

BRAIN CHOLINESTERASE ACTIVITY OF NESTLING GREAT EGRETS, SNOWY EGRETS AND BLACK-CROWNED NIGHT-HERONS

Authors: Custer, Thomas W., and Ohlendorf, Harry M.

Source: Journal of Wildlife Diseases, 25(3): 359-363

Published By: Wildlife Disease Association

URL: https://doi.org/10.7589/0090-3558-25.3.359

The BioOne Digital Library (https://bioone.org/) provides worldwide distribution for more than 580 journals and eBooks from BioOne's community of over 150 nonprofit societies, research institutions, and university presses in the biological, ecological, and environmental sciences. The BioOne Digital Library encompasses the flagship aggregation BioOne Complete (https://bioone.org/subscribe), the BioOne Complete Archive (https://bioone.org/archive), and the BioOne eBooks program offerings ESA eBook Collection (https://bioone.org/esa-ebooks) and CSIRO Publishing BioSelect Collection (https://bioone.org/esa-ebooks) and CSIRO Publishing BioSelect Collection (https://bioone.org/csiro-ebooks).

Your use of this PDF, the BioOne Digital Library, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at www.bioone.org/terms-of-use.

Usage of BioOne Digital Library content is strictly limited to personal, educational, and non-commmercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne is an innovative nonprofit that sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

BRAIN CHOLINESTERASE ACTIVITY OF NESTLING GREAT EGRETS, SNOWY EGRETS AND BLACK-CROWNED NIGHT-HERONS

Thomas W. Custer¹² and Harry M. Ohlendorf¹³

- 1 U.S. Fish and Wildlife Service, Patuxent Wildlife Research Center, Laurel, Maryland 20708, USA
- ² Mailing address: Gulf Coast Research Station, P.O. Box 2506, Victoria, Texas 77902, USA
- ³ Mailing address: Pacific Coast Research Station, c/o Department of Wildlife and Fisheries Biology, University of California, Davis, California 95616, USA

ABSTRACT: Inhibition of brain cholinesterase (ChE) activity in birds is often used to diagnose exposure or death from organophosphorus or carbamate pesticides. Brain ChE activity in the young of altricial species increases with age; however, this relationship has only been demonstrated in the European starling (Sturnus vulgaris). Brain ChE activity of nestling great egrets (Casmerodius albus) collected from a colony in Texas (USA) increased significantly with age and did not differ among individuals from different nests. Brain ChE activity of nestling snowy egrets (Egretta thula) and black-crowned night-herons (Nycticorax nycticorax) collected in one colony each from Rhode Island, Texas and California (USA) also increased significantly with age and did not differ among individuals from different nests or colonies. This study further demonstrates that age must be considered when evaluating exposure of nestling altricial birds to ChE inhibitors.

Key words: Brain cholinesterase activity, black-crowned night-herons, Nycticorax nycticorax, great egrets, Casmerodius albus, snowy egrets, Egretta thula, pesticide monitoring, field study.

INTRODUCTION

The National Contaminant Biomonitoring Program (NCBP) (U.S. Fish and Wildlife Service, Washington, D.C. 20240, USA) includes sampling several vertebrate species for contaminant analysis: freshwater fish, European starlings (Sturnus vulgaris) and wings of hunter-killed mallards (Anas platyrhynchos) and American black ducks (Anus rubripes) (Jacknow et al., 1986). In an effort to include an estuarine component in the NCBP, the U.S. Fish and Wildlife Service is evaluating the possibility of monitoring contaminant exposure in nestling colonial waterbirds.

Detecting exposure to organophosphorus or carbamate pesticides is one type of contaminant monitoring being evaluated for colonial waterbirds. Measurement of brain cholinesterase (ChE) activity is frequently used as an indication of exposure to these anti-ChE chemicals (Hill and Fleming, 1982). An approach to using brain ChE activity for diagnosis of anti-ChE exposure has been suggested by Ludke et al. (1975). Brain ChE activity >20% below controls or two standard deviations below average of controls indicates probable exposure to a ChE inhibitor, and mortality

is usually associated with brain ChE activities >50% below controls.

