

Repellency and Bioactivity of Caatinga Biome Plant Powders against *Callosobruchus maculatus* (Coleoptera: Chrysomelidae: Bruchinae)

Authors: Melo, Bruno Adelino de, Molina-Rugama, Adrián José, Haddi, Khalid, Leite, Delzuite Teles, and Oliveira, Eugênio Eduardo de

Source: Florida Entomologist, 98(2) : 417-423

Published By: Florida Entomological Society

URL: <https://doi.org/10.1653/024.098.0204>

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at www.bioone.org/terms-of-use.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

Repellency and bioactivity of Caatinga biome plant powders against *Callosobruchus maculatus* (Coleoptera: Chrysomelidae: Bruchinae)

Bruno Adelino de Melo¹, Adrián José Molina-Rugama^{1,2,*}, Khalid Haddi³, Delzuite Teles Leite¹, and Eugênio Eduardo de Oliveira^{3,*}

Abstract

The Caatinga biome represents the 4th-largest area covered by single vegetation in Brazil and contains dry forests rich in aromatic bushes, vines, herbs, and trees. The flora of this ecological region is widely known and employed in folk medicine and has other utilitarian and economic uses; however, its potential for controlling or repelling insects is poorly investigated. In this study, we evaluated the potential use of Caatinga plant species for controlling infestations of *Callosobruchus maculatus* F. (Coleoptera: Chrysomelidae: Bruchinae), the most important insect pest of cowpea, *Vigna unguiculata* (L.) Walp. (Fabales: Fabaceae). Powders of the leaves and stems of 9 plant species, including *Amburana cearensis* A. C. Smith ("cumaru") (Fabales: Fabaceae), *Croton sonderianus* Müll. Arg. ("marmeleiro") (Malpighiales: Euphorbiaceae), *Cleome spinosa* Jacq. ("mussambê") (Capparales: Cleomaceae), *Mimosa tenuiflora* Benth. ("jurema-preta") (Fabales: Fabaceae), *Anadenanthera macrocarpa* (Benth.) Brenan ("angico-vermelho") (Fabales: Fabaceae), *Aspidosperma pyrifolium* Mart. ("pereiro") (Gentianales: Apocynaceae), *Senna occidentalis* (L.) H.S. Irwin & R.C. Barneby ("manjerioba") (Fabales: Fabaceae), *Hyptis suaveolens* (L.) Poit. ("alfazema-brava") (Lamiales: Lamiaceae), and *Ziziphus joazeiro* Mart. ("juazeiro") (Rosales: Rhamnaceae), were applied on masses of cowpea seeds, and their effects on *C. maculatus* longevity as well as their repellent activities were evaluated. All the leaf and stem powders reduced only the longevity of males and showed strongly repellent activities against females. The preference level of females for untreated beans varied between 73 and 94%, indicating that all the leaf and stem powders can be a part of the integrated management of *C. maculatus* in storage facilities.

Key Words: stored grain pest; bruchid; *Vigna unguiculata*; plant powder; alternative pest control

Resumo

O bioma Caatinga representa a quarta maior área coberta por um único tipo de vegetação no Brasil. Este bioma se constitui de florestas secas com considerável diversidade de arbustos, ervas, trepadeiras e árvores aromáticas. A flora desta região ecológica é amplamente conhecida e tem sido utilizada para diversos fins utilitários e econômicos, principalmente na medicina popular. No entanto, o potencial destas plantas para controlar ou repelir insetos ainda é pouco investigado. Neste estudo, foi avaliado o uso potencial de espécies de plantas da Caatinga para controlar infestações de *Callosobruchus maculatus* F. (Coleoptera: Chrysomelidae: Bruchinae), uma das mais importantes pragas no feijão caupi, *Vigna unguiculata* (L.) Walp. (Fabales: Fabaceae). Pó das folhas e de caules de nove espécies de plantas, incluindo *Amburana cearensis* A. C. Smith ("cumaru") (Fabales: Fabaceae), *Croton sonderianus* Müll. Arg. ("marmeleiro") (Euphorbiaceae), *Spinosa cleome* Jacq. ("Mussambê") (Capparales: Cleomaceae), *Mimosa tenuiflora* Benth. ("jurema-preta") (Fabales: Fabaceae), *Anadenanthera macrocarpa* (Benth.) Brenan ("angico-vermelho") (Fabales: Fabaceae), *Aspidosperma pyrifolium* Mart. ("pereiro") (Gentianales: Apocynaceae), *Senna occidentalis* (L.) H.S. Irwin & R.C. Barneby ("mangirioba") (Fabales: Fabaceae), *Hyptis suaveolens* (L.) Poit. ("alfazema-brava") (Lamiales: Lamiaceae) e *Ziziphus joazeiro* Mart. ("juazeiro") (Rosales: Rhamnaceae), foram aplicados em massas de feijão caupi e seus efeitos sobre a longevidade e repelência de *C. maculatus* foram avaliados. Todos os pós de folhas e de caules reduziram apenas a longevidade de machos de *C. maculatus*. Entretanto, estes mesmos pós mostraram alta atividade repelente contra fêmeas destes insetos. O nível de preferência de fêmeas de *C. maculatus* para grãos não tratados variou entre 73 e 94%, indicando que os pós das folhas e de caules destas plantas podem se constituir importantes ferramentas para o manejo integrado de *C. maculatus* em unidades de armazenamento.

Palavras Chave: pragas de grãos armazenados; bruquídeo; *Vigna unguiculata*; pós vegetais; controle alternativo de pragas

The Caatinga biome accounts for about 60% of the northeast Brazilian territory and extends to a small part of the northeastern Minas Gerais State (Sampaio et al. 2002). This area is mainly covered by xeric

shrub lands rich in aromatic bushes, vines, herbs, and trees (Almeida et al. 2005) with its native plants presenting utilitarian and economic potential (Albuquerque & Andrade 2002; Lucena et al. 2007, 2008; Ca-

¹Unidade Acadêmica de Ciências Agrárias, Universidade Federal de Campina Grande, Pombal, Paraíba 58840-000, Brazil

²Current address: Departamento de Ciências Vegetais, Universidade Federal Rural do Semi-Árido, Mossoró, Rio Grande do Norte 59625-900, Brazil

³Departamento de Entomologia, Universidade Federal de Viçosa, Viçosa, Minas Gerais 36570-900, Brazil

*Corresponding authors; E-mail: eugenio@ufv.br, amolina@ufersa.edu.br

nuto et al. 2012). Many of the Caatinga plant species are used by native communities as firewood (Ramos et al. 2008), in carpentry (owing to their recognized durability), as seasoning (Canuto et al. 2012), or in folk medicine to treat several diseases (Leal et al. 2000; Albuquerque et al. 2007; Alviano et al. 2008; Cartaxo et al. 2010; Canuto et al. 2012). The great diversity of the Caatinga vegetation is underexploited, and few searches for active biological substances, including those with insecticidal or repellent activity, have been conducted (Almeida et al. 2005; Albuquerque et al. 2007).

