norma Mexicana NMX-AA-014-1980. Lineamientos para el muestreo en cuerpos receptores de aguas superficiales. Secretaría de Comercio y Fomento Industrial, Estados Unidos Mexicanos. <https://faolex.fao.org/docs/pdf/mex51857.pdf>
- NMX. (2001). NMX-AA-051-SCFI-2001. Análisis de agua - determinación de metales por absorción atómica en aguas naturales, potables, residuales y residuales tratadas - Método de prueba. Secretaría de Economía, Estados Unidos Mexicanos. https://www.dof.gob.mx/normasOficiales/6280/seeco3a12_C/seeco3a12_C.html
- NMX. (2006). Norma Mexicana NMX-AA-132-SCFI-2006. Muestreo de suelos para la identificación y la cuantificación de metales y metaloides, y manejo de la muestra. Secretaría de Economía, Estados Unidos Mexicanos. https://www.dof.gob.mx/nota_detalle.php?codigo=5475373&fecha=06/03/2017#gsc.tab=0
- NMX (2016). Norma Mexicana NMX-AA-051-SCFI-2016. Análisis de agua-Medición de metales por absorción atómica en aguas naturales, potables, residuales y residuales tratadas-Método de prueba. Secretaría de Economía, Estados Unidos Mexicanos. https://www.dof.gob.mx/normasOficiales/6280/seeco3a12_C/seeco3a12_C.html
- NMX. (2017). Norma Mexicana NMX-AA-112-SCFI-2017. Análisis de agua y sedimentos-evaluación de toxicidad aguda con *Vibrio fischeri*. Método de prueba. *Secretaría de Economía, Estados Unidos Mexicanos*. https://www.dof.gob.mx/nota_detalle.php?codigo=5501793&fecha=19/10/2017#gsc.tab=0
- Ortiz, J. M. (2019). Plan municipal de desarrollo de Tlaquiltenango, Morelos. H. *Ayuntamiento de Tlaquiltenango Morelos*. <https://www.tlaquiltenango.gob.mx/wp-content/uploads/2019/11/Plan-Municipal-Tlaquiltenango-2019-2021.pdf>. Accessed 28 September 2021.
- Pandey, L., Park, J., Son, D. H., Kim, W., Islam, M. S., Choi, S., Lee, H., & Han, T. (2019). Assessment of metal

- contamination in water and sediments from major rivers in South Korea from 2008 to 2015. *Science of The Total Environment*, 651, 323-333. <https://doi.org/10.1016/j.scitotenv.2018.09.057>
- Pansonato, A., Mott, T., & Strüssmann, C. (2011). Anuran amphibians diversity in a northwestern area of the Brazilian pantanal. *Biota Neotropica*, 11(4), 77-86. <https://doi.org/10.1590/s1676-06032011000400008>
- Penicaud, V., Lacoue-Labarthe, T., & Bustamante, P. (2017). Metal bioaccumulation and detoxification processes in cephalopods: A review. *Environmental Research*, 155, 123-133. <https://doi.org/10.1016/j.envres.2017.02.003>
- Prokić, M. D., Borković-Mitić, S. S., Krizmanić, I. I., Mutić, J. J., Trifković, J., Gavrić, J. P., Despotović, S. G., Gavrilović, B. R., Radovanović, T. B., Pavlović, S. Z., & Saičić, Z. S. (2016). Bioaccumulation and effects of metals on oxidative stress and neurotoxicity parameters in the frogs from the *Pelophylax esculentus* complex. *Ecotoxicology*, 25(8), 1531-1542. <https://doi.org/10.1007/s10646-016-1707-x>
- Proshad, R., Ahmed, S., Rahman, M., & Kumar, T. (2017). Apportionment of hazardous elements in agricultural soils around the vicinity of brick kiln in Bangladesh. *Journal of Environmental and Analytical Toxicology*, 07(02). <https://doi.org/10.4172/2161-0525.1000439>
- Proshad, R., Islam, S., Tusher, T. R., Zhang, D., Khadka, S., Gao, J., & Kundu, S. (2020). Appraisal of heavy metal toxicity in surface water with human health risk by a novel approach: a study on an urban river in vicinity to industrial areas of Bangladesh. *Toxin Reviews*, 40(4), 803-819. <https://doi.org/10.1080/15569543.2020.1780615>
- Proshad, R., Kormoker, T., & Islam, M. S. (2019). Distribution, source identification, ecological and health risks of heavy metals in surface sediments of the Rupsa River, Bangladesh. *Toxin Reviews*, 40(1), 77-101. <https://doi.org/10.1080/15569543.2018.1564143>
- Ramírez-Bautista, A., Torres-Hernández, L. A., Cruz-Elizalde, R., Berriozabal-Islas, C., Cruz-Elizalde, R., Wilson, L. D., Johnson, J. D., Porras, L. W., Balderas-Valdivia, C. J., González-Hernández, A., & Mata-Silva, V. (2023). An updated list of the Mexican herpetofauna: with a summary of historical and contemporary studies. *ZooKeys*, 1166, 287-306. <https://doi.org/10.3897/zookeys.1166.86986>
- Ramos-Arroyo, Y. R., & Siebe-Grabach, C. D. (2006). Estrategia para identificar jales con potencial de riesgo ambiental en un distrito minero: Estudio de caso en el Distrito de Guanajuato, México. *Revista Mexicana de Ciencias Geológicas*, 23(1), 54-74.