Previous research suggests that the development of brain ChE activity differs between altricial and precocial or semiprecocial avian species. Brain ChE activity increases during embryonic development in precocial or semi-precocial species (Hoffman and Eastin, 1981) and activity is similar for immature and adult birds (Ludke et al., 1975; White et al., 1979). In contrast, brain ChE activity for the altricial European starling is age-dependent (Grue et al., 1981; Grue and Hunter, 1984; Robinson et al., 1988). This phenomenon has not been investigated in other altricial species. The objectives of this study were to determine brain ChE activity of nestling great egrets (Casmerodius albus), snowy egrets (*Egretta thula*) and black-crowned night-herons (Nycticorax nycticorax); and to deterine if brain ChE activity of these altricial species is age-dependent.

MATERIALS AND METHODS

Herons and egrets were collected in 1987 from one colony each along the Rhode Island, Texas and California coasts. The Rhode Island colony was located on Gould Island in Newport County (Colony number 352003; 41°37′N, 71°13′W; Os-

TABLE 1. P values generated from analysis of variance of the effects of nest, colony and age on brain cholinesterase in three colonial waterbird species.

Source of variation	P values		
	Great egret	Snowy egret	Black- crowned night-heron
Nest within colony	0.91	0.69	0.91
Colony ^b		0.66	0.99
Age	< 0.01	< 0.01	< 0.01
Colony × age		0.34	0.73

^{*}Age was divided into three categories: 4 to 7, 8 to 12, and 13 to 17 days of age.

born and Custer, 1978); the Texas colony was located on Goat Island, Harris County (29°45′N, 95°35′W); and the California colony was located on West Marin Island in Marin County, California (37°58′N, 122°28′W).

Complete three, four, or five-egg clutches of snowy egrets and black-crowned night-herons were located in each of the three colonies and complete four-egg clutches of great egrets were located in the Texas colony. In Texas and Rhode Island, nests were examined every 2 to 5 days from early incubation through 15 days after the first egg hatched. In California, nestlings were marked at hatching and followed at 2 to 3 day intervals for 15 days. Day of hatch was determined by observing hatching or estimated by assuming cracking of the eggshell occurred 3 days before hatch and pipping 1 day before hatch (T. W. Custer, pers. obs.) or by body measurements including weight (g) and/or length (mm) of tarsus, forearm or bill of newly found nestlings (T. W. Custer, pers. obs.). Nestlings were individually marked with bands, ink or nail polish as soon as they were discovered. Within each clutch sampled, the nestling from the first egg to hatch was collected when about 15-days-old, the second to hatch was collected when about 10-day-old, and the third to hatch was collected when about 5-day-old. If it was not possible to collect three nestlings from a clutch, the priority of collection was 15-, 10and then 5-day-old nestlings. The actual ages of nestlings collected were assigned to three categories: 4 to 7, 8 to 12 or 13 to 17 days of age. In addition to nestlings, eggs of snowy egrets were collected at the Texas colony 1 or 3 days prior to hatching.

Nestlings and embryos within eggs were killed by CO₂ gas asphyxiation within 2 hr of collection. The carcasses and eggs were preserved immediately on wet ice and frozen at -20 C within 4 hr of death. The ChE assay was conducted

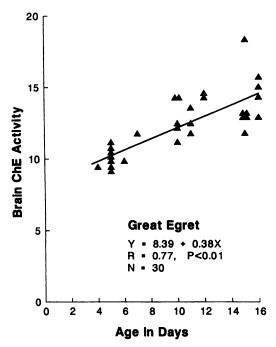


FIGURE 1. Brain ChE activity (µmoles acetylthiocholine iodide hydrolyzed/min/g of tissue, wet weight) of great egret embryos and nestlings.

within 5 months on longitudinally bisected half-brains using the method described by Ellman et al. (1961) and modifications by Hill and Fleming (1982). The reaction was measured on Bausch & Lomb Spectronic 70 spectrophotometer (Bausch & Lomb Inc., Rochester, New York 14604, USA). Acetylthiocholine iodide was used as a substrate. Each brain sample was analyzed in duplicate and an average brain ChE activity was calculated. All samples for each species were analyzed at room temperature (21 to 25 C) on the same day at the Gulf Coast Research Station (Victoria, Texas 77902, USA). Brain ChE activity was expressed as µmoles acetylthiocholine iodide hydrolyzed/min/g of tissue (wet weight).

To determine if colony location, nest or age influenced brain ChE activity of snowy egrets, black-crowned night-herons and great egrets (nest and age only), a two- or three-way AN-OVA was conducted. The relationship between nestling age and brain ChE activity was determined using linear and polynomial regression (Snedecor and Cochran, 1967). A t-test was used to compare slopes of nestling age and brain ChE activity among species. The level of significance for all statistical tests was $\alpha = 0.05$.