Food availability in the Brazilian Caatinga heavily depends on the capacity of farmers (most of them are subsistence producers) to preserve the post-harvest quality of their production. In this region, cereals and beans are grown predominantly by small farmers with little or no technological inputs (Vieira 2004; Ferreira et al. 2013). These farmers have low family income, and they usually keep their production inside their own small storage facilities with high quantitative and qualitative losses, most of them due to insect damage. Natural products from locally available plants with insecticide activity represent a low-cost and sustainable alternative to protect agricultural production. Furthermore, botanical insecticides supposedly pose little threat to the environment or human health compared with synthetic insecticides, and they represent a suitable alternative to controlling mites and insect pests worldwide (Isman 2006; Regnaut-Roger et al. 2012; Kedia et al. 2013).

The cowpea weevil, *Callosobruchus maculatus* F. (Coleoptera: Chrysomelidae: Bruchinae), damages 20–30% of legume seeds in the tropical countries (Kirado & Srivastava 2010) and can cause up to 100% loss when masses of cowpea beans are untreated (Gbaye et al. 2011). Adults mate after emergence and typically live not more than 2 wk depending on ambient temperature. The females deposit eggs on the surface of maturing cowpea pods and seeds. The newly emerged larvae burrow into and feed on a single seed until pupation, and adults do not need to feed (Mitchell 1975; Southgate 1978). Several holes are left in the seed by the emerging adults with severe weight loss facilitating fungal and mycotoxin contamination, which reduces the commercial bean value (Kirado & Srivastava 2010; Kedia et al. 2013).

Insecticidal natural products, such as powders of locally available plants, used by farmers in developing countries in their storage facilities, appear to be safe and promising (Paul et al. 2009; Silva et al. 2013; Tavares et al. 2013, 2014; Fouad et al. 2014; Melo et al. 2014). Thus, we evaluated the repellent activity and the effects of powders from 9 Caatinga plant species on *C. maculatus* longevity.

Material and Methods

INSECT REARING

The original population of *C. maculatus* was field-collected from small farms in the region of Pombal (Paraíba State, Brazil) and established under laboratory conditions ($25 \pm 2^\circ\text{C}$, $70 \pm 5\%$ RH, and 12:12 h L:D photoperiod), starting with at least 500 individuals. The identification was based on the traits described previously (Athié & Paula 2002). The population was reared on cowpea bean (*Vigna unguiculata* [L.] Walp.; Fabales: Fabaceae) grains (free of insecticides) bought from the local market. In order to avoid possible infestations from the field and to reduce any potential insecticide residual effect, the bean grains were kept at a temperature of -10°C for 14 d prior to being offered to *C. maculatus*. To obtain newly emerged *C. maculatus* of the same generation, adult insects were released in cowpea bean grain masses that were placed in plastic containers (0.4 L capacity) covered with “organza” cloth. After 5 d of colonization, the adults were removed and the egg-infested grains were maintained under laboratory conditions. The new adults emerged after around 4 wk.

PLANT POWDERS

The plant powders used in this study were obtained from the leaves and stems of 9 Caatinga plant species, including *Amburana cearensis* A. C. Smith (“cumuru-nordestino”) (Fabales: Fabaceae), *Croton sonderianus* Müll.Arg. (“marmeleiro-do-mato”) (Malpighiales: Euphorbiaceae), *Cleome spinosa* Jacq. (“mussambê”) (Capparales: Cleomaceae), *Mimosa tenuiflora* Benth. (“jurema-preta”) (Fabales: Fabaceae), *Anadenanthera macrocarpa* (Benth.) Brenan (“angico-vermelho”) (Fabales: Fabaceae), *Aspidosperma pyrifolium* Mart. (“pereiro”) (Gentianales: Apocynaceae), *Senna occidentalis* (L.) H.S. Irwin & R.C. Barneby (“mangirioba”) (Fabales: Fabaceae), *Hyptis suaveolens* (L.) Poit. (“alfazema-brava”) (Lamiales: Lamiaceae), and *Ziziphus joazeiro* Mart. (“juazeiro”) (Rosales: Rhamnaceae) (Table 1), collected in the region of Pombal (Paraíba State, Brazil). We chose only plant species that are used by native communities to treat several diseases, and some of their biological activities have been described (Table 1). During the period between the years of 2009 and 2012, leaves and stems were randomly collected from the adult plants by using pruning scissors. Samples of these plants were compared with material deposited in the herbarium of the Universidade Federal Rural do Semi-Árido (UFERSA, Mossoró-RN, Brazil). All the plant materials were individually wrapped in plastic bags, identified, and brought to the laboratory. Then, these materials were dried by direct exposure to sunlight over a 7 d period, and leaves and stem were separately milled with a manual grinder to powder. The resulting powder was passed through a 25 mesh sieve to obtain a fine dust. The fine dusts were stored individually in glass containers (hermetically closed) that were maintained at a controlled temperature (5°C) to ensure supply of the material throughout the investigation period.

LONGEVITY BIOASSAYS

The effects of each plant powder on insect longevity were assessed in survival bioassays conducted according to previously described methods (Procópio et al. 2003). Briefly, a pair of newly emerged weevils was confined in a plastic container (100 mL) containing 45 g of untreated (control) or plant powder treated cowpea bean seeds. Each weevil pair in 45 g of bean seeds was an experimental unit. In the treated bean unit, 2 g of the plant powder had been homogeneously distributed among the seeds. Five replicates were used for each plant powder tested, and the male and female insect mortality was monitored daily until the last day of survival. As these insects are excellent fliers, we customized an escape-proof cage that allowed measurements of mortality. This cage had the following dimensions: 40 cm length \times 20 cm width \times 20 cm height, and its base, back, and front sides were made of wood. Openings of 10 cm diameter were drilled in the back and front sides and were closed with organza cloth. These openings facilitated the insertion and handling of experimental materials. Furthermore, complete and easy viewing and handling of the experimental materials were achieved through the glass used at the top and lateral sides of the cage. The bean seeds were carefully poured onto the plastic trays placed inside the cage. After counting the number of dead insects, all the live insects, bean grains, and plant powders were added back into the experimental units. The insect longevity measurements were subjected to analysis of variance and subsequently to Tukey’s test ($\alpha = 0.05$), when appropriate.

FREE-CHOICE REPELLENCE TEST

The repellent activity of each plant powder was assessed in bioassays conducted in custom-made plastic arenas (35 cm diameter, 12 cm high), according to the modified protocols reported previously (Burkholder & Dicke 1966; Phillips & Burkholder 1981). Six 50 mL plastic

Table 1. Caatinga plant species collected in the county of Pombal, Paraíba State, Brazil.

