

# THE PREVALENCE OF ANTIBODY TO CONTAGIOUS CAPRINE PLEUROPNEUMONIA (Mycoplasma strain F38) IN SOME WILD HERBIVORES AND CAMELS IN KENYA 1

Authors: PALING, R. W., MACOWAN, K. J., and KARSTAD, L.

Source: Journal of Wildlife Diseases, 14(3): 305-308

Published By: Wildlife Disease Association

URL: https://doi.org/10.7589/0090-3558-14.3.305

The BioOne Digital Library (<u>https://bioone.org/</u>) provides worldwide distribution for more than 580 journals and eBooks from BioOne's community of over 150 nonprofit societies, research institutions, and university presses in the biological, ecological, and environmental sciences. The BioOne Digital Library encompasses the flagship aggregation BioOne Complete (<u>https://bioone.org/subscribe</u>), the BioOne Complete Archive (<u>https://bioone.org/archive</u>), and the BioOne eBooks program offerings ESA eBook Collection (<u>https://bioone.org/esa-ebooks</u>) and CSIRO Publishing BioSelect Collection (<u>https://bioone.org/csiro-ebooks</u>).

Your use of this PDF, the BioOne Digital Library, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at <u>www.bioone.org/terms-of-use</u>.

Usage of BioOne Digital Library content is strictly limited to personal, educational, and non-commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne is an innovative nonprofit that sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

## THE PREVALENCE OF ANTIBODY TO CONTAGIOUS CAPRINE PLEUROPNEUMONIA (Mycoplasma strain F38) IN SOME WILD HERBIVORES AND CAMELS IN KENYA <sup>III</sup>

R. W. PALING, K. J. MACOWAN and L. KARSTAD, Veterinary Research Laboratory, Kabete, Kenya.

Abstract: Sera of 11 species of wild herbivores were tested for antibody to Mycoplasma strain F38 which causes contagious caprine pleuropneumonia (CCPP) in Kenya. Antibodies were found in buffalo (Syncerus caffer) (32%), impala (Aepyceros melampus) (10%) and camels (Camelus dromedarius) (49%) but not in bushbuck (Tragelaphus scriptus), eland (Taurotragus oryx), Grant's gazelle (Gazella granti), kongoni (Alcelaphus buselaphus cokei), oryx (Oryx beisa), Thomson's gazelle (Gazella thomsonii), waterbuck (Kobus defassa) and wildebeest (Connochaetus taurinus).

The infection of a single Thomson's gazelle resulted in the development of antibody but no clinical disease. A goat kept in contact did not become infected.

## **INTRODUCTION**

Contagious caprine pleuropneumonia (CCPP) has been recognized in Kenya for several decades but only recently has the causal agent been isolated.<sup>4,5</sup> Three strains of Mycoplasma were isolated, F38 and G69 causing acute contagious pleuropneumonia<sup>5</sup> and F30 causing a non-contagious pleuropneumonia.4 Infection experiments were conducted with the three strains F38, G69 and F30 and the other known CCPP-associated Mycoplasma strains PG3 (National Type Culture Collection 10137), the Nigerian reference strains N  $108^1$  and Vom and M. mycoides subspecies mycoides (Gladysdale). F38 and G69 caused acute fatal fibrinous pleuropneumonia by intratracheal-endobronchial inoculation and by contact with experimentally infected animals. 3,5,6 F30 and Vom were highly pathogenic by inoculation<sup>3,4</sup> and strains PG3, IV108 and M. mycoides subspecies mycoides (Gladysdale) did not cause pleuropneumonia in goats.<sup>3</sup>

Serological tests were developed and the findings of the complement fixation and agar gel diffusion tests indicated that F38 is related to M. mycoides. In addition, the growth inhibition test indicated that F38 and G69, differed from PG3, N108, Smith,<sup>2</sup> F30, Vom and M. mycoides subspecies mycoides (Gladysdale).