RESULTS

Brain ChE activity of great egrets, snowy egrets, and black-crowned night-herons

b Colonies included one each from California, Rhode Island and Texas

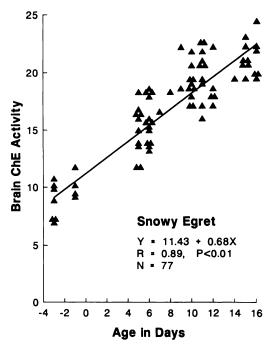
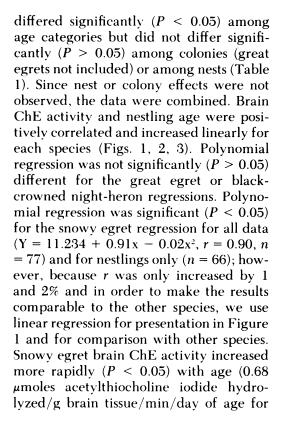


FIGURE 2. Brain ChE activity (µmoles acetylthiocholine iodide hydrolyzed/min/g of tissue, wet weight) of snowy egret nestlings.



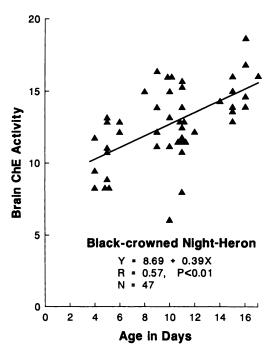


FIGURE 3. Brain ChE activity (µmoles acetylthiocholine iodide hydrolyzed/min/g of tissue, wet weight) of black-crowned night-heron nestlings.

all data; 0.58 μ moles/g/day for nestlings only) than great egrets (0.38 μ moles/g/day) or black-crowned night-herons (0.40 μ moles/g/day); however, there was no difference (P > 0.05) between great egrets and black-crowned night-herons.

Two black-crowned night-heron nestlings had unusually low brain ChE values (Fig. 3). One 11-day-old bird from California had 7.9 μ moles/g and was 40% lower than the value predicted by the regression (13.1 μ moles/g). A 10-day-old nestling from Texas had 6.0 μ moles/g and was 53% lower than the predicted value (12.7 μ moles/g). These values were included in the regression of brain ChE activity versus age (Fig. 3).

DISCUSSION

Nestlings of three altricial waterbird species studied demonstrated increased brain ChE activity with age. These results are similar to those described for starlings by Grue et al. (1981) and Grue and Hunter (1984) which suggest that age must be con-

sidered in evaluating exposure of nestling altricial birds to ChE inhibitors.

This study demonstrates the feasibility of measuring brain ChE activity as part of a National Contaminant Biomonitoring Program using nestling colonial waterbirds. Neither colony location nor nest had a significant effect on nestling brain ChE activity. These results are similar to those of Grue et al. (1981) that demonstrated nest to nest variation did not have a significant effect on brain ChE activity in developing starlings. Although nestling brain ChE activity is age dependent, age can be determined and the relationship between age and brain ChE activity calculated. Age of nestlings can be determined by repeated visits to nests; however, a combination of weight and body measurements may give similar results (T. W. Custer, unpubl. data) and would be less time consuming and would minimize disturbance in the colony.

Brain ChE activity in snowy egrets increased more rapidly with age than great egrets or black-crowned night-herons. This also seemed to be associated with differences in mobility of the nestlings. Fifteenday-old black-crowned night-herons or great egrets usually remained at the nest when disturbed, but 15-day-old snowy egrets often left the nest when disturbed (T. W. Custer, pers. obs.).

As mentioned elsewhere (Grue and Hunter, 1984), brain ChE levels presented here should not be used as normal values in subsequent studies because the measurement techniques are not standardized. However, brain ChE activity can be used as an indicator of exposure to ChE-inhibitors if proper controls are collected and concurrently analyzed (Ludke et al., 1975). Also, measurements of brain ChE activity from field-collected control specimens (within species) are reproducible provided that consistent assay procedures are followed (Hill, 1988).

A relationship between brain ChE activity and nestling age was demonstrated using data from field-collected birds. The

correlation coefficients for brain ChE activity and nestling age for snowy egrets (r = 0.89), great egrets (r = 0.77) and blackcrowned night-herons (r = 0.57) were statistically significant but not as high as observed by Grue et al. (1981) for wild starlings (r = 0.93). Additional variance in these field-collected data could have occurred because of errors in aging the nestlings, food or heat stress (Rattner, 1982), or potential exposure to ChE-inhibiting chemicals. The variance surrounding all age estimates is unknown, but probably was within 2 days. Food and/or heat stress has been shown to reduce brain ChE activity by 10 to 17% (Rattner, 1982). None of the three colonies was located within major agricultural areas; however, local exposure to ChE-inhibiting chemicals was possible from other sources such as chemicals used to control mosquitoes.