Scientific name	Family	Common name	Biological activity	Isolated compounds	References
<i>Amburana cearensis</i> A. C. Smith	Fabaceae	"cumaru"	anticholinesterase, antinociceptive, antiinflammatory, bronchodilator, antibacterial	courmarin, phenolic glycosides	(Bravo et al. 1999; Leal et al. 2000; Trevisan & Macedo 2003; Figueredo et al. 2013)
<i>Croton sanderianus</i> Müll. Arg.	Euphorbiaceae	"marmeleiro"	anticholinesterase, antimicrobial, antifungal	clerodane and cleistanthane type diterpenes; 1,8-cineol	(McChesney et al. 1991; Santos et al. 2005; Fontenelle et al. 2008)
<i>Cleome spinosa</i> Jacq.	Cleomaceae	"musambê"	neuroprotection, cytotoxicity against a number of human cancer cell lines, anti-HIV activity	unsaturated polyphenols, cembranoids, diterpenes	(Faulkner 2001; Hanson 2002; Collins et al. 2004)
<i>Mimosa tenuiflora</i> Willd. Benth.	Fabaceae	"jurema-preta"	anti-inflammatory, antioxidant, antifungal, healing properties	tannins and flavonoids	(Araújo et al. 2008)
<i>Anadenanthera macrocarpa</i> (Benth.) Brenan	Fabaceae	"angico-vermelho"	antibacterial, anti-inflammatory	phenols, flavonoids, free xanthenes, leucoantho-cyanidins	(Desmarchelier et al. 1999; Figueredo et al. 2013)
<i>Aspidosperma pyrifolium</i> Mart.	Apocynaceae	"pereiro"	antiplasmodial	monoterpenoid indole alkaloids	(Araújo et al. 2007)
<i>Senna occidentalis</i> (L.) H.S. Irwin & R.C. Barneby	Fabaceae	"mangirioba"	anti-inflammatory, antioxidant, antifungal, healing properties	tannins and flavonoids	(Araújo et al. 2008)
<i>Hyptis suaveolens</i> (L.) Poit.	Lamiaceae	"alfazema-brava"	anti-inflammatory, antioxidant, antifungal, healing properties, anticonvulsant	tannins and flavonoids, β -Caryophyllene, 1,8-cineole	(Akah & Nwambie 1993; Peertzada 1997; Araújo et al. 2008)
<i>Ziziphus joazeiro</i> Mart.	Rhamnaceae	"juazeiro"	antimicrobial, antioxidant, anticholinesterase	saponins	(Alviano et al. 2008; Farias et al. 2013; Ribeiro et al. 2013)

containers were placed at equidistance inside the arena, with 30 g of cowpea bean seeds in each container. Plant powder to be tested (1.5 g per container) was added to alternate containers (3 per arena). To facilitate odor removal, a 5 cm diameter hole was drilled in the center of the arena's lid for the insertion of a 5 cm diameter polyvinyl chloride (PVC) tube 10 cm in height. The external extremity of this tube was covered with organza cloth to prevent escape of the insects. Thirty adult females (aged 1–5 d) were released into the center of the arena, and after 24 h, the total number of insects per container was registered. Five replicates were used for each plant powder tested. In preliminary tests, we found even distribution of insects among containers when all the 6 plastic containers were filled only with untreated cowpea, so there was no indication of a position effect within the arena.

A binomial test ($P < 0.01$) was used to evaluate the significance of differences between the percentages of females that moved to untreated and powder treated bean seeds. The percentage of repellency was calculated as proposed by Mazzonetto & Vendramin (2003): $RI = (2 \times T) \div (T + C) \times 100$, where RI = repellency index, C = number of insects in the untreated container, and T = number of insects in the treated container. The RI values ranged between 0 and 2, which denoted the following: $RI = 1$, neutral activity; $RI > 1$, attraction; and $RI < 1$, repellency. As a safety margin for this classification, the standard deviation (SD) of each treatment was added/subtracted from the value of 1 (indicative of neutrality). The repellency index results were subjected to analysis of variance, and the averages were compared by using the Scott–Knott groupment analysis test (Scott & Knott 1974) at a probability level of 0.05.

Results

LONGEVITY BIOASSAYS

There were no significant differences ($F_{8,76} = 1.99$; $P > 0.05$) among the longevity of females exposed to leaf or stem powders of each plant tested, which allowed us to pool these longevity data and compare them with the longevity of females on untreated bean masses (Fig. 1). In general, the average longevity of females treated with plant powders was 7.4 ± 1.01 d and did not differ significantly ($F_{1,76} = 0.86$; $P > 0.05$) from that of the control females (7.8 ± 1.09 d; Fig. 1A). Likewise, the males showed similar longevity ($F_{8,76} = 0.82$; $P > 0.05$) when exposed to leaf or stem powders of each plant tested. However, the average longevity of males was significantly reduced ($F_{1,76} = 8.15$; $P < 0.01$) from 7.8 ± 1.79 d (control males) to 6.06 ± 1.25 d (males that lived on plant powder-treated beans) (Fig. 1B).

REPELLENT ACTIVITIES

All of the Caatinga plant powders were strongly repellent to females. The percentages of the females that preferred untreated beans ranged from 77% to 94% and were significantly greater ($P < 0.01$, binomial test) than those of females that preferred the leaf powder-treated beans (Fig. 2A). Similar results were obtained in the repellency bioassays with stem powders, where females significantly preferred ($P < 0.01$, binomial test) untreated bean seeds (Fig. 2B).

Although all the plant powders significantly repelled females, the leaf powders of *A. pyrifolium*, *S. occidentalis*, *H. suaveolens*, and *Z. joazeiro* exhibited greater repellency levels (Table 2). With regard to the stem powders, the plant species *C. sonderianus*, *C. spinosa*, *H. suaveolens*, and *Z. joazeiro* presented greater repellency levels. Furthermore, the leaf and stem powders of *A. pyrifolium* and *S. occidentalis* showed differential repellent activities (Table 2), with the leaf powders presenting greater repellency levels.

