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## Screening strains of the mulberry silkworm, *Bombyx mori*, for thermotolerance

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### Abstract

A tropical climate prevails in most of the sericultural areas in India, where temperature increases during the summer lead to adverse effects on temperate bivoltine silkworm rearing and cause crop losses. Screening for thermotolerance in the silkworm, *Bombyxmori* L. (Lepidoptera: Bombycidae) is an essential prerequisite for the development of thermotolerant breeds/hybrids. In the current study, the aim was to identify potential bivoltine silkworm strains specific for tolerance to high temperature. The third day of fifth stage silkworm larvae of bivoltine strains were subjected to high temperature of  $36 \pm 1^\circ \text{C}$  with RH of  $50 \pm 5\%$  for six hours (10:00-16:00) every day until spinning for three consecutive generations. Highly significant differences were found among all genetic traits of bivoltine silkworm strains in the treated groups. Three groups of silkworm resulted including susceptible, moderately tolerant, and tolerant by utilizing pupation rate or survival rate with thermal stress as the index for thermotolerance. Furthermore, based on the overall silkworm rearing performance of nine quantitative genetic traits such as larval weight, cocoon yield by number and weight, pupation, single cocoon and shell weight, shell ratio, filament length and denier, three bivoltine silkworm strains, BD<sub>2</sub>-S, SOF-BR and BO<sub>2</sub> were developed as having the potential for thermotolerance. The data from the present study enhance knowledge for the development of thermo tolerant silkworm breeds/ hybrids and their effective commercial utilization in the sericulture industry.

**Keywords:** genetic traits, rearing performance, survival rate, temperature stress, thermotolerant

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## Introduction

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The mulberry silkworm, *Bombyx mori* L. (Lepidoptera: Bombycidae), is a very heat-sensitive organism. Intensive and careful domestication over centuries has apparently deprived this commercial insect of the opportunity to acquire thermotolerance. This vulnerability is more pronounced in bivoltine strains compared to polyvoltine strains. Thus, among many factors responsible for poor performance of the bivoltine strains under tropical conditions, the major one is lack of thermotolerance. Many quantitative characters decline sharply at higher temperatures. Therefore, one of the key considerations in developing bivoltine hybrids for tropics could be the need for thermotolerant bivoltine strains. The recent advances in silkworm breeding and those in stress-induced protein synthesis have opened up new avenues to evolve robust productive silkworm hybrids (Evgenov et al. 1987; Kumar et al. 1989; Coulon-Bublex and Mathelin 1991; Wu and Hou 1993; Joy and Gopinathan 1995; Kumar et al. 2001, 2002, 2003; Reddy et al. 2002; Vasudha et al. 2006; Srivastava et al. 2007; Moghaddam et al. 2008).

Sericulture in India is practiced predominantly in tropical environmental regions such as Karnataka, Tamil Nadu, Andhra Pradesh, and West Bengal and to a limited extent in the temperate environment of Jammu and Kashmir. This situation provides scope for creating polyvoltine x bivoltine hybrids as a commercial venture as hybrids are hardy and have the ability to survive and reproduce under varied or fluctuating environmental climatic conditions.

However, hybrid quality is low when compared to the existing international

standards. For example, cross breeds of polyvoltine female x bivoltine male are generally reared in these regions during the summer, but the quality of cocoon production is not as high as it is for bivoltine silkworm hybrids (Ramesha et al. 2009). Bivoltine silkworm breeds are known for their qualitative and quantitative traits in the sericulture industry. During the last decade, a number of silkworm hybrids have been developed (Basavaraja et al. 1995; Data et al. 1997) and selected for exploitation at the field level during favorable seasons. These productive bivoltine breeds are susceptible to varied environmental conditions, as bivoltine silkworm breeds originated from temperate regions. Many important qualitative characters such as viability and cocoon traits decline sharply when temperature exceeds 28° C. As India has fluctuating temperature and humidity conditions, estimation of phenotypic stability at high temperature is considered of prime importance for sustainable progress in bivoltine breeding.

Previous studies have demonstrated fundamental thermotolerance in silkworms (Ueda and Lizuka 1962; Pillai and Krishnaswami, 1980, 1987; Kato et al. 1989) but information regarding the implications of these aspects of selection of parental resources for breeding programs is lacking. Some earlier studies addressed selection of silkworm breeds in respect of thermotolerance by identifying thermotolerant silkworm breeds (Shirota 1992; Tazima and Ohnuma 1995; Kumar et al. 2001). However, a clear understanding of the genetic basis and variability in the expression of quantitative and qualitative genetic traits during exposure to high temperatures is an important step for the selection of potential thermotolerance parental resources for breeding programs.

The purpose of this study is to obtain new data about screening for thermotolerance in silkworm larvae, not only to augment current knowledge on gene expression under stress conditions, but also to provide valuable information that will allow identification of thermotolerant bivoltine silkworm breeds based on the silkworm rearing performances relative to nine important economical genetic traits.

## Materials and Methods

### Silkworm breeds

The 24 bivoltine silkworm breeds used were, BD<sub>2</sub>, BO<sub>1</sub>N, BD<sub>3</sub>N, SOF-BR, BD<sub>2</sub>-S, BD<sub>1</sub>C-BR, BO<sub>3</sub>-W, SOW, SOCB, SB-F, BD<sub>2</sub>-G, BO<sub>3</sub>BL, BO<sub>1</sub>BL, BO<sub>2</sub>, SOHW, BO<sub>1</sub>BR, BD<sub>1</sub>LC, BD<sub>1</sub>O, BD<sub>2</sub>-LC, BD<sub>3</sub>, BO<sub>1</sub>-S, BD<sub>4</sub>, BO<sub>3</sub>, NB<sub>4</sub>D<sub>2</sub>. These strains with varied phenotypic and quantitative traits, including hibernation, are maintained at the Biology Division, Indian Institute of Chemical Technology, Hyderabad, AP, India.

### Silkworm rearing and estimation of genetic traits

Disease free eggs from each strain were reared and cocoons were harvested and maintained until eclosion of moths. Healthy female moths emerging on the peak day of eclosion were allowed to mate for three to four hours and held until oviposition. The eggs were acid-treated within 20 hours after oviposition, following the method developed by Yokoyama (1962) to prevent hibernation. The eggs were incubated at  $25 \pm 1^\circ \text{C}$  temperature and 70 to 80% RH after surface treatment with 2% formalin solution. Twenty to 30 eggs were chosen from each brood and pasted onto egg sheets. Three such egg sheets for each breed were prepared, wrapped in white tissue paper and boxed with black paper to

synchronize the embryonic development. On the day of hatching, the eggs were exposed to light in order to obtain uniform hatching and freshly chopped mulberry leaves were fed to the young larvae. The whole process, from silkworm egg incubation to completion of rearing activities, was carried out under hygienic conditions in a silkworm rearing laboratory thoroughly disinfected with bleaching powder and formalin solution.