Mycoplasma of the F38 strain were isolated from 21 acute cases from 14 outbreaks.<sup>5</sup> Sera from 10 species of mainly free-ranging wild herbivores and from camels (Camelus dromedarius) were screened by the complement fixation test (CFT) for antibody to Mycoplasma strain F38. This strain F38 which proved to be highly infectious for goats<sup>3,5</sup> was used to inoculate one Thomson's gazelle (Gazella thomsonii).

## MATERIALS AND METHODS

Blood was collected and sera were obtained from nine species of free-ranging

Research conducted as part of a Kenya Government project supported by the United Nations Development Program, with the Food and Agriculture Organisation as the executing agency; and the Canadian International Development Agency, through the International Development Research Centre.

wild herbivores from five districts, two species of wild herbivores in captivity (Kabete) and three species of domesticated wild herbivores and camels (Kilifi District) (Table 1). The animals were either shot, manually restrained or immobilized by the injection of a mixture of xylazine, <sup>(2)</sup> etorphine HCl <sup>(2)</sup> and acepromazine maleate <sup>(2)</sup> by projectile syringe. <sup>(2)</sup> The presence of antibody to *Mycoplasma* strain F38 was determined by CFT.<sup>4,6</sup> CFT titres of 1:32 and over were regarded as evidence of exposure (positive).

#### **EXPERIMENT**

An adult male Thomson's gazelle captured several months earlier and found free of antibody to CCPP, was inoculated by the intratracheal-endobronchial route with 20 ml of a culture of *Mycoplasma* (F38) containing over  $10^9$  colony forming units. For collection of serum and recording of body temperatures the gazelle was immobilized weekly, for 4 months, by the injection of 10 mg xylazine delivered by projectile syringe.

## RESULTS

Six of 19 (32%) buffalo (Syncerus caffer) from three areas, one of 10 (10%) impala (Aepyceros melampus) and 66 of 134 (49%) camels from two areas were positive (Table 1). The experimentally inoculated Thomson's gazelle did not develop clinical disease but serum antibody was present from day 14 to day 54 postexposure, with a peak titre of 1:128 between days 35 and 42. The goat kept in contact with the gazelle did not become infected.

## DISCUSSION

To date attempts to isolate Mycoplasma causing CCPP from wild herbivores and camels have been unsuccessful.<sup>8</sup> This is the first report of antibody to this organism in wild herbivores and camels. Shifrine et al.10 attempted to isolate M. mycoides from lymph nodes from 36 buffalo in East Africa, but failed. Significant CF antibody to M. mycoides subspecies mycoides were found in wildebeest (Connochaetes taurinus) and hippopotamus (Hippopotamus amphibius) but not in impala, eland, waterbuck, buffalo and zebra.9,10 The CFT enabled M. mycoides subspecies mycoides to be distinguished from the caprine strains.<sup>5</sup>

The prevalence of antibody to Mycoplasma strain F38 was low (4.9%) in 10 species of free-ranging herbivores from 6 districts of Kenya. Although the disease was readily transmitted by contact among goats, 5,6 wild animals except buffalo in infected areas did not have complement fixing antibodies to the causative organism and may not have a role in the epidemiology of CCPP. Following acute caprine pleuropneumonia, CF antibodies persisted in goats for at least 4 months.7 The possible involvement of the buffalo requires further study. The experimental infection of the Thomson's gazelle indicated that this species was susceptible to infection although clinical disease did not develop. For this reason Thomson's gazelle probably would not be an important source of infection for goats. The significance of antibody in camels is not clear, as the causal agent has not been identified.8 Goats on the same farms as the camels had no history of CCPP, but it is possible that a Mycoplasma identical to or closely related to strain F38 may have been present.

<sup>(2)</sup> Rompun 2%. Bayer, Leverkusen, Germany.

<sup>&</sup>lt;sup>3</sup> Large Animal Immobilon. Reckitt and Colman Ltd., Hull, England.

<sup>&</sup>lt;sup>(3)</sup> Palmer Chemical and Equipment Company. Douglasville, Georgia, USA.