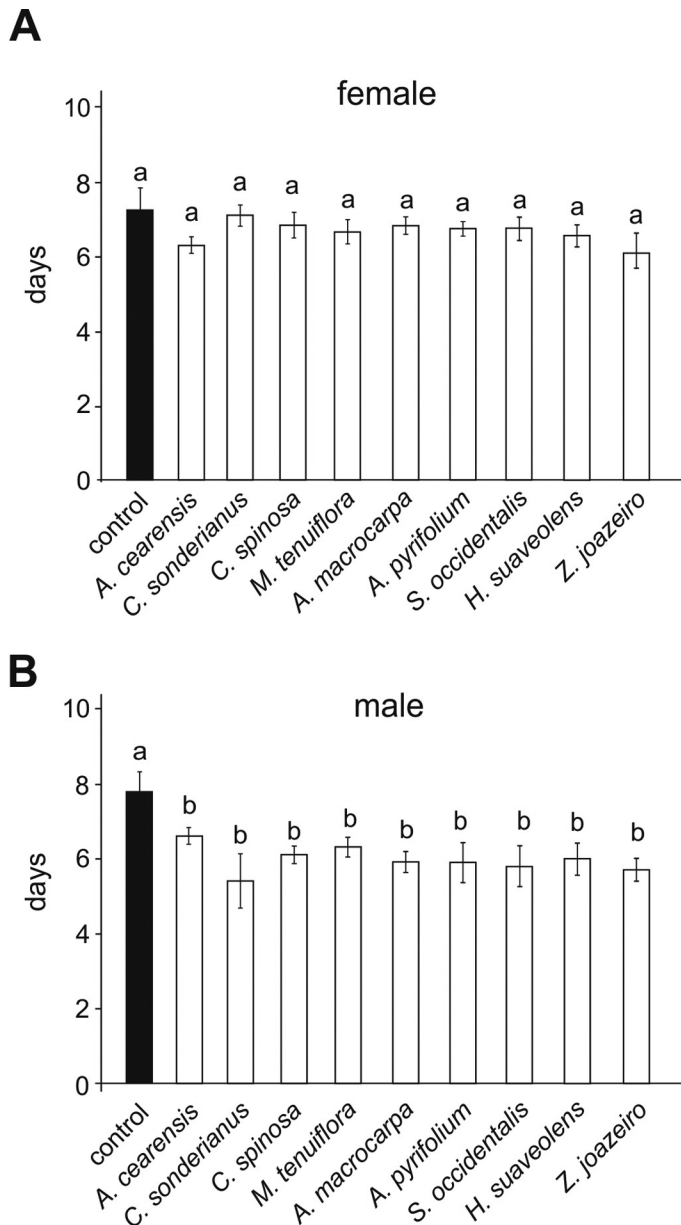


Fig. 1. Average longevity of *Callosobruchus maculatus* (Coleoptera: Chrysomelidae) females (A) and males (B) in the presence of the powders of 9 Caatinga plant species. Bars with the same letter indicate that no significant differences were noted among *C. maculatus* by Tukey's test ($\alpha = 0.05$). The following plant species were tested: *Amburana cearensis* ("cumarú"), *Croton sonderianus* ("marmeleiro"), *Cleome spinosa* ("mussambê"), *Mimosa tenuiflora* ("jurema-preta"), *Anadenanthera macrocarpa* ("angico-vermelho"), *Aspidosperma pyrifolium* ("pereiro"), *Senna occidentalis* ("manjerioba"), *Hyptis suaveolens* ("alfazema-brava"), and *Ziziphus joazeiro* ("juazeiro").

Discussion

Despite its great territorial expanse and significant biodiversity, the Caatinga biome is still an underexploited source of molecules with insecticidal/repellent activities. Most studies with plant products from this ecological region have focused on extracts or essential oils to control insect disease vectors (Lima et al. 2006; Farias et al. 2010; Souza et al. 2011; Santos et al. 2012; Barbosa et al. 2014). Few studies investigated

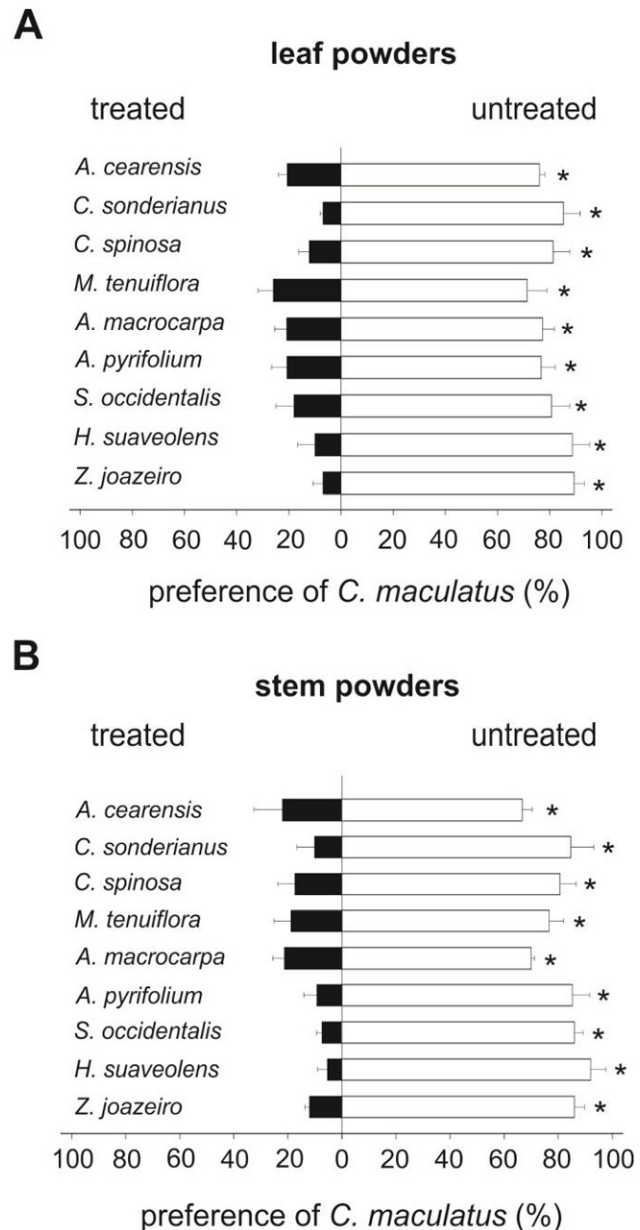


Fig. 2. Percentage of *Callosobruchus maculatus* (Coleoptera: Chrysomelidae) that moved toward bean seeds untreated and treated with leaf (A) and stem (B) powders. An asterisk by a bar indicates a significant difference in repellency between leaf powder-treated and untreated bean seeds (binomial test, $P < 0.01$). The following plant species were tested: *Amburana cearensis* ("cumarú"), *Croton sonderianus* ("marmeleiro"), *Cleome spinosa* ("mussambê"), *Mimosa tenuiflora* ("jurema-preta"), *Anadenanthera macrocarpa* ("angico-vermelho"), *Aspidosperma pyrifolium* ("pereiro"), *Senna occidentalis* ("manjerioba"), *Hyptis suaveolens* ("alfazema-brava"), and *Ziziphus joazeiro* ("juazeiro").

the potential of Caatinga plant powders as commodity protectants, and they normally evaluated only mortality effects (Souza & Tróvão 2009; Cruz et al. 2013). Here, we evaluated the insecticidal and repellent activities of 9 Caatinga plant species (*A. cearensis*, *C. sonderianus*, *C. spinosa*, *M. tenuiflora*, *A. macrocarpa*, *A. pyrifolium*, *S. occidentalis*, *H. suaveolens*, and *Z. joazeiro*) against the cowpea weevil, *C. maculatus*. Leaf and stem powders from these plants had major insecticidal effects on males and repelled the females, demonstrating their potential for use in the integrated management of *C. maculatus* in storage facilities.

Table 2. The repellency index (RI) obtained for each plant powder tested against *Callosobruchus maculatus* (Coleoptera: Chrysomelidae).

