



## **Biometrics, Sexual Dimorphism and Gender Determination of Griffon Vultures *Gyps fulvus* from Crete**

Authors: Xirouchakis, Stavros M., and Poulakakis, Nikos

Source: *Ardea*, 96(1) : 91-98

Published By: Netherlands Ornithologists' Union

URL: <https://doi.org/10.5253/078.096.0110>

---

BioOne Complete ([complete.BioOne.org](https://complete.BioOne.org)) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at [www.bioone.org/terms-of-use](https://www.bioone.org/terms-of-use).

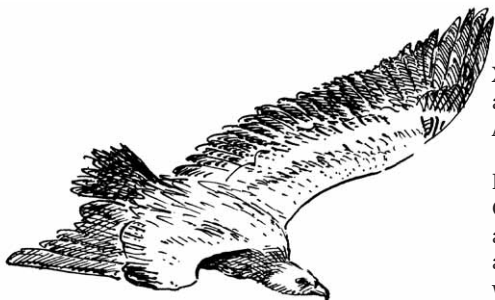
Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

---

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

# Biometrics, sexual dimorphism and gender determination of Griffon Vultures *Gyps fulvus* from Crete

Stavros M. Xirouchakis<sup>1,\*</sup> & Nikos Poulakakis<sup>1</sup>



Xirouchakis S.M. & Poulakakis N. 2008. Biometrics, sexual dimorphism and gender determination of Griffon Vultures *Gyps fulvus* from Crete. *Ardea* 96(1): 91–98.

In the present study we describe the morphometrics of 97 live Eurasian Griffon Vultures *Gyps fulvus* collected on the island of Crete (Greece) and a sex determination model on the basis of various field techniques and multivariate data analysis. We determined the sex of 49 individuals with sex-specific DNA markers and polymerase chain reaction using DNA extracted from blood samples. Adult birds were generally larger than young ones with the wing chord being the only significantly different character. Male Griffons were smaller than females for most body characters and significantly larger (c. 3–5%) for head length, head width, bill length and bill length including the cere. A stepwise discriminant function analysis showed a high predictive power in gender determination by classifying 94.1% of the individuals of known sex that was not reduced by a jackknife cross-validation technique.

Key words: morphometrics, sexual dimorphism, molecular sexing, *Gyps fulvus*, Crete

<sup>1</sup>Natural History Museum of Crete, Biology Department, University of Crete, P.O. Box 2208, Heraklion 71409, Crete, Greece;

\*corresponding author (sxirouch@nhmc.uoc.gr)

## INTRODUCTION

Biometrics and sex identification are of significance in avian biology. Data on body characteristics and gender differences have been used in a range of ecological or behavioural studies (Hughes 1998). Moreover, mating systems and demography have serious management implications in wild and captive populations, and sex identification is therefore important (Newton 1998, Dunn *et al.* 2001).

Eurasian Griffon Vultures *Gyps fulvus*, like all vultures of the hypergenus *Gyps*, lack plumage characteristics or external features from which sexes can be identified. Cloaca examination, laparoscopy, analysis of steroid hormones and

DNA analysis, which have been used for sex determination in birds including vultures (Fry 1983, Richner 1989, Griffiths *et al.* 1998, Wink *et al.* 1998, Ito *et al.* 2003), require trained researchers and specialized equipment, and are expensive and time consuming (Palma *et al.* 2001). Therefore, being able to reliably distinguish sexes by measuring morphological traits in the field would be especially useful.

Gender determination is essential to assess sex-specific behavioural patterns (e.g. foraging strategy) or life-history traits (e.g. survival, dispersal). Although the species is widely distributed and still common among European vultures in the Mediterranean region, published accounts on its body

and sex morphology are scarce (Fernández & Fernández 1974, Cramp & Simmons 1980). The objectives of this paper were (1) to describe the morphometrics of the species and make comparisons of this island population to continental ones, (2) to test the existing molecular techniques for sex identification in vultures, and (3) to determine body measurements that best differentiate males and females.