Silkworm rearing was conducted for each breed in plastic boxes by feeding them the V<sub>1</sub> variety of mulberry leaves from the well-maintained irrigated mulberry garden on campus. A standard rearing procedure was adopted as recommended by Datta (1992). The young larvae (1<sup>st</sup>-3<sup>rd</sup> instars) were reared at  $26\text{-}28^\circ \text{C}$  with 80-90% RH and late age larvae (4<sup>th</sup> and 5<sup>th</sup> instars) were maintained at  $24\text{-}26^\circ \text{C}$  with 70-80% RH until the 3rd day of fifth instar. Each group was divided into two, one of which was maintained as a control under standard rearing conditions and the second was exposed to high temperature treatment.

### High temperature treatment

The study was carried out between March 2007 and January 2008. Silkworm rearing was conducted following the standard method under the recommended temperature and relative humidity until the second day of the fifth instar. On the third day of the fifth instar, 300 larvae per breed in three replications of 100 larvae were selected for the high temperature treatment. High temperature treatment was obtained by utilizing the NK System Biotron ([www.nihonika.co.jp](http://www.nihonika.co.jp)), an environmental growth chamber with precise and automatic control facilities for uniform maintenance of temperature and humidity. The temperature used was  $36 \pm 1^\circ \text{C}$  and RH  $50 \pm 5\%$ . Fresh mulberry leaves were given

twice a day and silkworm rearing was continued as suggested by Kato et al. (1989) and Kumar et al. (2001) using appropriate plastic boxes/trays. A control group was maintained at ambient temperature of standard rearing conditions at  $25 \pm 1^\circ\text{C}$  and RH  $65 \pm 5\%$ . Thermal exposure was given every day for six hours until spinning (10:00-16:00) since continuous exposure to high temperature conditions reduces quantitative traits drastically in high thermotolerance screening experiments. Observations were carried out daily and mortality due to high temperature in each of the 24 breeds was noted. After thermal treatment, the treated silkworm larvae were shifted to the moutage for spinning at normal ambient temperature of  $25 \pm 2^\circ\text{C}$  and RH  $65 \pm 5\%$ . Cocoons were harvested 4-5 days later after completion of cocoon spinning. Harvested cocoons were accessed for survival to pupation using the equations detailed below. The pupation or survival rate was utilized as the measure of index for assessing thermotolerance by calculating the number of healthy live pupae obtained relative to the number of larvae at the beginning of the treatment both in treated and control groups.

During the process of silkworm rearing, data on larvae and cocoons for the nine genetic traits (larval weight, cocoon yield for 10,000 larvae by number and weight, pupation rate, cocoon and shell weight, shell ratio, filament length and denier) were collected and calculated according to the equations below:

**Larval weight (g).** Mean larval weight (g) recoded for 10 randomly selected larvae at the peak of growth of fifth instar larvae from each replication. This was an indicator of the general health of the larvae.

**Cocoon yield for 10,000 larvae.** The mean number of cocoons harvested relative to the

number of larvae at the beginning of the experiment, converted to 10,000 larvae.

**Cocoon yield by weight (kg) for 10,000 larvae.** The mean weight of the cocoons harvested in kilogram (kg) for every 10,000 larvae by weight.

**Pupation rate (%).** The live pupa present inside the cocoon during metamorphosis of larva into pupa expressed as a percentage. This genetic trait was kept as the ultimate index for assessing thermotolerance in this investigation.

$$\frac{\# \text{ of good cocoons} + (\# \text{ of double cocoons} \times 2)}{\# \text{ of larvae beginning the experiment}} \times 100$$

**Cocoon weight (g).** The average single cocoon weight in grams for 10 male and 10 female cocoons chosen randomly on the 6<sup>th</sup> or 7<sup>th</sup> day of spinning.

**Shell weight (g).** The average single cocoon shell weight in grams of 10 male and 10 female cocoons shell chosen randomly. The shells used were the same cocoons used for the cocoon weight determination.

**Shell ratio (%).** The total quantity of silk available from a single cocoon was expressed as a percentage using the following equation.

$$\frac{\text{(single cocoon shell weight (g))}}{\text{single cocoon weight (g)}} \times 100$$

**Filament length (m).** This is the most important parameter used by the industry. Silk filament length indicates the reelable length of silk filament from a cocoon. It was calculated using the average length of unwound silk filament from 10 cocoons (obtained using a

**Table 1.** Silkworm rearing performance of bivoltine germplasm resources under normal and high temperature conditions.

Breed	Weight of 10 larvae (g)		Yield /10,000 larvae by number		Yield /10,000 larvae by weight (g)		Pupation rate (%)	
	Control	Treated	Control	Treated	Control	Treated	Control	Treated
BD <sub>2</sub>	40.9	39.3	9218	5784	15.38	7.39	90.4	66.8
BO <sub>1</sub> N	41.8	38.2	9143	7146	14.99	9.61	89.56	69.78
BD <sub>1</sub> N	41.3	37.7	9276	6967	14.1	9.93	92.28	68.78
SOF-BR	37.7	35	8967	4377	12.44	6	89.4	74.29
BD <sub>2</sub> -S	41.3	37.8	9345	6199	15.2	9.02	91.61	76.07
BD <sub>1</sub> C-BR	40.3	38.1	9100	3869	13.43	3.73	89.35	67.85
BO <sub>1</sub> -W	42	40.4	9259	3576	13.46	3.49	88.96	65.44
SOW	37.8	35.6	9306	6232	12.2	10.4	88.18	61.6
SOCB	38.3	36.2	9219	4206	12.52	4.08	87.29	71.21
SB-F	38.4	34.7	9160	6795	12.52	7.85	89.32	67.09
BD <sub>2</sub> -G	41.1	38.5	9153	6420	12.88	8.99	90.41	63.78
BO <sub>1</sub> BL	41	37.9	9150	4239	12.32	4.12	87.59	71.8
BO <sub>1</sub> BL	41	38.7	9083	5869	12.53	9.01	90.13	58.09
BO <sub>2</sub>	42.2	38.5	9133	7386	13.5	9.02	88.71	73.25
SOHW	38.4	35.8	9251	3912	12.25	3.78	90.85	68.18
BO <sub>1</sub> BR	39.7	38	9197	5891	13.77	9.59	90.42	58.09
BD <sub>1</sub> LC	40.8	39.1	9091	6685	14.17	9.73	89.99	65.9
BD <sub>1</sub> O	41	37.8	9066	7190	13.86	12.06	90.12	71.11
BD <sub>2</sub> -LC	41.4	38.8	9030	6683	13.94	8.66	89.38	65.78
BD <sub>1</sub>	40.3	38.1	9037	3894	12.95	3.74	89.28	68
BO <sub>1</sub> -S	41	38.3	9138	2191	13.14	2.12	88.9	71.39
BD <sub>1</sub>	41.4	38.9	8997	5796	13.37	8.78	89.16	57.12
BO <sub>1</sub>	41.1	37.2	9156	5590	13.36	9.39	89.54	54.87
NB <sub>1</sub> D <sub>2</sub>	40.8	37.2	9087	6089	15.35	8.06	89.39	59.95
Mean	40.5**	37.7**	9132**	6666**	12.64**	9.34**	88.56**	66.50**
Minimum	37.7	34.7	8967	2191	12.2	2.12	87.29	54.87
Maximum	42.2	40.4	9345	7386	15.38	12.06	92.28	76.07
Std. Error	0.161	0.04648	17.7057	5.1112	0.05847	0.01688	0.87944	0.25387
CD at 5%	0.45197	0.13047	49.7033	14.34811	0.16415	0.04739	2.46874	0.71267
F Ratio	67.31	5.04	232.1	242.7	170.25	68.56	11.24	11.84