Plant species	Repellency index ^a	
	Leaves	Stems
<i>Amburana cearensis</i>	0.46 ± 0.05 Aa	0.42 ± 0.04 Aa
<i>Croton sonderianus</i>	0.22 ± 0.10 Ba	0.15 ± 0.03 Ba
<i>Cleome spinosa</i>	0.35 ± 0.08 Aa	0.26 ± 0.06 Ba
<i>Mimosa tenuiflora</i>	0.39 ± 0.07 Aa	0.54 ± 0.07 Aa
<i>Anadenanthera macrocarpa</i>	0.46 ± 0.05 Aa	0.42 ± 0.06 Aa
<i>Aspidosperma pyrifolium</i>	0.20 ± 0.06 Bb	0.43 ± 0.07 Aa
<i>Senna occidentalis</i>	0.16 ± 0.03 Bb	0.36 ± 0.08 Aa
<i>Hyptis suaveolens</i>	0.11 ± 0.04 Ba	0.20 ± 0.07 Ba
<i>Ziziphus joazeiro</i>	0.25 ± 0.02 Ba	0.14 ± 0.04 Ba

^aMeans followed by the same lowercase letter in a row or the same capital letter in a column are not significantly different based on the Scott–Knott groupment analysis test at $P < 0.05$.

Similar to the lack of insecticide activity against *C. maculatus* females observed here for all the plant powders, root powder of *M. tenuiflora* showed very small insecticidal activity against termites (Isoptera) (Cruz et al. 2013), and powders of *A. macrocarpa* did not show any insecticidal activity against the maize weevil, *Sitophilus zeamais* Motschulsky (Coleoptera: Curculionidae) (Souza & Tróvão 2009). Furthermore, powders from other medicinal plants (thyme, *Thymus vulgaris* L. [Lamiales: Lamiaceae]; lavender cotton, *Santolina chamaecyparissus* L., and stinking bean trefoil, *Anagyris foetida* L. [Fabales: Fabaceae]) neither affected the longevity of southern cowpea weevil, *Callosobruchus chinensis* L. (Coleoptera: Chrysomelidae) males nor females (Righi-Assia et al. 2010). These differential insecticidal activities of plant powders might have resulted from multiple factors involving the way they work and the resistance mechanisms of the insects. Plant powders can control insects by eroding the cuticle layer and causing dehydration (Kedia et al. 2013); blocking the spiracles and causing asphyxiation (Denloye 2010); or impairing physiological processes by penetrating the insect body via the respiratory or alimentary system (Ofuya & Dawodu 2002). Plant powders of *S. occidentalis* caused significant mortality in *C. maculatus* (Adesina et al. 2011), and insecticidal properties of *A. pyrifolium* (Torres et al. 2006), *C. sonderianus* (Moraes et al. 2006; Lima et al. 2006, 2013), *A. cearensis* (Farias et al. 2010; Souza et al. 2011), and *Z. joazeiro* (Souza et al. 2011) have been documented in different insect species. The repellent activities of Caatinga plant powders need further study although the repellency of many other plant powders against stored pests has been reported (Elhag 2000; Kéita et al. 2001; Mazzonetto & Vendramin 2003; Silva-Aguayo et al. 2005; Sanon et al. 2006; Kabir & Muhammad 2010).

The present study extends knowledge on Caatinga plants for use as stored product protectants because it demonstrates that the leaf and stem powders of 9 Caatinga plants show strong repellent activities against *C. maculatus* females. Leaf powders of *A. pyrifolium* and *S. occidentalis* repelled *C. maculatus* more efficiently than the stem powders of these plants, suggesting that these plants possess different active constituents or that they have the same constituents but with different concentrations in various plant parts (Ravi Kiran et al. 2006; Autran et al. 2009). Such differential activities of the powders of leaves and stems of other plants such as neem, *Azadirachta indica* A. Juss. (Sapindales: Meliaceae) (leaf and stem bark powders), have been described, with the leaf powder showing higher repellent activities against *C. maculatus* than the stem powder (Kabir & Muhammad 2010). The striking repellency results obtained here for *M. tenuiflora* and *A. macrocarpa* powders are noteworthy, because these plant products had been previously reported to have no (Souza & Tróvão 2009; Santos et al. 2012)

or very low insecticidal activity (Cruz et al. 2013). We also found that *S. occidentalis* strongly repels *C. maculatus* females, which differs from the results described by Pålsson & Jaenson (1999), who observed no repellent activities of this plant against mosquitoes (Diptera: Culicidae), reinforcing the hypothesis that repellent activity of plant products might be species specific.

Furthermore, *H. suaveolens* plant products demonstrated noticeable repellent activity against *C. maculatus* females, as demonstrated with other insect species (Sanon et al. 2006; Ilboudo et al. 2010; Benelli et al. 2012). However, products from this plant species can cause detrimental effects on natural enemies in storage environments (Sanon et al. 2011), thus requiring caution when used as grain protectants. In addition, other plant species of the *Cleome* genus showed repellent actions against ticks (Parasitiformes) and insects (Ndungu et al. 1995; Nyalala & Grout 2007), but the present study is the first to report on the insecticidal/repellent potential against *C. spinosa*.

The application of plant materials with insecticidal or repellent properties to stored grains is a common traditional method in rural areas around the world (Regnault-Roger et al. 2012; Kedia et al. 2013). Tropical ecosystems (such as the Caatinga biome) are particularly rich in plants that are used by local communities to treat diseases, thus indicating the potential to discover new compounds (Albuquerque et al. 2007, 2008). Further investigations exploring the toxicological aspects of the major constituents or identifying the principal volatiles produced by the Caatinga plants tested here will provide new insights on how these plants exhibit their insecticidal/repellent activities.

Our findings not only extend the knowledge on the Caatinga plants but also provide information about plants that can be used to protect cowpeas against *C. maculatus* infestations. All the plants tested are readily available in the Caatinga Region, and these anti-insect materials are affordable to low-income farmers who are normally constrained to sell their production early after harvest or, even worse, have their stored bean seeds (normally saved on the farm from the previous harvest) prone to infestation by stored product pests.

Acknowledgments

This work was supported by grants of the “Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq)” and “Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES)” to all authors, and by grants of “Fundação Arthur Bernardes (FUNARBE)” and “Fundação de Amparo à Pesquisa do Estado de Minas Gerais (FAPEMIG)” to EEO.