## MATERIAL AND METHODS

### Study area and sample

Between 1998 and 2006 97 live vultures were collected from Crete (34°54'N 23°30'E – 35°41'N 26°19'E, 8261 km<sup>2</sup>). All birds were found wounded, poisoned, emaciated or swept by strong northern winds into the sea. The majority came from central and eastern parts of the island, where population density is highest and most colonies occur (Xirouchakis & Mylonas 2004). Griffons were classified into three age groups according to the plumage characteristics provided by Forsman (1999): juvenile (0–1 yrs), immature (2–4 yrs) and adult (>5 yrs). After having been marked with metal rings from the Hellenic Ringing Scheme and with plastic engraved rings (Pro-Touch Engraving), birds were sent to wildlife rehabilitation centres. Normally the recovery period lasted from a few months to one year. Body measurements and blood samples were taken a few hours prior to the release in the wild.

### Molecular sexing

In 2002–2006 a small blood sample (0.5 ml) was taken from each vulture with an insulin syringe from the brachial vein ( $n = 55$ ). The blood was transferred to capillary tubes with 95% ethanol and stored at  $-20^{\circ}\text{C}$ . DNA was extracted from 200  $\mu\text{l}$  of blood using the Macherey-Nagel Kit (Germany) following the manufacturer's instructions. Contamination with modern DNA or Polymerase chain reaction (PCR) products was ruled out by including two extraction blanks in every extraction round. For DNA sexing we used the method

described by Griffiths *et al.* (1998) and Conway *et al.* (2004) based on two conserved CHD (chromohelicase-DNA-binding) genes that are located on the avian sex chromosomes of all birds.

### Morphological measurements

The following biometric data were taken, using standard measurements as described for vultures (Mendelssohn *et al.* 1989, Mundy *et al.* 1992): 1) wing chord (WC), from the carpal joint of the bent wing to the tip of the longest straightened primary; 2) wing span (WS), the distance between the extended wing tips by keeping the bird on its back with the wings fully outstretched; 3) tail length (TL), from the insertion of the central rectrices to their tips; 4) tarsus length (TL), the distance from the tarsometatarsal joint to the articulation of the middle toe; 5) middle toe (MTO), from the first scale of the extended toe to the base of its claw; 6) middle talon (MTA), the distance between the base of the claw to its tip; 7) head length (HL), from the supraoccipital to the tip of the bill; 8) head width (HW), distance between the widest points in the auricular patches behind the eyes; 9) bill length (BL), from the tip of the culmen to its junction to the cere; 10) bill plus cere length (BCL), from the bill tip to the edge of implantation of feathers; 11) bill width (BW), from the junction of the tomium of the upper jaw to the cere at both sides of the culmen; 12) bill depth (BD), dorso-ventral distance at the nostrils, and 13) body mass (W).

Wing span was measured with a tape meter (accuracy 0.5 cm). Wing chord and tail length were measured with a steel stopped ruler to the nearest millimetre. The remaining measurements were taken to the nearest 0.1 mm by using a vernier caliper. Body mass was determined with a pesola 15-kg spring balance to the nearest 0.1 kg. Vultures without physical injuries were weighed before being dispatched to rehabilitation centres, recovered birds again after having fasted for 2–3 days prior to release.

Two aerodynamic statistics were estimated (Pennycuik 1972), namely wing loading (weight/wing area) expressed as  $\text{kg m}^{-2}$  and aspect ratio ( $\text{wingspan}^2/\text{wing area}$ ). Wing area was calculated

as the average wingspan divided by average wing width (Mendelsohn *et al.* 1989, Mundy *et al.* 1992). The latter character was measured from a sample of 14 alive (8 males and 6 females) and 5 dead specimens that were all immature. All measurements were made by the same person to minimize variability, and are presented as means and standard deviations.