\*\*Significant at p &lt; 0.01

mono cocoon reeling unit) and expressed in meters according to the following formula:

$$\frac{\text{length of raw silk (m)} \times 1.125 \text{ (circumference)}}{\text{\# of reeling cocoons}}$$

**Denier (d).** The thickness of the silk filament measured by following formula.

$$\frac{\text{(weight of total silk filament (g)) / total filament length of silk}}{\times 9000}$$

The data on rearing performance and post cocoon parameters of both high temperatures treated and control groups were recorded on nine genetic traits for each replicate. This was subjected to statistical analysis with assistance of 'Indostat' software for understanding the significance of the study by ANOVA.

## Results

### Morphological salient features of bivoltine breeds

**Table 2.** Post cocoon parameters of bivoltine germplasm resources under normal and high temperature conditions.

Breed	Cocoon weight		Shell ratio		Filament length		Denier			
	Control	Treated	Control	Treated	Control	Treated	Control	Treated		
BD <sub>2</sub>	1.669	1.31	0.34	0.26	20.38	19.82	831	791	2.5	2.3
BO <sub>1</sub> N	1.624	1.364	0.341	0.283	21.13	20.75	1022	1069	2.4	2.3
BD <sub>1</sub> N	1.524	1.305	0.304	0.254	19.95	19.5	809	818	2.3	2.2
SOF-BR	1.407	1.421	0.246	0.243	17.5	17.11	623	634	2.4	2.3
BD <sub>2</sub> -S	1.635	1.329	0.211	0.287	21.08	20.08	859	855	2.6	2.6
BD <sub>1</sub> C-BR	1.502	1.375	0.297	0.253	19.74	18.39	764	755	3.2	3.1
BO <sub>1</sub> -W	1.508	1.453	0.284	0.238	18.82	16.38	637	590	2	1.9
SOW	1.315	1.235	0.248	0.245	18.87	19.86	742	689	2.3	2.2
SOCB	1.364	1.331	0.265	0.236	19.43	17.77	671	659	2	1.9
SB-F	1.357	1.258	0.254	0.245	18.68	19.45	721	750	2.4	2.2
BD <sub>2</sub> -G	1.39	1.398	0.277	0.274	19.9	19.61	827	813	2.5	2.3
BO <sub>1</sub> BL	1.366	1.388	0.254	0.267	18.46	19.24	627	722	2.3	2.1
BO <sub>1</sub> BL	1.5	1.399	0.289	0.257	19.26	18.37	716	780	2.3	2.2
BO <sub>2</sub>	1.509	1.416	0.329	0.303	21.78	21.53	810	822	2.4	2.3
SOHW	1.349	1.484	0.271	0.288	20.1	19.41	758	730	2.3	2.3
BO <sub>1</sub> BR	1.52	1.382	0.311	0.277	20.44	20.01	839	810	2	1.9
BD <sub>1</sub> LC	1.542	1.39	0.314	0.272	20.38	19.58	781	816	3.3	3.1
BD <sub>1</sub> O	1.54	1.454	0.303	0.289	19.69	19.84	776	737	3.1	3
BD <sub>1</sub> -LC	1.513	1.311	0.293	0.238	19.37	17.88	732	707	2.5	2.3
BD <sub>2</sub>	1.445	1.411	0.306	0.252	21.22	17.88	867	607	2.3	2.1
BO <sub>1</sub> -S	1.45	1.359	0.308	0.24	21.25	17.7	886	617	2	2
BD <sub>1</sub>	1.506	1.433	0.326	0.28	21.64	19.53	784	934	2.4	2.2
BO <sub>2</sub>	1.492	1.428	0.305	0.303	20.45	21.23	772	806	2.1	2
NB <sub>1</sub> D <sub>2</sub>	1.699	1.352	0.314	0.25	18.47	18.49	733	712	2.6	2.4
Mean	1.429**	1.390**	0.284**	0.266**	19.87**	19.12**	775**	759**	2.4**	2.3**
Minimum	1.315	1.235	0.211	0.236	17.5	16.38	623	590	2	1.9
Maximum	1.699	1.484	0.341	0.303	21.78	21.53	1022	1069	3.3	3.1
Std. Error	0.003	0.001	0.002	0	0.099	0.028	2.181	0.63	0.005	0.0015
CD at 5%	0.008	0.002	0.004	0.001	0.277	0.08	6.122	1.767	0.015	0.0043
F Ratio	435.62	228.24	281.67	71.24	164.28	39.62	1645.04	426.21	4380.3	40.53

\*\*Significant at p &lt; 0.01

Morphological differences were found between the germplasm breeds with respect to origin, egg, larval and cocoon traits. The 24 breeds were not of Indian origin. The chorion of 13 breeds was brown, whereas 11 breeds were pigmented, and the color of the serosa of all breeds was white. Larval marking of all breeds was plain, 19 breeds had a stout body and 5 had a slender body. Cocoon shape in 13 breeds was oval, 6 were dumbbell-shaped and 5 had peanut-shaped cocoons. Most of the breeds spun a white cocoon but SOCB and SOHW spun dull white color cocoons. Fine cocoon grains occurred in 17 breeds, medium grains occurred in 4 breeds, and coarse grains occurred in 3 breeds.