<b>TABLE 1. Serce</b>	TABLE 1. Serologic survey for antibody to CCPP (Mycoplasma F38) in Kenya (1974-1977)	body to CCPF	(Mycoplas	sma F38) ii	n Kenya (	1974-1977).				
		Kabete								
SPECIES	AREA	Captivity	Kajiado	Kilifi	Laikipia	Kajiado Kilifi Laikipia Machakos Nakuru Narok total	Nakuru	Narok		positive
Buffalo	Syncerus caffer			4/4*	1/6	1/7		0/2	6/19	32%
Bushbuck	Tragelaphus scriptus	us 0/1			0/3		0/6		0/10	0%
Camel	<b>Camelus</b> dromedarius	ius		64/102	2/32				66/134	49%
Eland	Taurotragus oryx	0/5	0/11	0/12					0/28	0%0
Grant's Gazelle	Gazella granti		0/10				9/0		0/16	9%0
Impala	Aepyceros melampus	18	1/9				0/1		1/10	10%
Kongoni	Alcelaphus buselaphus cokei					0/4	9/0		0/10	9%0
Огух	Oryx beisa			0/20					0/20	0%0
Thomson's Gazelle	Gazella thomsonii				6/0		0/1		0/10	00%
Waterbuck	Kobus defassa				0/1	0/2	2/0		0/10	0%
Wildebeest	Connochaetes taurinus		0/10						0/10	06%
*The number o over the total	*The number of sera recorded "positive" in the complement fixation test (titres of 1/32 or higher) are given, as the numerator, over the total tested, as denominator.	ive" in the co or.	mplement	fixation te	st (titres o	f 1/32 or hig	(her) are	given, a	us the nu	merator,

## Acknowledgement

This paper is published with kind permission of the Director of Veterinary Services, Kenya.

## LITERATURE CITED

- COTTEW, G.S. and R.H. LEACH. 1969. Mycoplasma of cattle, sheep and goats. pp. 527-570. In: The Mycoplasmatales and L-Phase of Bacteria. L. Hayflick (ed.). North Holland, Amsterdam.
- W.A. WATSON, O. ERDAG and F. ARISOY. 1969. Mycoplasma of caprine pleuropneumonia in Turkey and their relationship to other Mycoplasma of goats and M. mycoides var. mycoides. J. Comp. Path., 79: 541-551.
- KALINER, G. and K.J. MACOWAN. 1976. The pathology of experimental and natural contagious caprine pleuropneumonia in Kenya. Zbl. Vet Med. B, 23: 652-661.
- 4. MACOWAN, K.J. 1976. A *Mycoplasma* from chronic caprine pleuropneumonia in Kenya. Trop. Anim. Hlth. Prod. 8: 20-26.
- 5. —— and J.E. MINETTE. 1976. A *Mycoplasma* from acute contagious caprine pleuropneumonia in Kenya. Trop. Anim. Hlth. Prod. 8: 91-95.
- and ——. 1977. Contact transmission of experimental contagious caprine pleuropneumonia (CCPP). Trop. Anim. Hlth. Prod. 9: 185.
- and ——. 1977. The effect of high passage Mycoplasma strain F38 on the cause of contagious caprine pleuropneumonia (CCPP). Trop. Anim. Hlth. Prod. in press.
- 8. PALING, R.W., S. WAGHELA and K.J. MACOWAN. The role of diseases in mixed farming of domesticated wild animals and domestic animals including camels on a ranch in Kenya. Patr II, Bacterial diseases. In preparation.
- 9. SHIFRINE, M. and C.H. DOMERMUTH. 1967. Contagious bovine pleuropneumonia in wildlife. Bull. epiz. Dis. Afr. 15: 319-322.
- 10. ——, S.S. STONE and C. STAAK. 1970. Contagious bovine pleuropneumonia in African buffalo (Syncerus caffer). Bull. epiz. Dis. Afr. 18: 201-205.

Received for publication 15 November 1977

308