### Statistical analysis

Data were tested for normality by using the Shapiro-Wilk's statistic, and homogeneity of variances with the Levene's test. A univariate analysis (ANOVA) was applied for differences in body measurements between age groups and sexes, and Tukey tests and *t*-tests on a post-hoc analysis, respectively. When the assumption of normality was violated, comparisons were made with Kruskal-Wallis and Mann-Whitney tests (Zar 1996). A multiple analysis of variance (MANOVA) was performed on both age and sex in order to investigate their main effect and interaction on body characters (Hair *et al.* 1998). For sex determination, frequency distributions, although overlapping, indicated that some characters might discriminate birds. The percent of sexual dimorphism in each measurement was taken as  $\frac{\bar{x}_m - \bar{x}_f}{\bar{x}_m}$  where  $\bar{x}_m$  and  $\bar{x}_f$  were the mean values of males and females respectively. Gender determination was pursued by applying Discriminant Function Analysis (DFA; Klecka 1980) with sex being the categorical variable. For selection of the dependent variable (body measurements) we used Wilks' lambda statistic. The main criteria established for the application of the method were: a) normality of the data, b) linearity of inter-variable relationships (inspection of scatterplot matrix), c) lack of multicollinearity (inter-variable correlation coefficients  $r < 0.6$ ) and d) homoscedasticity (equality of the variance/covariance matrices, Box's M statistic). Prior probabilities in the groups formed in the classification were determined by the observed sample sizes.

The DFA with all variables combined detected the most informative ones and predicted membership into two mutually exclusive groups. It created a regression equation (discriminant function) that

provided individual functions (D scores) where a cut-point between male-female mean scores (the weighted average of group centroids) distinguished birds by gender. The effectiveness of the method was assessed by the proportion of birds of known sex that were correctly classified using all individuals in the analysis. A jackknife procedure (Sokal & Rohlf 1995) was also performed where each bird was singly removed from the analysis and classified using a separate function derived from the remainder of the data. This cross-validation technique was considered a good indicator of the DFA accuracy since it works better for small sample sizes. All analyses were made at a 0.05 level of significance by using SPSS 14.0 software (Norusis 1989).

## RESULTS

Descriptive statistics for all birds sampled are presented in Table 1. All variables were normally distributed (Shapiro-Wilk's statistic  $P > 0.05$ ), with the exception of body mass which was discarded from the discriminant function analysis. The weight of Griffons differed between age groups (Kruskal-Wallis  $\chi^2 = 8.6$ ,  $P = 0.014$ ), but not between the sexes (Mann-Whitney  $U = 294$ ,  $P = 0.634$ ) although females were slightly heavier than males. Extreme values below 6 kg (with a minimum of 4 kg), apparently the threshold for a Griffon to become grounded, were excluded from the comparisons.

Regarding age, 41 vultures were classified as juveniles, 46 as immatures and 10 as adults. Means for body measurements did not differ between age groups apart from wing chord that was significantly larger in adult than in juvenile and immature birds (Tukey test  $P = 0.003$ ). However, non-adult birds (pooled data) were smaller than adults for eight out of 12 morphometrics variables examined (Table 1). Although the small sample size might have produced some bias, adult birds were on average smaller in tail, middle talon, bill width and bill depth. By examining the coefficient of variation we concluded that

**Table 1.** Descriptive statistics of body measures of Griffon Vultures in Crete. Units in mm unless otherwise mentioned. Variable that differs among age groups (*F*-test) in bold.