### Rearing performance

Considerable variations were found for 9 genetic traits among the bivoltine breeds at high temperature. Data were obtained for larva weight, yield by 10,000 larvae by number and weight, pupation, cocoon weight, shell weight, shell ratio, filament length and denier for 24 bivoltine breeds under normal and high temperature treatments. There was evidence of clear declines in all nine genetic traits in all of the high temperature treated

groups (Tables 1 and 2). Among high temperature treated groups, maximum larval weight in BO<sub>3</sub>-W (40.4 g) followed by BD<sub>2</sub> (39.3 g), BD<sub>1</sub>LC (39.1 g) and minimum in SB-F (34.7 g) with an average of 37.7 g was estimated. Highest yield /10,000 larvae by number was observed in BO<sub>2</sub> (7386) and lowest in BO<sub>1</sub>- S (2191) with an average of 5541. Maximum yield /10,000 larvae by weight were in SOW (10.40 kg), minimum in BO<sub>1</sub>-S (2.12 kg) with the mean value of 7.35 kg. The highest pupation rate of 76.07% in BD<sub>2</sub>-S was followed by 74.29% in SOF-BR, 73.25% in BD<sub>2</sub> and lowest of 54.87% in BO<sub>3</sub> with a mean value of 66.51% (Table 1).

### **Pupation / survival rate (index of thermotolerance)**

Based on the pupation/survival rate, all of the breeds were grouped for relative temperature tolerance. Among them, 7 breeds had pupation rate > 70% for the high temperature treated groups and were considered to be tolerant to high temperature (Table 1). Amongst them, maximum pupation was observed in BD<sub>2</sub>-S (76.07%) followed by SOF-BR (74.29%), BO<sub>2</sub> (73.25%), BO<sub>3</sub>- BL (71.80 %), BO<sub>1</sub>-S (71.39%), SOCB (71.21%), BD<sub>1</sub>O (71.11%). Twelve germplasm breeds were found to exhibit < 60% of a pupation rate ranging from 69.78 % (BO<sub>1</sub>N) to 61.60 % (SOW) and were considered to be moderately tolerant to high temperature. While 5 germplasm breeds *viz.*, BO<sub>1</sub>BR, BO<sub>1</sub>-BL, BD<sub>4</sub>, BO<sub>3</sub> and NB<sub>4</sub>D<sub>2</sub> were identified as less tolerant (< 60% of pupation) to high temperature with pupation rates ranging from 58.09% (BO<sub>1</sub>BR) to 54.87% (BO<sub>3</sub>). By considering the pupation/survival rate as the index for thermotolerance, the bivoltine germplasm resource was classified as susceptible (BO<sub>1</sub>BR, BO<sub>1</sub>-BL, BD<sub>4</sub>, BO<sub>3</sub> and NB<sub>4</sub>D<sub>2</sub>), moderately tolerant (BD<sub>2</sub>, BO<sub>1</sub>N, BD<sub>3</sub>N, BD<sub>1</sub>C-BR, BO<sub>3</sub>-W, SOW, SB-F, BD<sub>2</sub>-

G, SOHW, BD<sub>1</sub>LC, BD<sub>2</sub>-LC, BD<sub>3</sub>) and tolerant (BO<sub>1</sub>-S, BD<sub>2</sub>-S, SOCB, BO<sub>3</sub>BL, BO<sub>2</sub>, BD<sub>1</sub>O, SOF-BR) to thermal stress using the following ranges: susceptible, 50-60%; moderately tolerant, 61-70%; and tolerant, 71-90%. With respect to percentage difference between the high temperature groups and control, negative correlations were seen for all genetic traits (Figure 1).

### **Performance of cocoon traits**

Maximum single cocoon weight in SOHW was 1.484 g and minimum single cocoon weight in SOW was 1.235 g, with an average cocoon weight of 1.374 g. With respect to shell weight, the highest was 0.303 g in BO<sub>2</sub> and the lowest was 0.236 g in SOCB, with an average of 0.266 g. An average of 19.14% shell ratio with the maximum of 21.53% and minimum of 16.38% was found. BO<sub>1</sub>N showed highest filament length of 1069 m, followed by BD<sub>4</sub> with 934 m, BD<sub>2</sub>-S with 855 m and lowest of 590 m in BO<sub>3</sub>-W. The average among the breeds was 759 m. The relevant to negative correlated trait of denier reached a maximum of 3.1d in BD<sub>1</sub>C-BR, BD<sub>1</sub>LC and a minimum of 1.9d in BO<sub>2</sub>-W, SOCB, BO<sub>1</sub>BR with an average of 2.3d among bivoltine breeds (Table 2).

### **Percentage of difference in treated over control groups**

The percentage of difference among treated and control groups was the highest in BO<sub>3</sub>-W (-3.81%) and the lowest in SB-F (-9.64%) for the trait of larval weight. BO<sub>2</sub> (19.13%) and BD<sub>1</sub>-S (76.02%) showed high and low percentages of difference for the traits of cocoon yield by number, respectively. Maximum difference occurred in BD<sub>1</sub>O (-12.99%) and minimum difference in BO<sub>1</sub>-S (-83.87%) for cocoon yield by weight. With respect to cocoon weight, a positive difference in SOHW (10.01%) and negative difference in

**Table 3.** Percentage of difference on genetic traits in bivoltine germplasm resources of high temperature conditions groups over control.

