

## **Salmonella from Gopher Tortoises (*Gopherus polyphemus*) in South Georgia**

Authors: Lockhart, J. Mitchell, Lee, Gregory, Turco, Jenifer, and Chamberlin, Linda

Source: Journal of Wildlife Diseases, 44(4) : 988-991

Published By: Wildlife Disease Association

URL: <https://doi.org/10.7589/0090-3558-44.4.988>

---

BioOne Complete ([complete.BioOne.org](https://complete.BioOne.org)) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at [www.bioone.org/terms-of-use](https://www.bioone.org/terms-of-use).

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

---

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

## ***Salmonella* from Gopher Tortoises (*Gopherus polyphemus*) in South Georgia**

**J. Mitchell Lockhart,<sup>1,4</sup> Gregory Lee,<sup>2</sup> Jenifer Turco,<sup>1</sup> and Linda Chamberlin<sup>1,3</sup>** <sup>1</sup>1500 North Patterson St., Biology Department, Valdosta State University, Valdosta, Georgia 31698, USA; <sup>2</sup>Department of the Air Force, Moody Air Force Base, Valdosta, Georgia 31699, USA; <sup>3</sup>Deceased; <sup>4</sup>Corresponding author (email: jmlockha@valdosta.edu)

**ABSTRACT:** From 2002 to 2006, gopher tortoises (*Gopherus polyphemus*) were collected at Moody Air Force Base, Lowndes/Lanier counties, Georgia, USA, and opportunistically surveyed for the presence of *Salmonella* species. Four of 155 (2.6%) cloacal swabs collected from 80 tortoises were positive for the presence of *Salmonella enterica*, and the following serovars were identified: Give, Hartford, Javiana, and Luciana. Female tortoises (5%) were infected at a rate similar to male tortoises (5%). All isolates were obtained from adult tortoises ( $n=73$ ); subadults ( $n=7$ ) were all negative. Each isolated serovar is a potential human pathogen, suggesting appropriate precautions should be emphasized when handling these animals.

**Key words:** Gopher tortoise, *Gopherus polyphemus*, *Salmonella*.

Salmonellosis is a major cause of human illness in the United States, with nearly 1.4 million cases and 600 deaths a year (Mead et al., 1999). In the United States, approximately 74,000 cases of *Salmonella* per year are attributed to handling pet reptiles or amphibians (Mermin et al., 2004). Reptile-associated salmonellosis is particularly associated with invasive disease (Cieslak et al., 1994) and can frequently involve children (Mermin et al., 1997). *Salmonella* usually causes a self-limiting gastrointestinal disorder, but it may have severe manifestations, even causing death (Mermin et al., 1997; CDC, 2003).

Reptiles have long been recognized as a source of *Salmonella* infections (Hinshaw and McNeil, 1945; McNeil and Hinshaw, 1946). Pet turtles were a common source of reptile-associated salmonellosis until 1975 when the Food and Drug Administration banned the sale of turtles less than 4 inches long. Direct transmission of *Salmonella* to humans

may occur through handling of an infected reptile; indirect transmission may occur through contact with feces or by handling objects contaminated by reptiles. In turtles, *Salmonella* is routinely isolated in healthy animals, although *Salmonella* may cause significant pathology (Gonzalez et al., 2005).

The gopher tortoise (*Gopherus polyphemus*) is the only tortoise to occur in the southeastern United States and as a result of degradation, fragmentation, and loss of sandhill habitat (Bozeman, 1971; Wharton, 1978; Noss, 1989), gopher tortoises are state-listed as threatened or endangered across much of their range (Ernst et al., 1994). Gopher tortoises are considered a keystone organism because their burrows are home to more than 350 species of vertebrates and invertebrates, including several threatened and endangered species, such as the eastern indigo snake (*Drymarchon corais cooperi*), gopher frog (*Rana sevosa*), and Florida mouse (*Peromyscus floridanus*) (Jackson and Milstrey, 1989; Lips, 1991; Witz et al., 1991). Although long recognized in pet reptiles, the epizootiologic status of *Salmonella* in free-living tortoises is not well known. The objective of this study was to determine the prevalence of *Salmonella* species in a local gopher tortoise population.

From March 2002 to September 2006, gopher tortoises were captured as part of a study to evaluate a population located on Moody Air Force Base, Lowndes/Lanier counties, Georgia, USA (30°58'00.21"N, 83°10'32.70"W) for the presence of *Mycoplasma agassizii*, the causative agent of upper respiratory tract disease. Cloacal swabs were opportunistically collected from tortoises and examined for the

presence of *Salmonella*. Captured gopher tortoises were manually restrained, placed with the plastron facing dorsally, and a Cary Blair (Becton Dickinson & Company, Sparks, Maryland, USA) transport media swab was collected from the cloaca. Collected swabs were placed on ice and transported to the laboratory. Cloacal swabs taken in the field were broken off into 10 ml of Selenite F broth (Becton Dickinson & Company) and incubated overnight at 37 C. The next day, 2 XLD (Becton Dickinson & Company) agar plates were streaked for isolation from each tube of broth; plates were incubated overnight at 37 C. Single nonlactose- and nonsucrose-fermenting, black (H<sub>2</sub>S-producing) colonies were restreaked to another XLD plate for isolation and further evaluation. If the isolated colonies still demonstrated H<sub>2</sub>S production and did not ferment lactose or sucrose, they were streaked on nutrient agar (Becton Dickinson & Company) and inoculated into an Enterotube II for identification. *Salmonella* isolates were submitted to the National Veterinary Services Laboratory (Ames, Iowa, USA) for serotyping.