Variable	<i>n</i>	Pooled $\bar{x} \pm SD$	Range	Juvenile $\bar{x} \pm SD$ ( <i>n</i> )	Immature $\bar{x} \pm SD$ ( <i>n</i> )	Adult $\bar{x} \pm SD$ ( <i>n</i> )	<i>F</i> -value	<i>P</i>
<b>Wing chord, cm</b>	97	69.9 ± 2.65	64–75.3	70 ± 2.56 (41)	69.1 ± 2.47 (46)	72.9 ± 1.58 (10)	9.74	< 0.001
Wingspan, cm	97	252.5 ± 9.13	230–272	250.3 ± 8.55 (41)	253.6 ± 8.57 (46)	256.2 ± 12.35 (10)	2.45	0.09
Tarsus	97	118.3 ± 7.93	99.5–135.04	117.9 ± 7.99 (41)	118.4 ± 7.09 (46)	119.2 ± 11.57 (10)	0.13	0.88
Tail, cm	97	30.4 ± 2.80	24–38	30.3 ± 2.63 (41)	30.7 ± 2.88 (46)	29.3 ± 3.12 (10)	1.08	0.34
Middle toe	97	107.9 ± 3.83	100–116.8	107.7 ± 3.10 (41)	107.7 ± 4.27 (46)	110.6 ± 3.86 (10)	2.63	0.07
Middle talon	97	35.5 ± 2.73	27.8–43	35.6 ± 2.42 (41)	35.6 ± 3.01 (46)	35 ± 2.78 (10)	0.20	0.82
Head length	89	142.8 ± 5.07	133.3–153.8	142.8 ± 5.33 (35)	142.5 ± 5.10 (44)	144 ± 4.24 (10)	0.36	0.70
Head width	59	61.1 ± 2.54	56.9–66.8	60.6 ± 2.83 (16)	61.2 ± 2.45 (37)	62.2 ± 2.30 (6)	0.97	0.38
Bill length	91	52.6 ± 2.58	45.9–61.1	52.1 ± 2.44 (35)	52.6 ± 2.74 (46)	54 ± 1.84 (10)	2.12	0.13
Bill-cere length	74	73.5 ± 2.5	67–78.6	73.7 ± 2.78 (25)	73.4 ± 2.37 (40)	73.9 ± 0.53 (9)	0.20	0.82
Bill width	75	24.8 ± 1.84	21.4–29	24.9 ± 1.89 (27)	24.8 ± 1.82 (39)	24.4 ± 1.95 (9)	0.25	0.78
Bill depth	74	35.5 ± 1.94	30.8–40	35 ± 1.96 (26)	35.9 ± 1.70 (39)	35.4 ± 2.66 (9)	1.79	0.17
Weight, kg	92	7.44 ± 0.72	6–9	7.16 ± 0.68 (37)	7.66 ± 0.55 (45)	7.47 ± 1.13 (10)		
Wing area, m <sup>2</sup>	19		0.88–0.89		0.88 ± 0.01			
Wing loading, kg m <sup>-2</sup>	19		8–9.5		8.69 ± 0.38			
Aspect ratio	19		7.2–7.3		7.23 ± 0.04			

**Table 2.** Sexual dimorphism, univariate comparisons and stepwise discriminant function analysis of morphometric measurements of male and female Griffon Vultures in Crete. See Table 1 for units. Variables that differ between sexes (by Wilks' lambda) in bold.

Variable	Male ( <i>n</i> = 29) $\bar{x} \pm SD$ (range)	Female ( <i>n</i> = 22) $\bar{x} \pm SD$ (range)	% dimorphism	Wilks' lambda	<i>F</i> -value	<i>P</i>
Wing chord	69.5 ± 2.72 (64–75)	70.3 ± 2.84 (65–75)	-1.3	0.976	1.23	0.27
Wingspan	253.4 ± 7.80 (238–270)	253.9 ± 10.58 (230–272)	-0.2	0.999	0.03	0.87
Tarsus	120.5 ± 9.67 (99.5–135.04)	117.2 ± 8.42 (101–134)	2.8	0.967	1.69	0.20
Tail	30 ± 3.02 (24–35)	31 ± 2.79 (27.2–38)	-3.4	0.969	1.55	0.22
Middle toe	108.2 ± 4.53 (101.3–116.8)	107.6 ± 3.41 (100.4–113.8)	0.6	0.994	0.27	0.60
Middle talon	35.2 ± 2.24 (30.2–39.1)	36.5 ± 3.74 (27.8–43)	-3.7	0.952	2.46	0.12
<b>Head length</b>	146.1 ± 3.92 (139.5–153.8)	138.9 ± 2.50 (134.3–143.2)	5	0.461	57.35	<0.001
<b>Head width</b>	62.1 ± 2.14 (58.4–66.1)	59.8 ± 2.51 (57.3–66.8)	3.7	0.799	12.29	0.001
<b>Bill length</b>	54.1 ± 2.18 (49.9–61.1)	51.4 ± 1.81 (45.9–55)	5	0.687	22.35	<0.001
<b>Bill-cere length</b>	74.7 ± 1.93 (71.5–78.6)	72.6 ± 1.77 (69.1–76)	2.8	0.754	16.01	<0.001
Bill width	24.8 ± 1.98 (21.4–29)	25.3 ± 1.86 (21.8–29)	-1.8	0.986	0.70	0.40
Bill depth	35.4 ± 1.64 (30.8–38.6)	36 ± 1.90 (32.5–40)	-1.8	0.969	1.56	0.22
Body mass	7.648 ± 0.69 (6.5–9)	7.741 ± 0.42 (6.2–9)	-0.012			
Wing area	0.89 ± 0.004 (0.8–0.89, <i>n</i> = 8)	0.89 ± 0.01 (0.8–0.89, <i>n</i> = 6)	0			
Wing loading	8.7 ± 0.21 (8.5–9.1, <i>n</i> = 8)	9.04 ± 0.28 (8.8–9.5, <i>n</i> = 6)	-3.9			
Aspect ratio	7.24 ± 0.03 (7.2–7.3, <i>n</i> = 8)	7.26 ± 0.04 (7.2–7.3, <i>n</i> = 6)	-0.3			