Breed	Weight of 10 larvae	Yield /10,000 larvae by No	Yield /10,000 larvae by weight	Pupation rate	Cocoon weight	Shell weight	Shell ratio	Filament length	Denier
BD <sub>2</sub>	-3.91	-37.25	-51.95	-26.11	-21.51	-23.53	-2.74	-4.81	-5.09
BO <sub>1</sub> N	-8.61	-21.84	-35.89	-22.09	-16.01	-17.01	-1.82	4.6	-3.33
BD <sub>3</sub> N	-8.72	-24.89	-29.57	-25.47	-14.37	-16.45	-2.26	1.11	-5.66
SOF-BR	-7.16	-51.19	-51.77	-16.9	1	-1.22	-2.24	1.77	-4.84
BD <sub>2</sub> -S	-8.47	-33.67	-40.66	-16.96	-18.72	36.02	-4.75	-0.47	-2.27
BD <sub>1</sub> C-BR	-5.46	-57.48	-72.23	-24.06	-8.46	-14.81	-6.85	-1.18	-3.28
BO <sub>1</sub> -W	-3.81	-61.38	-74.07	-26.44	-3.65	-16.2	-12.97	-7.38	-3.32
SOW	-5.82	-33.03	-14.75	-30.14	-6.08	-1.21	5.25	-7.14	-6.51
SOCB	-5.48	-54.38	-67.41	-18.42	-2.42	-10.94	-8.56	-1.79	-2.81
SB-F	-9.64	-25.82	-37.3	-24.89	-7.3	-3.54	4.13	4.02	-10.21
BD <sub>1</sub> -G	-6.33	-29.86	-30.2	-29.45	0.58	-1.08	-1.47	-1.69	-5.7
BO <sub>1</sub> BL	-7.56	-53.67	-66.56	-18.03	1.61	5.12	4.23	15.15	-9.68
BO <sub>1</sub> BL	-5.61	-35.38	-28.09	-35.55	-6.73	-11.07	-4.62	8.94	-2.82
BO <sub>2</sub>	-8.77	-19.13	-33.19	-17.43	-6.16	-7.9	-1.13	1.48	-5.82
SOHW	-6.77	-57.71	-69.14	-24.95	10.01	6.27	-3.43	-3.69	-2.36
BO <sub>1</sub> BR	-4.28	-35.95	-30.36	-35.76	-9.08	-10.93	-2.1	-3.46	-3.79
BD <sub>1</sub> LC	-4.17	-26.47	-31.33	-26.77	-9.86	-13.38	-3.95	4.48	-4.29
BD <sub>1</sub> O	-7.8	-20.69	-12.99	-21.09	-5.58	-4.62	0.78	-5.03	-3.05
BD <sub>2</sub> -LC	-6.28	-25.99	-37.88	-26.4	-13.35	-18.77	-7.7	-3.42	-8.91
BD <sub>1</sub>	-5.46	-56.91	-71.12	-23.84	-2.35	-17.65	-15.75	-29.99	-5.96
BO <sub>1</sub> -S	-6.59	-76.02	-83.87	-19.7	-6.28	-22.08	-16.72	-30.36	-0.76
BD <sub>4</sub>	-6.04	-35.58	-34.33	-35.94	-4.85	-14.11	-9.76	19.13	-11.29
BO <sub>3</sub>	-9.49	-38.95	-29.72	-38.72	-4.29	-0.66	3.83	4.4	-7.94
NB <sub>4</sub> D <sub>2</sub>	-8.82	-32.99	-47.49	-32.93	-20.42	-20.38	0.12	-2.86	-7.5

BD<sub>2</sub> (-21.51%) was found. For shell weight, a maximum positive difference was found in BD<sub>2</sub>-S (36.02%) and a minimum positive difference was found in BD<sub>2</sub> (-23.53%). For shell ratio, a high positive difference was found in SOW (5.25%) and a low positive difference was found in BO<sub>1</sub>-S (-16.72%). With respect to filament length, maximum positive difference occurred in BD<sub>4</sub> (19.13%) and minimum positive difference occurred in BO<sub>1</sub>-S (-30.36%). For the trait of denier, a high difference occurred in BO<sub>1</sub>-S (-0.76%) and low difference occurred in BD<sub>4</sub> (-11.29%) (Table 3).

### Positive percentage difference on genetic traits

Among the 9 genetic traits of breeds, 4 important traits such as cocoon weight, shell weight, shell ratio and filament length were found to exhibit the positive difference in the high temperature treated group over the control group (Table 3). In the case of cocoon weight, positive percentage differences were observed in SOHW (10.01%), followed by BO<sub>3</sub> BL (1.61%), SOF-BR (1.00%), BD<sub>2</sub>-G (0.58%) (Figure 2). The maximum difference in BD<sub>2</sub>-S (36.02%), which was followed by

SOHW (6.27%) and BO<sub>3</sub>BL (5.12%) was revealed on shell weight (Figure 3). High positive difference in SOW (5.25%) followed by BO<sub>3</sub>BL (4.23%), SB-F (4.13%), BO<sub>3</sub> (3.83%) and BD<sub>1</sub>O (0.78%), NB<sub>4</sub>D<sub>2</sub> (0.12%) occurred on shell ratio (Figure 3). For filament length, maximum difference occurred in BD<sub>4</sub> (19.13%) followed by BO<sub>3</sub>BL (15.15%), BO<sub>1</sub>BL (8.94%), BO<sub>1</sub>N (4.60%), BD<sub>1</sub>LC (4.48%), BO<sub>3</sub> (4.40%), SB-F (4.02%), SOF-BR (1.77%), BO<sub>2</sub> (1.48%) and BD<sub>3</sub>N (1.11%) (Figures 2 and 4).

The collected data for the 9 genetic traits were statistically analyzed by the variance test for significant differences between the normal and treated groups. Results obtained after analysis of variance of mean squares revealed highly significant ( $p \leq 0.001$ ) differences among all genetic traits for the breeds. The maximum critical difference of 5% was demonstrated for the trait of yield/10,000 larvae by number (14.3481) among high temperature treated groups, which was followed by filament length (1.767), pupation rate (0.71267) yield /10,000 larvae by weight, filament length (6.122) and shell weight (0.001) (Tables 1 and 2). All genetic traits showed a decline with increase of temperature compared to the controls.

Based on all morphological genetic traits along with the maximum thermotolerance level on pupation rate, three bivoltine silkworm germplasm breeds *viz.*, BD<sub>2</sub>-S, SOF-BR and BO<sub>2</sub>, were identified as potential thermo tolerant breeding resources for breeding programs.

### Discussion

On thermal treatment, all genetic traits of silkworms showed a decline with the increase of temperature above standard level. A similar



result was reported by Kumar et al. (2002). They found that biological molecules like DNA, RNA, lipids, etc. were vulnerable to heat stress. Temperature stress causes a number of abnormalities at the cellular level as the normal pattern of protein synthesis halts. Another important effect of temperature (or stress of any kind) is the unfolding of cellular proteins. Cellular proteins are typically folded in their native conformations while functioning in cells. This process can result in aggregates of unfolded protein that at best diminish the pool of functional proteins and at worst are cytotoxic (Feder 1996; Feder et al. 1996). However, a brief exposure of cells to sub-lethal high temperature was found to render protection to the organism of subsequent and more severe temperature changes (Denlinger and Yocum 1998; Gilchrist and Huey 1999; Kevin 2002). In a study with heat-shocked *Drosophila*, continued survival and acclimatization of the experimental insects occurred at higher temperatures (Dingley and Maynard Smith 1968). Further, every form of stress is known to induce a set of proteins in all tested organisms from bacteria to man. Inducing stressors include ethanol, heavy metals, hypoxia, hyperoxia, changes in pH, free radicals, various poisons and toxins, ischemia, osmotic shock, ionizing radiation and many others (Feder 1996). The heat shock response in *B. mori* cells induced active transcription of heat shock mRNAs (Evegnev et al. 1987; Kiuchi et al. 2007). It was reported that 93, 70, 46 and 28 kDa protein bands appeared after exposure to high temperature in both bivoltine and polyvoltine strains, but with varying kinetics. The isolated hemocytes of polyvoltine breeds exhibited the induction of 70 kDa protein (Joy and Gopinathan 1995). For instance, polyvoltine breeds reared in tropical countries are known to tolerate slightly higher temperature, as are cross

breeds that have evolved for a tropical climate (Ramesha et al. 2009).