- Rinklebe, J., Antoniadis, V., Shaheen, S. M., Rosche, O., & Altermann, M. (2019). Health risk assessment of potentially toxic elements in soils along the Central Elbe River, Germany. *Environment International*, 126, 76-88. <https://doi.org/10.1016/j.envint.2019.02.011>
- Roa-Mata, M. F. (2017). Análisis de la comunidad de anfibios y su relación con la variabilidad ambiental en la presa Lorenzo Vázquez, Tlaquiltenango, Morelos, México. [Bachelor of Science in Biology. Universidad Autónoma del Estado de Morelos].
- Rodríguez-Rodríguez, R. M. (2020). Elementos potencialmente tóxicos en el polvo urbano de la zona metropolitana de San Luis Potosí. [Master of Applied Geoscience, Instituto Potosino de Investigación Científica y Tecnológica, A.C. México]. <https://repositorio.ipicyt.edu.mx/handle/11627/5279>
- Ruiz de Velasco, A. (1890). Geografía y Estadística del Estado de Morelos. Oficina Tipográfica de la Secretaría de Fomento. México. 7, 13-17.
- Ryzhenko, N., & Bokhonov, Y. (2020). Trace metals migration from soil to seepage water. *Polish Journal of Soil Science*, 53(2), 261-271. <https://doi.org/10.17951/pjss/2020.53.2.261>
- Santos-Barrera, A. G., Hammerson, G., Wilson, L. D., Bolaños, F., & Ponce-Campos, P. (2010). *Smilisca baudinii*, Common Mexican Treefrog. Red List, IUCN, 8235.
- Santoyo-Martínez, M., Mussali-Galante, P., Hernández-Plata, I., Valencia-Cuevas, L., Flores-Morales, A., Ortiz-Hernández, L., & Tovar-Sánchez, E. (2020). Heavy metal bioaccumulation and morphological changes in *Vachellia campechiana* (Fabaceae) reveal its potential for phytoextraction of Cr, Cu, and Pb in mine tailings. *Environmental Science and Pollution Research*, 27(10), 11260-11276. <https://doi.org/10.1007/s11356-020-07730-7>
- Sasaki, K., Lesbarrères, D., Beaulieu, C., Watson, G. D., & Litzgus, J. D. (2016). Effects of a mining-altered environment on individual fitness of amphibians and reptiles. *Ecosphere*, 7(6). <https://doi.org/10.1002/ecs2.1366>
- Scott, N. J., & Woodward, B. D. (2001). Relevamientos de lugares de reproducción. In: W. M. Heyer, A. Donnelly, R. W. McDiarmid, L. A. Hayek, & M. C. Foster (eds.), *Medición y Monitoreo de la diversidad biológica. Métodos estandarizados para anfibios* (pp. 113-120). Editorial Universitaria de la Patagonia, Chubut, República Argentina.
- SE (Secretaría de Economía). (2020). *Minería*. <https://www.gob.mx/se/acciones-y-programas/mineria.Fechadeconsulta:21-julio-2023>
- SEMARNAT (Secretaría del Medio Ambiente y Recursos Naturales). 2010. *Norma Oficial Mexicana Nom-059-Semarnat-2010, Protección ambiental-Especies nativas de México de flora y fauna silvestres-Categorías de riesgo y especificaciones para su inclusión, exclusión o cambio-Lista de especies en riesgo*. Diario Oficial de la Federación, 30 de diciembre de 2010, Segunda Sección, México.
- Severtsova, E. A., & Aguillón-Gutiérrez, D. R. (2013). Postembryonic development of anurans in ponds littered with metal-containing refuse (simulation experiments). *Biology bulletin of the Russian Academy of Sciences*, 40(9), 738-747. <https://doi.org/10.1134/S1062359013090094>
- Severtsova, E. A., Nikiforova, A. I., & Aguillón-Gutiérrez, D. R. (2013). Spectrochemical and histochemical analyses of tissues of grass frog and gray toad tadpoles developing under simulation of pollution by plumbum and ferrum. *Moscow University Biological Sciences Bulletin*, 68(4), 186-191. <https://doi.org/10.3103/S009639251304010X>

- SGM (Sistema Geológico Mexicano). (2020). Anuario Estadístico de la Minería Mexicana. Servicio Geológico Mexicano. M. T. A. López, & H.D.N. Betancourt (eds.). México, D.F. Vol. 49, 555.
- Soliman, M. M., Hesselberg, T., Mohamed, A. A., & Renault, D. (2022). Trophic transfer of heavy metals along a pollution gradient in a terrestrial agro-industrial food web. *Geoderma*, 413, 115748. <https://doi.org/10.1016/j.geoderma.2022.115748>
- Solís, M. B. M. (2016). *Aislamiento de bacterias de jales mineros y análisis de uso potencial para la remediación de sitios contaminados con metales pesados* [Master of Biotechnology, Universidad Autónoma del Estado de Morelos, México].