relative variability was marginally significant (homogeneity Levene's test,  $P = 0.053$ ), increasing with age for wingspan, tarsus length, tail length and bill depth, and decreasing for wing chord and bill length.

Molecular techniques (i.e. P2 and P8 sexing methods) proved successful in gender identification. We determined the sex of 49 individuals (27 males, 22 females) out of a total of 55. Males showed a single band at 379bp (CHD-Z) on the polyacrylamide gel, while the females exhibited two bands (379bp, CHD-Z) and (388bp, CHD-W), both of them readily discernible. The identifying fragments of CHD gene were sequenced to confirm their size. GenBank accession numbers to the nucleotide sequence of these regions are EU430640 (CHD-W) and EU430641 (CHD-Z). Two additional birds were sexed as males by gonad inspection after dissection.

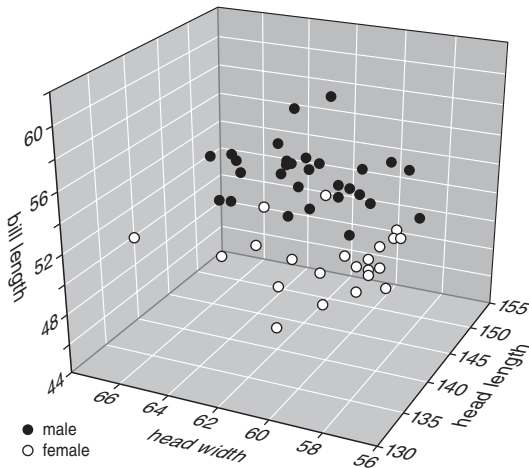
Males were smaller than females for most body measurements, including wing loading and aspect ratio, though the degree of sexual dimorphism differed significantly only between head and bill dimensions ( $F$ -tests, all  $P < 0.05$ , Table 2). In particular females were larger for all physical characters, apart from tarsus length and middle toe length, but male birds were significantly larger than females (c. 3–5%) for length and width of the head as well as length of the bill up to the cere or to the edge of the feathering (Table 2). The effect of age on sexual dimorphism (MANOVA, Wilks' Lambda = 0.399,  $F = 1.65$ ,  $P = 0.055$ , interaction: Wilks' Lambda = 0.421,  $F = 1.53$ ,  $P = 0.087$ ) was not significant, therefore we pooled data of all age classes and performed a discriminant function analysis with sex as independent variable. A linear

discriminant function analysis was used since all relevant assumptions were met, i.e. linear relationship between the variables, no inter-variable correlations ( $r < 0.5$ ) and homoscedasticity (Box's M statistic = 15.1,  $F = 1.38$ ,  $P = 0.18$ ). The Wilks' Lambda criterion indicated that head length, head width, bill length and bill-cere length were the best physical characters in discriminating gender (Table 2). The application of DFA where all four variables were entered simultaneously, exhibited a success rate of 96.6% and 95.5% in correctly predicting males and females respectively (eigenvalue = 2.92,  $\chi^2 = 0.255$ ,  $df = 4$ ,  $P < 0.001$ ). On the whole sex was accurately predicted for 96.1% of the sample, although it dropped to 94.1% when the jackknife cross-validation technique was performed. However, a stepwise procedure showed that sex discrimination was possible by using head length, head width and bill length (eigenvalue = 2.89,  $\chi^2 = 0.257$ ,  $df = 3$ ,  $P < 0.001$ ) with an overall accuracy of 94.1% (Table 3).