References Cited

- Adesina JM, Afolabi LA, Aderibigbe ATB. 2011. Efficacy of *Senna occidentalis* leaves powder on oviposition, hatchability of eggs and emergence of *Callosobruchus maculatus* (Fab) on treated cowpea seeds. *South Asian Journal of Experimental Biology* 1: 168-171.
- Akah PA, Nwambie AI. 1993. Nigerian plants with anti-convulsant properties. *Fitoterapia* 64: 42-44.
- Albuquerque UP, Andrade LHC. 2002. Uso de recursos vegetais da caatinga: o caso do agreste do estado de Pernambuco (Nordeste do Brasil). *Interciência* 27: 336-346.
- Albuquerque UP, Medeiros PM, Almeida ALS, Monteiro JM, Freitas Lins Neto EM, Melo JG, Santos JP. 2007. Medicinal plants of the Caatinga (semi-arid) vegetation of NE Brazil: a quantitative approach. *Journal of Ethnopharmacology* 114: 325-354.
- Albuquerque UP, Silva VA, Cabral MC, Alencar NL, Andrade LHC. 2008. Comparisons between the use of medicinal plants in indigenous and rural Caatinga (dryland) communities in NE Brazil. *Boletín de la Sociedad Latinoamericana y del Caribe de Plantas Medicinales y Aromáticas* 7: 156-170.
- Almeida CFCBR, Lima e Silva TC, de Amorim ELC, Maia MBDS, Albuquerque UP. 2005. Life strategy and chemical composition as predictors of the selection