ACKNOWLEDGMENTS

We thank David Peterson for performing the field work in Rhode Island and assisting in ChE activity determinations; Timothy Harris for ChE activity determinations, field work in Texas and statistical assistance; Kathy Morois and Grey Pendleton for statistical assistance; Thomas B. Crowley for providing access to West Marin Island; James Myers for organizing the Rhode Island study and assisting in the field; Charles Allin, Dexter Miller, David Vitello and Doug Campbell for field assistance in Rhode Island; Brian Cain, Kirke King and Lawrence Gamble for field assistance in Texas; Carol Allen, Joy Jackman, Roger Hothem, Kathy Marois, Dan Welsh and Max Ohlendorf for field assistance in California; Christian E. Grue, Elwood Hill and Barnett Rattner for reviewing the manuscript; and Clementine Glenn for typing the manuscript.

LITERATURE CITED

ELLMAN, G. L., K. D. COURTNEY, V. ANDRES, JR., AND M. R. FEATHERSTONE. 1961. A new and rapid colorimetric determination of acetylcholinesterase activity. Biochemical Pharmacology 7: 88-98.

GRUE, C. E., AND C. C. HUNTER. 1984. Brain cholinesterase activity in fledgling starlings: Implications for monitoring exposure of songbirds to ChE inhibitors. Bulletin of Environmental Contamination and Toxicology 32: 282-289.

——, G. V. N. POWELL, AND N. L. GLADSON.

- 1981. Brain cholinesterase (ChE) activity in nestling starlings: Implications for monitoring exposure of nestling songbirds to ChE inhibitors. Bulletin of Environmental Contamination and Toxicology 26: 544–547.
- HILL, E. F. 1988. Brain cholinesterase activity of apparently normal wild birds. Journal of Wildlife Diseases 24: 51-61.
- ——, AND W. J. FLEMING. 1982. Anticholinesterase poisoning of birds: Field monitoring and diagnosis of acute poisoning. Environmental Toxicology and Chemistry 1: 27–38.
- HOFFMAN, D. J., AND W. C. EASTIN, JR. 1981. Effects of malathion, diazinon, and parathion on mallard embryo development and cholinesterase activity. Environmental Research 26: 472-485.
- JACKNOW, J., J. L. LUDKE, AND N. C. COON. 1986. Monitoring fish and wildlife for environmental contaminants: The National Contaminant Biomonitoring Program. U.S. Department of the Interior, Fish and Wildlife Service, Fish and Wildlife Leaflet 4, Washington, D.C., 15 pp.
- LUDKE, J. L., E. F. HILL, AND M. P. DIETER. 1975. Cholinesterase (ChE) response and related mortality among birds fed ChE inhibitors. Archives of Environmental Contamination and Toxicology 3: 1-21.

- OSBORN, R. G., AND T. W. CUSTER. 1978. Herons and their allies: Atlas of Atlantic Coast colonies: 1975 and 1976. Biological Services Program, U.S. Fish and Wildlife Service, FWS/OBS-77/08, U.S. Government Printing Office, Washington, D.C., 211 pp.
- RATTNER, B. A. 1982. Diagnosis of anticholinesterase poisoning in birds: Effects of environmental temperature and underfeeding on cholinesterase activity. Environmental Toxicology and Chemistry 1: 329–335.
- ROBINSON, S. C., R. J. KENDALL, R. ROBINSON, C. J. DRIVER, AND T. E. LACHER, JR. 1988. Effects of agricultural spraying of methyl parathion on cholinesterase activity and reproductive success in wild starlings (Sturnus vulgaris). Environmental Toxicology and Chemistry 7: 343–349.
- SNEDECOR, G. W., AND W. G. COCHRAN. 1967. Statistical methods, 6th ed. Iowa State University Press, Ames, Iowa, 593 pp.
- WHITE, D. H., K. A. KING, C. A. MITCHELL, E. F. HILL, AND T. G. LAMONT. 1979. Parathion causes secondary poisoning in a laughing gull breeding colony. Bulletin of Environmental Contamination and Toxicology 23: 281-284.

Received for publication 21 October 1988.