Both unstandardized discriminant functions were similarly effective, as shown by the cross-validated values (Table 3). However, we considered the discriminant formula with the fewer variables to be the most useful (Fig. 1), because the cross-validation technique did not reduce its predictive ability even if prior probabilities in the DFA were taken of equal values (Table 3, correctly classified cases,  $D_2$ : 96.6% of males, 90.9% of females, 94.1% overall vs.  $D_1$ : 89.7% of males, 90.0% of females, 90.2% overall). For the simpler discriminant function the group centroids were 1.45 for males and -1.912 for females, thus the cutting score 0.31 could be used as a gender index (males:  $D$  scores  $> 0.31$ , females:  $D$  scores  $< 0.31$ ).

**Table 3.** Discriminant functions (four and three variables) for gender determination of Griffon Vultures in Crete. HL = head length, HW = head width, BL = bill length, BCL = bill-cere length.

Discriminant function	Group centroids		Selftest (%)	Cross-validated (%)
	Males	Females		
$D1 = 0.26 \text{ HL} + 0.17 \text{ HW} + 0.34 \text{ BL} + 0.06 \text{ BCL} - 71.4$	1.459	-1.924	96.1	94.1
$D2 = 0.27 \text{ HL} + 0.19 \text{ HW} + 0.37 \text{ BL} - 69.12$	1.450	-1.912	94.1	94.1



**Figure 1.** Head length, head width and bill length (all in mm) are effective measurements for gender determination of Griffon Vultures in Crete.

## DISCUSSION

Regarding age-related biometrics, relevant conclusions should be drawn with caution as the sample size of adult birds was small. Nevertheless, excluding characters of significant sexual dimorphism, adult griffons were larger than immatures in all morphological measurements apart from tail length, probably a feature that serves manoeuvrability of flight-inexperienced birds. The length of the middle talon was found to be smaller in adult birds but this difference could have been caused by some degree of wear in a few individuals. The negative trend in the variation of measurements with regard to age should be attributed to the variable body condition of fledglings and first-year birds. However, an increase in wing chord and a decrease of tail length with age seems rather common among medium-sized raptors and eagles (Mundy 1982, Bortolotti 1984, Ferrer & de le Court 1992, Donohue & Dufty 2006).

In general raptors exhibit reverse sexual dimorphism where females are generally larger than males (Newton 1979). In our case the differences

observed, with female vultures being marginally larger than males for most linear measurements as well as weight, were in line with the existing literature (Cramp & Simmons 1980, Donázar 1993). More specifically, our results on sex-specific discrepancies in head and bill dimension corresponded with Fernández & Fernández (1974), who observed morphological differences in head width, bill length and width of the lower mandible of male and female Griffons in a sample of 16 pairs. The fact that head and bill dimensions were not influenced by age indicates that inter-sexual differences between these characters build up quite early, probably already during the nestling stage.

Head and bill length have a strong discriminant power to distinguish males from females. These 'skeletal' measurements have the advantage not to vary in relation to factors such as food availability, season, phase of the breeding cycle, moult or wear. Moreover, they are consistent and routinely measured in morphometrical studies in contrast to other body characters that are effective in adult gender discrimination but not traditionally collected in the field e.g. footpad length, forearm length or tarsus length (Edwards & Kochert 1986, Ferrer & de le Court 1992).

The present field method for gender determination of Griffon Vultures is rapid and simple, and cheaper than molecular techniques. Its applicability corresponds to live birds and possibly to study skins. Geographic variation may affect the sex discriminant coefficients of the function we developed. However, given the long-distance movements of the species within its distribution range (Bernis 1983, Berthold *et al.* 1991, Sušić 2000, Bahat *et al.* 2001), this is unlikely.

## ACKNOWLEDGEMENTS

We thank the Hellenic Wildlife Rehabilitation Centre at Aegina, the staff of the vertebrate section of the Natural History Museum of Crete and J.A. Donázar, A. Margalida, R.G. Bijlsma and two anonymous referees for their comments on an earlier draft of the manuscript. The study was supported by a PYTHAGORAS II research grant to SMX from the Hellenic Ministry of Education.