The success of the sericulture industry depends upon several variables, but environmental conditions such as biotic and abiotic factors are of particular importance. Among the abiotic factors, temperature plays a major role on growth and productivity of silkworms (Benchamin and Jolly 1986). It is also known that the older silkworms prefer relatively lower temperature than younger silkworms and that fluctuation of temperature during different stages of larval development was found to be more favorable for growth and development of larvae than constant temperature. There is ample literature stating that good quality cocoons are produced within a temperature range of 22-27° C and that cocoon quality is poorer above these levels (Krishnaswami et al. 1973; Datta 1992; Datta et al. 1996, 1997). However, polyvoltine breeds reared in tropical countries are known to tolerate slightly higher temperature (Hsieh et al. 1995), as are cross breeds that have been developed for tropical climates. In order to use bivoltine races in a tropical country like India, it is necessary to have a stable cocoon crop in a high temperature environment. High temperature affects nearly all biological processes including the rates of biochemical and physiological reactions (Hsieh et al. 1995; Willmer et al. 2004), and can eventually affect the quality or quantity of cocoon crops in the silkworm. Several reports (Ueda and Lizuka 1962; Shirota 1992; Tazima and Ohuma 1995; Hsieh et al. 1995) demonstrated that silkworms were more sensitive to high temperature during the fourth and fifth stages, which are recommended for the recognition and selection of thermo tolerant silkworm breeds, under high temperature conditions. Recently, the effects of temperature on the

development and survival of the Argentine ant also have been shown by Abril et al. (2010).

It is well understood that the majority of the economically important genetic traits of silkworms are qualitative in nature and that phenotypic expression is greatly influenced by environmental factors such as temperature, relative humidity, light and nutrition (Krishnaswami et al. 1971; Pillai and Krishnaswami 1980, 1987; Wu and Hau 1993; Tazima 1984; Thairarajan et al. 1993; Zhang et al. 2002; Zhao et al. 2007; Ramesha et al. 2010). Therefore, it is essential to gauge the degree of phenotypic difference of the economical traits to understand the genetic steadiness under the varied environmental conditions and the productivity of germplasm breeding resources. The problem of balancing and fixing the desirable traits for a given environment is a challenging task for the breeder. Hence, understanding the range of reactions of selected breeds to variable environmental conditions is important for the breeder to utilize them appropriately in hybrid programs. Intensive and careful domestication over centuries has apparently deprived the insect of opportunities to acquire thermotolerance. Among many factors responsible for poor performance of the bivoltine strains under tropical conditions, the main culprit is temperature. Indeed, many quantitative characters decline sharply when temperature is higher than 28° C. Breeders in the field agree that it is a difficult task to breed such bivoltine breeds that are suitable to high temperature environments with productive traits. It is a well-established fact that under tropical conditions, bivoltine races are more vulnerable to various stresses including climatic conditions, poor leaf quality and improper management during the summer. In order to efficiently select breeds with high temperature tolerance, it is

important to analyze the impact of high temperature on many genetic traits of these silkworm breeds.

In order to achieve greater success in this regard, it is important to understand the level of temperature tolerance in silkworm bivoltine breeds. The main objective of this study was to identify bivoltine silkworm breeds tolerant to high temperature among 24 germplasm resources evaluated for tolerance to thermal stress at  $36 \pm 1^\circ\text{C}$  and relative humidity (RH)  $50 \pm 5\%$  analysis by measuring qualitative and quantitative genetic traits for three successive generations. The results obtained support the earlier observation of Kumar et al. (1989), Kato et al. (1989), Koundinya et al. (2003). The latter authors' emphasis was on the phenotypic manifestations of 9 genetic traits under high temperature conditions. The results revealed highly significant ( $p \leq 0.01$ ) variability among the germplasm resources of bivoltine silkworm germplasm with respect to nine genetic traits on treated groups relating to survival or pupation rate. The benchmark for short listing toward thermotolerance was kept as pupation rate of  $>70\%$  for bivoltine as suggested by Kumar et al. (2002). Although earlier studies have shown that breeds like Hosa Mysore, KA and their hybrids are moderately tolerant to high temperature up to  $38^\circ\text{C}$  (Pillai and Krishnaswami 1987; Shirota 1992), the current study indicates that seven bivoltine germplasm were identified as temperature tolerant with a pupation rate of above 70%.

This study shows less percentage decline in  $\text{BO}_3\text{W}$ ,  $\text{BO}_2$ ,  $\text{BD}_1\text{O}$ ,  $\text{BD}_2\text{S}$  for larval weight, cocoon yield by number, by weight, and pupation rate respectively, better performance over control in  $\text{BD}_2\text{-G}$ ,  $\text{SOF-BR}$ ,  $\text{BO}_3\text{BL}$ ,  $\text{BD}_2\text{-S}$ ,  $\text{SOHW}$  for shell weight, and in  $\text{NB}_4\text{D}_2$ ,  $\text{BD}_1\text{O}$ ,  $\text{SB F}$ ,  $\text{BO}_3\text{-BL}$  for shell ratio

with maximum pupation rates in BD<sub>2</sub>-S, SOF-BR, BO<sub>2</sub>, BO<sub>3</sub>-BL, BD<sub>1</sub>O indicating a capacity for thermotolerance.

In view of the above observations, it was a difficult task to break the negative correlation associated with survival rate and productivity traits. In the present study, bivoltine germplasm resources with maximum pupation or survival rate were identified as potential breeding resource material for the development of thermotolerant breeds/hybrids. This resulted in the identification of BD<sub>2</sub>-S, SOF-BR, and BO<sub>2</sub> as temperature tolerant bivoltine germplasm resources based on their pupation or survival rate as the index and their performance on nine quantitative genetic traits. These resources are recommended as potential breeding resources for the development of thermotolerant silkworm breeds/ hybrids.

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### References

Abril S, Olveras J, Gómez C. 2010. Effect of temperature on the development and survival of the Argentine ant, *Linepithema humile*. *Journal of Insect Science*, 10:97. Available online, <http://www.insectscience.org/10.97>.

Basavaraja HK, Nirmal Kumar S, Suresh Kumar N, Mal Reddy N, Kshama Giridhar Ahsan MM, Datta RK. 1995. New productive bivoltine hybrids *Indian Silk* 34: 5-9.

Benchamin KV, Jolly MS. 1986. Principles of silkworm rearing. In: Mahalingam S. Editor. *Proceedings of Seminar on Problems and Prospects of Sericulture*. pp. 63-106. Vellore, India.

Coulon-Bublex M, Mathelin J. 1991. Variations in the rate of synthesis of heat shock proteins HSP70, between laying and neurula, the diapausing embryo of the silkworm, *Bombyx mori*. *Sericologia* 3: 295-300.