- of medicinal plants from the Caatinga (Northeast Brazil). *Journal of Arid Environments* 62: 127-142.
- Alviano WS, Alviano DS, Diniz CG, Antonioli AR, Alviano CS, Farias LM, Carvalho MAR, Souza MMG, Bolognese AM. 2008. In vitro antioxidant potential of medicinal plant extracts and their activities against oral bacteria based on Brazilian folk medicine. *Archives of Oral Biology* 53: 545-552.
- Araújo Jr JX, Antheaume C, Trindade RCP, Schmitt M, Bourguignon J-J, Sant'Ana AEG. 2007. Isolation and characterisation of the monoterpenoid indole alkaloids of *Aspidosperma pyrifolium*. *Phytochemistry Reviews* 6: 183-188.
- Araújo TAS, Alencar NL, Amorim ELC, Albuquerque UP. 2008. A new approach to study medicinal plants with tannins and flavonoids contents from the local knowledge. *Journal of Ethnopharmacology* 120: 72-80.
- Athié I, Paula D.C. 2002. Insetos de grãos armazenados: aspectos biológicos e identificação, 2nd edition. Varela, São Paulo, Brazil. 244 pp.
- Autran ES, Neves IA, da Silva CSB, Santos GKN, Câmara CAGD, Navarro DMAF. 2009. Chemical composition, oviposition deterrent and larvicidal activities against *Aedes aegypti* of essential oils from *Piper marginatum* Jacq. (Piperaceae). *Bioresource Technology* 100: 2284-2288.
- Barbosa PBBM, de Oliveira JM, Chagas JM, Rabelo LMA, de Medeiros GF, Giodani, RB, da Silva EA, Uchôa AF, Ximenes MDDM. 2014. Evaluation of seed extracts from plants found in the Caatinga biome for the control of *Aedes aegypti*. *Parasitology Research* 113: 3565-3580.
- Benelli G, Flamini G, Canale A, Molfetta I, Cioni PL, Conti B. 2012. Repellence of *Hyptis suaveolens* whole essential oil and major constituents against adults of the granary weevil *Sitophilus granarius*. *Bulletin of Insectology* 65: 177-183.
- Bravo JAB, Sauvain M, Gimenez TA, Muñoz OV, Callapa J, Le Men-Olivier L, Masiot G, Lavaud C. 1999. Bioactive phenolic glycosides from *Amburana cearensis*. *Phytochemistry* 50: 71-74.
- Burkholder WE, Dicke RJ. 1966. Evidence of sex pheromones in females of several species of Dermestidae. *Journal of Economic Entomology* 59: 540-543.
- Canuto KM, Silveira ER, Bezerra AME, Leal LKAM, Viana GSB. 2012. Phytochemistry, pharmacology and agronomy of medicinal plants: *Amburana cearensis*, an interdisciplinary study, pp. 353-374 In Rao V [ed.], *Phytochemicals – A Global Perspective of their Role in Nutrition and Health*. InTech, Rijeka, Croatia.
- Cartaxo SL, De Almeida Souza MM, Albuquerque UP. 2010. Medicinal plants with bioprospecting potential used in semi-arid Northeastern Brazil. *Journal of Ethnopharmacology* 131: 326-342.
- Collins DO, Reynolds WF, Reese PB. 2004. New cembranes from *Cleome spinosa*. *Journal of Natural Products* 67: 179-183.
- Cruz CSA, Medeiros MB, Gomes JP, Souza FC. 2013. Uso de plantas em pó seco com propriedades termiticida sobre a mortalidade de cupins arbóreos. *Revista Verde de Agroecologia e Desenvolvimento Sustentável* 7: 1-5.
- Denloye AA. 2010. Bioactivity of powder and extracts from garlic, *Allium sativum* L. (Alliaceae) and spring onion, *Allium fistulosum* L. (Alliaceae) against *Callosobruchus maculatus* F. (Coleoptera: Bruchidae) on cowpea, *Vigna unguiculata* (L.) Walp (Leguminosae) seeds. *Psyche* 2010: article ID 958348.
- Desmarchelier C, Lisboa Romão R, Coussio J, Ciccio G. 1999. Antioxidant and free radical scavenging activities in extracts from medicinal trees used in the 'Caatinga' region in northeastern Brazil. *Journal of Ethnopharmacology* 67: 69-77.
- Elhag EA. 2000. Deterrent effects of some botanical products on oviposition of the cowpea bruchid *Callosobruchus maculatus* (F.) (Coleoptera: Bruchidae). *International Journal of Pest Management* 46: 109-113.
- Farias DF, Cavaleiro MG, Viana MP, Queiroz VA, Rocha-Bezerra LCB, Vasconcelos IK, Morais SM, Carvalho AFU. 2010. Water extracts of Brazilian leguminous seeds as rich sources of larvicidal compounds against *Aedes aegypti* L. *Anais da Academia Brasileira de Ciências* 82: 585-594.
- Farias DF, Souza TM, Viana MP, Soares BM, Cunha AP, Vasconcelos IM, Ricardo NMPS, Ferreira PMP, Melo VMM, Carvalho AFU. 2013. Antibacterial, antioxidant, and anticholinesterase activities of plant seed extracts from Brazilian semiarid region. *BioMed Research International* 510736, 9 pp.
- Faulkner DJ. 2001. Marine natural products. *Natural Product Reports* 18: 1-49.
- Ferreira LVM, Nóbrega RSA, Nóbrega JCA, Aguiar FL, Moreira FMS, Pacheco LP. 2013. Biological nitrogen fixation in production of *Vigna unguiculata* (L.) Walp, family farming in Piauí, Brazilian *Journal of Agricultural Science* 5: 153-160.
- Figueredo FG, Ferreira EO, Lucena BFF, Torres CMG, Lucetti DL, Lucetti ECP, Silva JMFL, Santos FAV, Medeiros CR, Oliveira GMM, Colares AV, Costa JGM, Coutinho HDM, Menezes IRA, Silva JCF, Kerntopf MR, Figueiredo PRL, Matias EFF. 2013. Modulation of the antibiotic activity by extracts from *Amburana cearensis* A. C. Smith and *Anadenanthera macrocarpa* (Benth.) Brenan. *BioMed Research International* 640682, 5 pp.
- Fontenelle ROS, Morais SM, Brito EHS, Brilhante RSN, Cordeiro RA, Nascimento NRF, Kerntopf MR, Sidrim JJC, Rocha MFG. 2008. Antifungal activity of essential oils of *Croton* species from the Brazilian *Caatinga* biome. *Journal of Applied Microbiology* 104: 1383-1390.
- Fouad HA, Faroni LRD, Tavares, WD, Ribeiro RC, Freitas SD, Zanuncio JC. 2014. Botanical extracts of plants from the Brazilian cerrado for the integrated management of *Sitotroga cerealella* (Lepidoptera: Gelechiidae) in stored grains. *Journal of Stored Product Research* 57: 6-11.
- Gbaye OA, Millard JC, Holloway GJ. 2011. Legume type and temperature effects on the toxicity of insecticide to the genus *Callosobruchus* (Coleoptera: Bruchidae). *Journal of Stored Product Research* 47: 8-12.
- Hanson JR. 2002. Diterpenoids. *Natural Product Reports* 19: 125-132.
- Ilboudo Z, Dabiré LCB, Nébrié RCH, Dicko IO, Dugravot S, Cortesero AM, Sanon A. 2010. Biological activity and persistence of four essential oils towards the main pest of stored cowpeas, *Callosobruchus maculatus* (F.) (Coleoptera: Bruchidae). *Journal of Stored Product Research* 46: 124-128.
- Isman OA. 2006. Botanical insecticides, deterrents and repellents in modern agriculture and an increasingly regulated world. *Annual Review of Entomology* 51: 45-66.
- Kabir HY, Muhammad S. 2010. Comparative studies of seed oil extract, leaves and stem bark powders of *Azadirachta indica* Linn (Meliaceae) on adults *Callosobruchus maculatus* (Coleoptera Bruchidae). *Bioscience Research* 22: 345-350.
- Kedia A, Prakash B, Mishra PK, Singh P, Dubey NK. 2013. Botanicals as eco friendly biorational alternatives of synthetic pesticides against *Callosobruchus* spp. (Coleoptera: Bruchidae) – a review. *Journal of Food Science and Technology* 51: 2210-2215.
- Kéita SM, Vincent C, Schmit J-P, Arnason JT, Bélanger A. 2001. Efficacy of essential oil of *Ocimum basilicum* L. and *O. gratissimum* L. applied as an insecticidal fumigant and powder to control *Callosobruchus maculatus* (Fab.) (Coleoptera: Bruchidae). *Journal of Stored Product Research* 37: 339-349.
- Kirado MM, Srivastava M. 2010. A comparative study on the efficacy of two Lamiaceae plants on egg – laying performance by the pulse beetle *Callosobruchus chinensis* Linn. (Coleoptera: Bruchidae). *Journal of Biopesticides* 3: 590-595.
- Leal LKAM, Ferreira AAG, Bezerra GA, Matos FJA, Viana GSB. 2000. Antinociceptive, anti-inflammatory and bronchodilator activities of Brazilian medicinal plants containing coumarin: a comparative study. *Journal of Ethnopharmacology* 70: 151-159.
- Lima JKA, Albuquerque ELD, Santos ACC, Oliveira AP, Araújo APA, Blank AF, Arrigoni-Blank MDF, Alves PB, Santos DDA, Bacci L. 2013. Biototoxicity of some plant essential oils against the termite *Nasutitermes corniger* (Isoptera: Termitidae). *Industrial Crop Production* 47: 246-251.
- Lima MGA, Maia ICC, Sousa BD, Morais SM, Freitas SM. 2006. Effect of stalk and leaf extracts from Euphorbiaceae species on *Aedes aegypti* (Diptera, Culicidae) larvae. *Revista do Instituto de Medicina Tropical de São Paulo* 48: 211-214.
- Lucena RFP, Albuquerque UP, Monteiro JM, De Fátima C, Almeida CBR, Florentino ATN, Ferraz JSF. 2007. Useful plants of the semi-arid Northeastern region of Brazil – a look at their conservation and sustainable use. *Environmental Monitoring and Assessment* 125: 281-290.
- Lucena RFP, Nascimento VT, Araújo EL, Albuquerque UP. 2008. Local uses of native plants in an area of Caatinga vegetation (Pernambuco, NE Brazil). *Ethnobotany Research and Applications* 6: 3-13.
- Mazonetto F, Vendramin JD. 2003. Efeito de pós de origem vegetal sobre *Acanthoscelides obtectus* (Say) (Coleoptera: Bruchidae) em feijão armazenado. *Neotropical Entomology* 32: 145-149.
- McChesney JD, Clark AM, Silveira ER. 1991. Antimicrobial diterpenes of *Croton sonderianus*, 1. Hardwick and 3,4-secotrachylobanoic acids. *Journal of Natural Products* 54: 1625-1633.
- Melo BA, Molina-Rugama AJ, Leite, DT, de Godoy MS, de Araújo EL. 2014. Bioatividade de pós de espécies vegetais sobre a reprodução de *Callosobruchus maculatus* (Fabr. 1775) (Coleoptera: Bruchidae). *Bioscience Journal* 30: 346-353.
- Mitchell R. 1975. Evolution of oviposition tactics in bean weevil, *Callosobruchus maculatus* (F.). *Ecology* 56: 696-702.
- Morais SM, Cavalcanti ESB, Bertini LM, Oliveira CLL, Rodrigues JRB, Cardoso JHL. 2006. Larvicidal activity of essential oils from Brazilian croton species against *Aedes aegypti* L. *Journal of the American Mosquito Control Association* 22: 161-164.
- Ndungu M, Lwande W, Hassanali A, Moreka L, Chhabra SC. 1995. *Cleome monophylla* essential oil and its constituents as tick (*Rhipicephalus appendiculatus*) and maize weevil (*Sitophilus zeamais*) repellents. *Entomologia Experimentalis et Applicata* 76: 217-222.
- Nyalala, S, Grout B. 2007. African spider flower (*Cleome gynandra* L./*Gynandropsis gynandra* (L.) Briq.) as a red spider mite (*Tetranychus urticae* Koch) repellent in cut-flower rose (*Rosa hybrida* L.) cultivation. *Scientia Horticulturae* 114: 194-198.