## REFERENCES

- Bahat O., Hatzoffe O. Kaplan A. & Woodley B. 2001. Foraging range and movements of Griffon vultures (*Gyps fulvus*) in Israel, as determined by satellite tracking. Fourth Eurasian congress on raptors. Doñana Biol. Station – Raptor Research Foundation, Seville, pp. 11–12.
- Berthold P., Griesinger J., Nowak E. & Querner U. 1991. Satelliten-Telemetrik eines Gänsegeier (*Gyps fulvus*) in Spanien. *J. Ornithol.* 132: 327–329.
- Bernis F. 1983. Migration of the Common Griffon Vulture in the Western Palearctic. In: Wilbur S.R. & Jackson J.A. (eds) *Vulture Biology and management*. University of California Press, Berkeley, pp. 185–196.
- Bortolotti G.R. 1984. Age and sex variation in Golden Eagles. *J. Field Ornithol.* 55: 54–66.
- Conway W.C., Wickliffe J.K., Hoffmann F.G., Baker R.J. & Smith L.M. 2004. An improved PCR-based method for gender identification in birds. *Occ. Papers Museum of Texas Tech.* University 239: 1–7.
- Cramp S. & Simmons K.E.L. (eds) 1980. *The Birds of Western Palearctic*. Vol. II. Oxford University Press, Oxford.
- Donázar J.A. 1993. *Los Buitres Ibericos*, Biología y Conservación. Reyero JM, Madrid.
- Donohue K.C. & Dufty A.M. Jr. 2006. Sex determination of Red-tailed Hawks (*Buteo jamaicanensis calurus*) using DNA analysis and morphometrics. *J. Field Ornithol.* 77: 74–79.
- Dunn P.O., Whittingham L.A. & Pitcher T.E. 2001. Mating systems, sperm competition, and the evolution of sexual dimorphism in birds. *Evolution* 51: 161–175.
- Edwards T.C. Jr. & Kochert M. 1986. Use of body weight and length of footpad as predictors of sex in Golden Eagles. *J. Field Ornithol.* 57: 317–319.
- Fernández J.A. & Fernández J. 1974. Sobre sexo, mecanismos y proceso de reproducción en el buitre leonado (*Gyps fulvus*). *Doñana Acta Vertebrata* 1: 109–119.
- Ferrer M. & de le Court C. 1992. Sex identification in the Spanish Imperial Eagle. *J. Field Ornithol.* 63: 359–364.
- Forsman D. 1999. *The raptors of Europe and the Middle East*. Posyer, London.
- Fry D.M. 1983. Techniques for sexing monomorphic vultures. In: Wilbur S.R. & Jackson J.A. (eds) *Vulture Biology and Management*. University of California Press, Berkeley, pp. 356–374.
- Griffiths R., Double M.C., Orr K. & Dawson R.J.G. 1998. DNA test to sex most birds. *Mol. Ecol.* 7: 1071–1075.
- Hair J.F., Anderson R.E., Tatham R.L. & Black W.C. 1998. *Multivariate data analysis*. Prentice-Hall, New Jersey.
- Hughes C. 1998. Integrating molecular techniques with field methods in studies of social behaviour: a revolution results. *Ecology* 79: 383–399.
- Ito H., Sudo-Yamaji A., Abe M., Murase T. & Tsubota T. 2003. Sex identification by alternative polymerase chain reaction methods in Falconiformes. *Zool. Science* 20: 339–344.
- Klecka W.R. 1980. *Discriminant analysis. Quantitative Applications in the Social Sciences Series*, No. 19. Sage Publications, Thousand Oaks, CA.
- Mendelsohn J.M., Kemp A.C., Biggs H.C., Biggs R. & Brown C.J. 1989. Wing areas, wing loadings and wing spans of 66 species of African raptors. *Ostrich* 60: 35–42.
- Mundy P. 1982. *The comparative biology of Southern African vultures*. Vulture Study Group, Johannesburg.
- Mundy P., Butchart D., Ledger J. & Piper S. 1992. *The Vultures of Africa*. Academic Press, London.
- Newton I. 1979. *Population ecology of raptors*. Buteo Books, Vermillion.
- Newton I. 1998. *Population limitation in birds*. Academic Press, London.
- Norusis, M. 1989. *SPSS for Windows. Advanced Statistics*. Chicago University Press, Chicago.
- Palma L., Mira S., Cardia P., Beja P., Guillemaud T., Ferrand N., Cancela M.L. & da Fonseca L. 2001. Sexing Bonelli's eagle nestlings: morphometrics versus molecular techniques. *J. Raptor Res.* 35: 187–193.
- Pennyquick C.J. 1972. Soaring behaviour and performance of some East-African birds, observed from a motor-glider. *Ibis* 114: 178:218.
- Richner H. 1989. Avian laparoscopy as a field technique for sexing birds and an assessment of its effects on wild birds. *J. Field Ornithol.* 60: 137–142.
- Sokal R.R. & Rohlf F.J. 1995. *Biometry: the principles and practice of statistics in biological research*. Third edition. Freeman, New York.
- Sušić G. 2000. Regular long-distance migration of Eurasian Griffon *Gyps fulvus*. In: Chancellor R.D. & Meyburg B.-U. (eds) *Raptors at Risk*. WWGBP/Hancock House, Berlin, pp. 225–230.
- Wink M., Sauer-Gürth H., Martinez F., Doval G., Blanco G. & Hatzofe O. 1998. The use of (GACA) PCR to sex Old World vultures (Aves: Accipitridae). *Mol. Ecol.* 7: 779–782.
- Xirouchakis S.M. & Mylonas M. 2004. Griffon Vulture (*Gyps fulvus*) distribution and density in Crete. *Israel J. Zool.* 50: 341–354.
- Zar J. 1996. *Biostatistical analysis*. Third edition. Prentice Hall, New Jersey.