Datta RK. 1992. *Guidelines for Bivoltine Rearing*. pp. 24. Central Silk Board, Bangalore, India.

Datta RK, Basavaraja HK, Mano Y. 1996. *Manual on bivoltine rearing race maintenance and multiplication*. pp. 65. Central Sericultural Research and Training and Institute, Mysore.

Datta RK, Basavaraja HK, Suresh Kumar N, Kishor Kumar CM, Nirmal Kumar S. 1997. Evolution of robust hybrids of bivoltine silkworm, *Bombyx mori* L. for tropics. *XVIIth Congress of the International Sericulture Commission*, Brazil, 22-26<sup>th</sup> April.

Denlinger DL, Yocum GD. 1998. Physiology of heat sensitivity in insects. In: Hallman GJ, Denlinger DL, Editors. *Lethal Temperatures in Integrated Pest Management*. pp. 7. Westview Press.

Dingley F, Maynard Smith J. 1968. Temperature acclimatization in the absence of

protein synthesis of *Drosophila subobscura*. *Journal of Insect Physiology* 14: 1185-1194.

Evgenev MB, Braude-Zolotareva TY, Titarenko EA, Levin AV, Denisenko ON, Ulmasov K, Karaev K. 1987. Heat shock response in *Bombyx mori* cells infected by nuclear polyhedrosis virus. *Molecular Genetics* 215: 322-325.

Feder ME. 1996. Ecological and evolutionary physiology of stress proteins and the stress response: the *Drosophila melanogaster* model. In: Johnston IA, Bennett AF, Editors. *Animals and Temperature: Phenotypic and Evolutionary Adaptation to Temperature*. pp. 79-102. Cambridge University Press.

Feder ME, Cartaño NV, Milos L, Krebs RA, Lindquist SL. 1996. Effect of engineering hsp70 copy number on hsp70 expression and tolerance of ecologically relevant heat shock in larvae and pupae of *Drosophila melanogaster*. *Journal of Experimental Biology* 199: 410 1837-1844.

Gilchrist GW, Huey, RB. 1999. The direct response of *Drosophila melanogaster* to selection on knock-down temperature. *Heredity* 83: 15-29.

Hsieh FK, Yu SJ, Su SY, Peng SJ. 1995. Studies on the thermotolerance of the silkworm (*Bombyx mori*). *Chinese Journal Entomology* 15: 91-101.

Joy O, Gopinathan KP. 1995. Heat shock response in mulberry silkworm races with different thermo tolerances. *Journal of Biosciences* 20: 499-513.

Kato M, Nagayasu K, Ninagi O, Hara W, Watanabe A. 1989. Studies on resistance of the silkworm, *Bombyx mori* L. for high

temperature. *Proceedings of the 6th International Congress of SABRAO (II)* pp. 953-956.

Kevin CK. 2002. Molecular Biology of Thermoregulation Invited Review: Heat shock proteins: modifying factors in physiological stress responses and acquired thermotolerance. *Journal of Applied Physiology* 92: 2177-2186.

Kiuchi T, Aoki F, Nagata M. 2007. High temperature causes arrest of cell cycle in G2 phase in *BmN* cells derived from the silkworm, *Bombyx mori*. *Physiological Entomology* 32: 212-218.

Koundinya PR, Kumaresan P, Sinha RK, Thangavelu K. 2003. Screening of promising germplasm of polyvoltine silkworm (*Bombyx mori* L.) for thermotolerance. *Indian Journal of Sericulture* 42: 67-70.

Krishnaswami S, Sriharan TP, Asan M. 1971. Ecological studies on silkworm rearing to prevent crop losses in adverse seasons in West Bengal. *Indian Journal of Sericulture* 10: 72-78.

Krishnaswami S, Narasimhanna MN, Surayanarayana SK, Kumararaj S. 1973. *Manual on sericulture. Silkworm rearing. 2:* 54-88. UN Food and Agriculture Organisation, Rome

Kumar NS, Kishore Kumar CM, Basavaraju HK, Mal Reddy N, Ramesh Babu M, Datta RK. 1989. Comparative performance of robust and productive bivoltine hybrids of *Bombyx mori* L. under high temperature conditions. *Sericologia* 39: 567-571.

Kumar NS, Yamamoto T, Basavaraja HK, Datta RK. 2001. Studies on the effect of high

temperature on F1 hybrids between polyvoltines and bivoltine silkworm races of *Bombyx mori* L. *International Journal of Industrial Entomology* 2: 123-127.

Kumar NS, Basavaraja HK, Kishor Kumar CM, Mal Reddy N, Datta RK. 2002. On the breeding of "CSR18 x CSR19"- A robust bivoltine hybrid of silkworm, *Bombyx mori* L. for the tropics. *International Journal of Industrial Entomology* 5: 155-162.

Kumar NS, Basavaraja HK, Mal Reddy N, Dandin SB. 2003. Effect of high temperature and high humidity on the quantitative traits of parents, foundation crosses, single and double hybrids of bivoltine silkworm, *Bombyx mori* L. *International Journal of Industrial Entomology* 6: 197-202.

Moghaddam SHH, Du X, Li J, Cao J, Zhong BB, Chen YY. 2008. Proteome analysis on differentially expressed proteins of the fat body of two silkworm breeds, *Bombyx mori*, exposed to heat shock exposure. *Biotechnology and Bioprocess Engineering* 13: 624-631.

Pillai VS, Krishnaswami S. 1980. Effect of high temperature on the survival rate, cocoon quality and fecundity of *Bombyx mori* L. In: *Sericulture Symposium and Seminar*. pp. 141-148. Tamil Nadu Agriculture University.

Pillai VS, Krishnaswami S. 1987. Adoptability of silkworm *Bombyx mori* (L) to tropical conditions: III. Studies on the effect of high temperature during later development stages of silkworm. *Indian Journal of Sericulture* 26: 63-71.

Ramesha C, Seshagiri SV, Rao CGP. 2009. Evaluation and identification of superior polyvoltine crossbreeds of mulberry

silkworm, *Bombyx mori* L. *Journal of Entomology* 6: 179-188.

Ramesha C, Anuradha CM, Lakshmi H, Sugnana Kumari S, Seshagiri SV, Goel AK, Suresh Kumar C. 2010. Nutrigenetic traits analysis for identification of nutritionally efficient silkworm germplasm breeds. *Biotechnology* 9: 131-140.

Reddy PL, Sankar Naik S, Sivarami Reddy N. 2002. Implications of temperature and humidity on pupation patterns in the silkworm, *Bombyx mori* L. *International Journal of Industrial Entomology* 5: 67-71.

Shirota T. 1992. Selection of healthy silkworm strain through high temperature rearing of fifth instar larvae. *Reports of the silk Science Research Institute* 40: 33-40.