- Ofuya TI, Dawodu EO. 2002. Aspects of insecticidal action of *Piper guineense* Schum and Thonn fruit powders against *Callosobruchus maculatus* (F.) (Coleoptera: Bruchidae). *Niger Journal of Entomology* 19: 40-50.
- Pålsson K, Jaenson TGT. 1999. Plant products used as mosquito repellents in Guinea Bissau, West Africa. *Acta Tropica* 72: 39-52.
- Paul UV, Lossini JS, Edwards PJ, Hilbeck A. 2009. Effectiveness of products from four locally grown plants for the management of *Acanthoscelides obtectus* (Say) and *Zabrotes subfasciatus* (Boheman) (both Coleoptera: Bruchidae) in stored beans under laboratory and farm conditions in northern Tanzania. *Journal of Stored Product Research* 45: 97-107.
- Peerzada N. 1997. Chemical composition of the essential oil of *Hyptis suaveolens*. *Molecules* 2: 165-168.
- Phillips JK, Burkholder WE. 1981. Evidence for a male-produced aggregation pheromone in the rice weevil (Coleoptera, Curculionidae). *Journal of Economic Entomology* 74: 539-542.
- Procópio SO, Vendramin JD, Ribeiro Júnior JI, Santos J.B. 2003. Bioatividade de diversos pós de origem vegetal em relação a *Sitophilus zeamais* Mots. (Coleoptera: Curculionidae). *Ciencia e Agrotecnologia* 27: 1231-1236.
- Ramos MA, Medeiros PM, Almeida ALS, Feliciano ALP, Albuquerque UP. 2008. Use and knowledge of fuelwood in an area of Caatinga vegetation in NE Brazil. *Biomass Bioenergy* 32: 510-517.
- Ravi Kiran SR, Bhavani K, Devi PS, Rao BRR, Reddy KJ. 2006. Composition and larvicidal activity of leaves and stem essential oils of *Chloroxylon swietenia* DC against *Aedes aegypti* and *Anopheles stephensi*. *Bioresource Technology* 97: 2481-2484.
- Regnault-Roger C, Vincent C, Arnason JT. 2012. Essential oils in insect control: low-risk products in a high-stakes world. *Annual Review of Entomology* 57: 405-424.
- Ribeiro BD, Alviano DS, Barreto DW, Coelho MAZ. 2013. Functional properties of saponins from sisal (*Agave sisalana*) and juá (*Ziziphus joazeiro*): critical micellar concentration, antioxidant and antimicrobial activities. *Colloids and Surfaces A* 436: 736-743.
- Righi-Assia AF, Khelil MA, Medjdoub-Bensaad F, Righi K. 2010. Efficacy of oils and powders of some medicinal plants in biological control of the pea weevil (*Callosobruchus chinensis* L.). *African Journal of Agricultural Research* 5: 1474-1481.
- Sampaio EVSB, Giuiietti AM, Viginio J, Gamarra-Rojas CFL. 2002. Vegetação e flora da Caatinga, 1st edition. Associação Plantas do Nordeste, Recife, PE, Brazil, p. 176.
- Sanon A, Ilboudo Z, Dabiré CLB, Nebie RCH, Dicko IO, Monge JP. 2006. Effects of *Hyptis spicigera* Lam. (Labiatae) on the behaviour and development of *Callosobruchus maculatus* F. (Coleoptera: Bruchidae), a pest of stored cowpeas. *International Journal of Pest Management* 52: 117-123.
- Sanon A, Ba MN, Dabiré LCB, Nébié RCH, Monge JP. 2011. Side effects of grain protectants on biological control agents: how *Hyptis* plant extracts affect parasitism and larval development of *Dinarmus basalis*. *Phytoparasitica* 39: 215-222.
- Santos EA, Carvalho CM, Costa ALS, Conceição AS, Moura FBP, Santana AEG. 2012. Bioactivity evaluation of plant extracts used in indigenous medicine against the snail, *Biomphalaria glabrata*, and the larvae of *Aedes aegypti*. *Evidence-Based Complementary and Alternative Medicine* 846583: 1-9.
- Santos FA, Jeferson FA, Santos CC, Silveira ER, Rao VSN. 2005. Antinociceptive effect of leaf essential oil from *Croton sonderianus* in mice. *Life Sciences* 77: 2953-2963.
- Scott AJ, Knott M. 1974. A cluster analysis method for grouping means in the analysis of variance. *Biometrics* 30: 507-512.
- Silva M.L, Silva LB, Fernandes RM, Lopes GS. 2013. Efeito do extrato aquoso e etanólico do angico preto sobre larvas de *Rhipicephalus (Boophilus) microplus*. *Arquivo Brasileiro de Medicina Veterinária e Zootecnia* 65: 637-644.
- Silva-Aguayo GI, Kiger-Melivilu R, Hepp-Gallo R, Tapia-Vargas M. 2005. Control de *Sitophilus zeamais* con polvos vegetales de tres espécies del género *Chenopodium*. *Pesquisa Agropecuária Brasileira* 40: 953-960.
- Southgate BJ. 1978. The importance of the Bruchidae as pests of grain legumes, their distribution and control, pp. 219-229 *In* Singh SR, van Emden HF, Taylor TA. [eds.], *Pests of Grain Legumes: Ecology and Control*. Academic Press, London, United Kingdom.
- Souza MCC, Trovão MBM. 2009. Bioatividade do extrato seco de plantas da Caatinga e do Nim (*Azadirachta indica*) sobre *Sitophilus zeamais* Mots em milho armazenado. *Revista Verde de Agroecologia e Desenvolvimento Sustentável* 4: 120-124.
- Souza TM, Farias DF, Soares BM, Viana MP, Lima GPG, Machado LKA, Morais SM, Carvalho AFU. 2011. Toxicity of Brazilian plant seed extracts to two strains of *Aedes aegypti* (Diptera: Culicidae) and nontarget animals. *Journal of Medical Entomology* 48: 846-851.
- Tavares WD, Graziotti GH, de Souza AA, Freitas SD, Consolaro HN, Ribeiro PED, Zanuncio JC. 2013. Screening of extracts of leaves and stems of *Psychotria* spp. (Rubiaceae) against *Sitophilus zeamais* (Coleoptera: Curculionidae) and *Spodoptera frugiperda* (Lepidoptera: Noctuidae) for maize protection. *Journal of Food Protection* 76: 1892-1901.
- Tavares, WD, Faroni LRD, Ribeiro RC, Fouad HA, Freitas SD, Zanuncio JC. 2014. Effects of astilbin from *Dimorphandra mollis* (Fabaceae) flowers and Brazilian plant extracts on *Sitophilus zeamais* (Coleoptera: Curculionidae). *Florida Entomologist* 97: 892-901.
- Torres AL, Boiça Junior AL, Medeiros CAM, Barros R. 2006. Efeito de extratos aquosos de *Azadirachta indica*, *Melia azedarach* e *Aspidosperma pyrifolium* no desenvolvimento e oviposição de *Plutella xylostella*. *Bragantia* 65: 447-457.
- Trevisan MTS, Macedo FVV. 2003. Seleção de plantas com atividade anticolinérgica para tratamento da doença de Alzheimer. *Química Nova* 26: 301-304.
- Vieira C. 2004. Memórias de meio século de estudo sobre a cultura do feijão. Editora UFV, Viçosa-MG, Brazil. 214 pp.