## SAMENVATTING

Door middel van inspectie van de cloaca, laparoscopie en een analyse van steroïde hormonen en DNA is het mogelijk de sekse van Vale Gieren *Gyps fulvus* te bepalen. Deze technieken vergen echter veel ervaring, kennis en gespecialiseerde apparatuur. Ze zijn bovendien niet goedkoop. De onderhavige studie probeert morfologische maten te identificeren waarmee de geslachten van Vale Gieren uit elkaar kunnen worden gehouden. Onderscheid naar geslacht is wenselijk, omdat veel biologische parameters – zoals dispersie, overleving en gedrag – verschillen naar sekse. Om inzicht in de populatiedynamiek van een lokale populatie te krijgen, zoals in dit geval bij Vale Gieren op Cyprus, is seksebepaling van gevangen en gevonden vogels een eerste vereiste. Dat klinkt eenvoudiger dan het is, omdat Vale Gieren – in tegenstelling tot andere roofvogelsoorten – slechts een geringe seksuele dimorfie kennen. Verder is onbekend of de eilandpopulatie op Cyprus afwijkt van die op het vasteland van Europa. Het onderzoek werd verricht aan 97 gieren die tussen 1998 en 2006 in vogelasiels terecht waren gekomen, voornamelijk als juveniele (41) en onvolwassen

(46) vogels. Slechts 10 vogels werden als adult aangemerkt. Van alle vogels werden twaalf lengtematen bepaald, verdeeld over vleugel, staart, poot en kop, daarnaast gewicht en afgeleiden als vleugelbelasting en vleugelslankheid. Een deel van de vogels werd moleculair op geslacht gebracht. Voor de meeste lichaamsmaten waren mannen kleiner dan vrouwen. Significante verschillen tussen de seksen werden gevonden in koplengte, kopbreedte en snavelengte (zowel met als zonder washuid), waarbij mannen 3–5% groter waren dan vrouwen. Kopmaten zijn 'hard' (immers betrekking hebbend op bot), vertonen geen seizoenvariatie, zijn niet onderhevig aan leeftijdsafhankelijke variaties, en zijn beter reproduceerbaar dan 'zachte' maten als vleugel, staart en tarsus. Door gebruik te maken van snavelengte, koplengte en kopbreedte kon bijna 95% van de Vale Gieren foutloos op geslacht worden gebracht. Daarmee is het een goed en goedkoop alternatief voor meer geavanceerde methoden.

(RGB)

*Corresponding editor: Rob G. Bijlsma*

*Received 30 August 2007; accepted 27 February 2008*