- Soto-Sandoval, Y., Suazo-Ortuño, I., Urbina-Cardona, N., Marroquín-Páramo, J., & Alvarado-Díaz, J. (2017). Effect of tropical dry forest sucesional stages on microhabitat use by *Agalychnis dacnicolor* (Anura: Phyllomedusidae) and *Smilisca fodiens* (Anura: Hylidae). *Revista de Biología Tropical* 65 (2). <https://doi.org/10.15517/rbt.v65i2.24706>
- Sparling, D. W. (2017). *Basics of Ecotoxicology*. Taylor and Francis Group (eds.). Boca Ratón, Miami, Florida, USA. CRC Press. <https://doi.org/10.1201/9781315158068>
- Stolyar, O. B., Loumbourdis, N. S., Falfushinska, H. I., & Romanchuk, L. D. (2008). Comparison of metal bioavailability in frogs from urban and rural sites of Western Ukraine. *Archives of Environmental Contamination and Toxicology*, 54(1), 107–113. <https://doi.org/10.1007/s00244-007-9012-6>
- Suárez, V. R., Mercado-Silva, N., García, C. P., & Iñiguez Dávalos, L. I. (2016). Anurans of the Marabasco River, New Records for the State of Colima, Mexico. *Southwestern Naturalist*, 61(3), 241–245. <https://doi.org/10.1894/0038-4909-61.3.241>
- Téllez-Ramírez, I., & Sánchez-Salazar, M. T. (2018). La expansión territorial de la minería mexicana durante el periodo 2000-2017. Una lectura desde el caso del Estado de Morelos. *Investigaciones Geográficas, Boletín del Instituto de Geografía*. <https://doi.org/10.14350/riig.59607>
- Thanomsangad, P., Tengjaroenkul, B., Sriuttha, M., & Neeratanaphan, L. (2019). Heavy metal accumulation in frogs surrounding an e-waste dump site and human health risk assessment. *Human and Ecological Risk Assessment*, 26(5), 1313-1328. <https://doi.org/10.1080/10807039.2019.1575181>
- Timofeev, I., Kosheleva, N., & Kasimov, N. (2018). Contamination of soils by potentially toxic elements in the impact zone of tungsten-molybdenum ore mine in the Baikal region: A survey and risk assessment. *Science of the Total Environment*, 642, 63-76. <https://doi.org/10.1016/j.scitotenv.2018.06.042>
- Tovar-Sánchez, E., Galante, P. M., Pacheco, M., Hernández, M. H., Salinas, E. S., & Velona, A. O. (2016). Relationship between genotoxic damage and arsenic blood concentrations in individuals residing in an arsenic contaminated area in Morelos, Mexico. *Revista Internacional de Contaminación Ambiental*, 32(1), 101-117.
- Varol, M., Canpolat, Ö., Eriş, K. K., & Çağlar, M. (2020). Trace metals in core sediments from a deep lake in eastern Turkey: Vertical concentration profiles, ecoenvironmental risks and possible sources. *Ecotoxicology and Environmental Safety*, 189, 110060. <https://doi.org/10.1016/j.ecoenv.2019.110060>
- Venne, L. S., Cobb, G. P., Coimbatore, G., Smith, L. M., & McMurry, S. T. (2006). Influence of land use on metal concentrations in playa sediments and amphibians in the Southern High Plains. *Environmental Pollution*, 144(1), 112–118. <https://doi.org/10.1016/j.envpol.2006.01.003>
- Volke, S. T., & Velasco, J. A. (2005). Tecnologías de remediación para suelos contaminados. *Número 2*. Instituto de Ecología. México, D.F. 3 (2) pp. 60. <https://repositorio.unan.edu.ni/2986/1/5624.pdf>
- Whittaker, R. H. (1965). Dominance and diversity in land plant communities. *Science*, 147(3655), 250–260. <https://doi.org/10.1126/science.147.3655.250>
- Xiao, L., Yu, Z., Liu, H., Tan, T., Yao, J., Zhang, Y., & Wu, J. (2020). Effects of Cd and Pb on diversity of microbial community and enzyme activity in soil. *Ecotoxicology*, 29(5), 551-558. <https://doi.org/10.1007/s10646-020-02205-4>
- Zhelev, Z. M., Arnaudova, D. N., Popgeorgiev, G. S., & Tsonev, S. V. (2020). In situ assessment of health status and heavy metal bioaccumulation of adult *Pelophylax ridibundus* (Anura: Ranidae) individuals inhabiting polluted area in southern Bulgaria. *Ecological Indicators*, 115, 106413. <https://doi.org/10.1016/j.ecolind.2020.106413>
- Zhou, Q., Zhang, J., Fu, J., Shi, J., & Jiang, G. (2008). Bio-monitoring: An appealing tool for assessment of metal pollution in the aquatic ecosystem. *Analytica Chimica Acta*, 606(2), 135–150. <https://doi.org/10.1016/j.aca.2007.11.018>