Srivastava PP, Kar PK, Awasthi AK, Raje Urs S. 2007. Identification and association of ISSR markers for thermal stress in polyvoltine silkworm *Bombyx mori*. *Russian Journal of Genetics* 43: 858-864.

Tazima Y. 1984. Effect of dosage rate and fractionated delivery of ionizing radiation on mutataion induction in silkworm spermatogenesis, problems of threshold in chemical mutagenesis. *The Environmental Mutagen Society of Japan* 169-173.

Tazima Y, Ohnuma A. 1995. Preliminary experiments on the breeding procedure for synthesizing a high temperature resistant commercial strain of the silkworm, *Bombyx mori* L. *Silk Science Research Institute Japan* 43: 1-16.

Thaigarajan, VSK, Bhargava M, Ramesh B, and Nagaraju B.1993. Differences in seasonal performance of 26 strains of silkworm

*Bombyx mori* (Bombycidae) *Journal of Lepidopteron Society* 47: 321-337.

Ueda S, Lizuka H.1962. Studies on the effects of rearing temperature affecting the health of silkworm larvae and upon the quality of cocoons-1 Effect of temperature in each instar. *Acta Sericologia Japan* 41: 6-21.

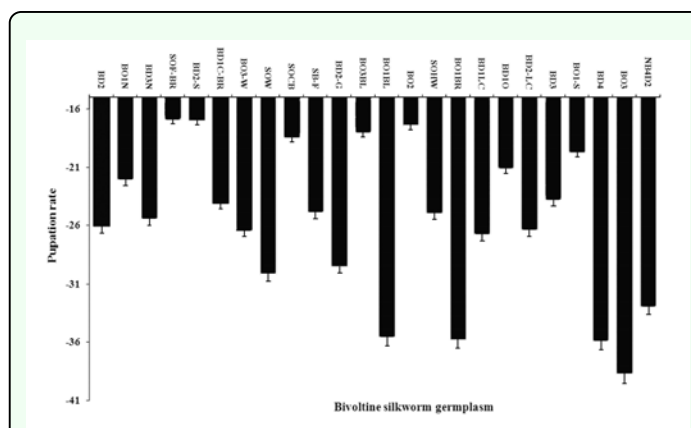
Vasudha B, Chavadi, Aparna H, Sosalegowda, Manjunatha MH. 2006. Impact of heat shock on heat shock proteins expression, biological and commercial traits of *Bombyx mori*. *Insect Science* 13: 243.

Wu DJ, Hou RF. 1993. The relation ship between thermotolerancy and heat stable esterase in the silkworm *Bombyx mori* L. (Lepidoptera: Bomycidae) *Applied Entomology and Zoology* 28: 371-377.

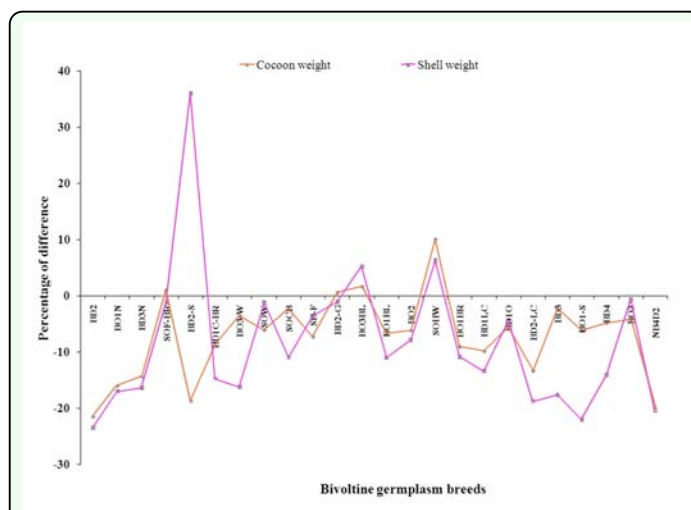
Yokoyama T. 1962. *Synthesized science of sericulture*. Central Silk Board.

Zhao Y, Chen K, He S. 2007. Key principles for breeding spring and autumn using silkworm varieties: from our experience of breeding 873×874. *Caspian Journal of Environmental Science* 5: 57-61.

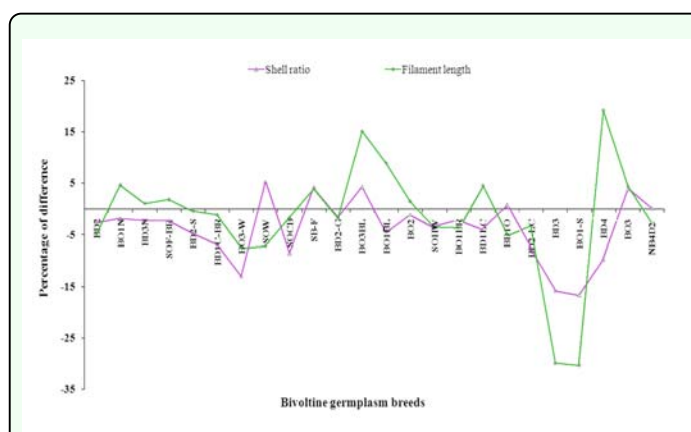
Zhang YH, Xu AY, Wei YD, Li MW, Hou CX, Zhang GZ. 2002. Studies on feeding habits of silkworm germplasm resources for artificial diet without mulberry. *Acta Sericologica Sinica* 28: 333-336.



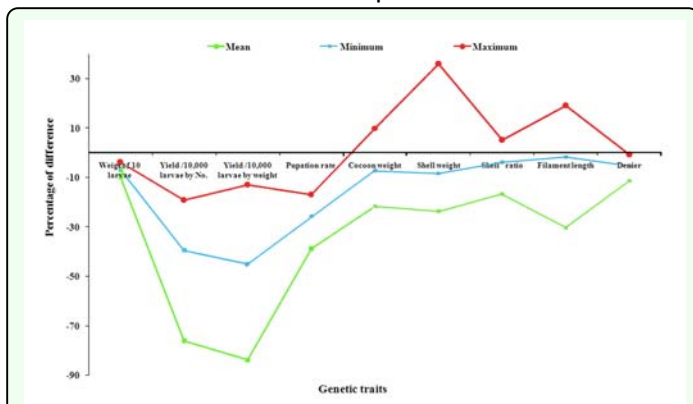
**Figure 1.** Pupaion rate difference in high temperature treated groups. High quality figures are available online.



**Figure 2.** Cocoon and shell weight difference in high temperature treated groups over control. High quality figures are available online.



**Figure 3.** Shell ratio and filament length difference in high temperature treated groups and control. High quality figures are available online.



**Figure 4.** Percentage difference among genetic traits in high temperature treated groups over control. High quality figures are available online.