Eighty unique tortoises were sampled from one to seven times between 2002 and 2006 for a total of 155 cloacal samples. Four tortoises (5%) tested positive at some point during the study. Four (2.6%) cloacal swabs produced *Salmonella enterica* isolates, and the following serovars were identified: Give, Hartford, Javiana, and Luciana. Thirty-nine female tortoises sampled 74 times produced two (3%) isolates: serovars Give and Luciana. Forty-one male tortoises sampled 81 times yielded the remaining two (3%) isolates: serovars Javiana and Hartford. Seventy-three adults sampled 144 times produced all four isolates; seven subadults, sampled 11 times, were all negative. No significant difference was noted ( $\chi^2 = 0.003$ ,  $P > 0.05$ ) in male versus female infection rate. No apparent trends were noted with regard to year or month of collection (Table 1). Isolates were obtained in June (2005, serovar Give), August (2002,

TABLE 1. Gopher tortoise cloacal swab sample distribution, Moody Air Force Base, Valdosta, Georgia, USA, 2002–2006.

Yr	Tortoise collected	<i>Salmonella</i> positive	Serovar(s)
2002	52	1	Javiana
2003	42	0	—
2004	27	0	—
2005	17	2	Give, Luciana
2006	17	1	Hartford

serovar Javiana; 2006, serovar Hartford), and September (2005, serovar Luciana). No *Salmonella*-positive animals were suspect or positive for the presence of *Mycoplasma*.

Two factors most likely limited the sensitivity of detecting *Salmonella* in the gopher tortoises: the small amounts of feces collected on the cloacal swabs and the enrichment procedure used. Funk et al. (2000), who used rectal swabs as well as fecal samples of 1, 10, and 25 g in their efforts to culture *S. enterica* from swine, found that the relative sensitivity of detection (number of positive animals for a given fecal sample weight/number positive for all fecal sample weights) was directly related to the fecal sample weight. They further determined that the relative sensitivity of detection for rectal swabs was only 8.7% (Funk et al., 2000). Using this estimate, it seems possible that the actual rates of *Salmonella* infection in the gopher tortoises sampled might have been 11 times higher than those reported in the present study. The enrichment procedure used in the present study consisted of incubating the cloacal swabs overnight at 37 C in Selenite F broth. However, in studies with other enrichment broths, Nietfeld et al. (1998) found that adding an additional delayed secondary enrichment (after the primary enrichment) increased the numbers of pig rectal swabs that tested positive for *Salmonella*. For naturally infected animals, only four of 367 pigs were positive for *Salmonella* after a 24-hr enrichment, whereas 10 additional pigs were positive for *Salmonella* after the

delayed secondary enrichment (Nietfeld et al., 1998). Based on these data, one can estimate that the actual rates of *Salmonella* infection in the gopher tortoises sampled might have been 3 times higher than those reported in the present study. Thus, it seems likely that the calculated *Salmonella* infection rates reported in the present study markedly underestimate the actual prevalence of *Salmonella* infection in the gopher tortoises sampled.

*Salmonella* has been recorded frequently in both terrestrial and aquatic chelonians. Previous studies have ranged from zero prevalence in several studies to 100% in *Testudo graeca* in Spain (Hidalgo-Vila et al., 2007). Among land-dwelling tortoises in the United States, *Salmonella* species have been isolated previously from 5.1% of 413 desert tortoises (*Gopherus agassizii*; Dickinson et al., 2001). The present study represents one of the few times wild tortoises have been surveyed repeatedly for the presence of *Salmonella*. Interestingly, each of the four positive tortoises was sampled on multiple occasions (up to seven times); one of the four positives (serovar Javiana) had a subsequent negative sample 2 yr later.

Serovars isolated in this study have been reported previously in human salmonellosis cases. Serovar Give is commonly found in animals, but it is rarely found in humans (Higgins et al., 1997). Serovar Javiana was among the 15 most common serovars isolated in the United States during 1987–1997 (Olsen et al., 2001), it was responsible for at least one outbreak of salmonellosis in Georgia in 2000 (Georgia Department of Human Resources, Division of Public Health, 2001), and it was documented from at least 56 individuals in Georgia in 2004 (Georgia Department of Human Resources, Division of Public Health, 2005). Serovar Hartford was among the top 20 increasing serovars in the United States during 1987–1997 (Olsen et al., 2001), and serovar Luciana is rarely reported from humans (CDC, 2004).

The range of different *Salmonella* ser-

ovars detected in this study is similar to results from other studies (Briane et al., 2004; Corrente et al., 2004). Although the four serovars isolated are documented human pathogens, the extent to which gopher tortoises serve as carriers of potentially zoonotic *Salmonella* remains unknown. Gopher tortoises are generally reclusive animals that spend significant periods sheltered in their burrows, suggesting limited human contact. The potential for human origin of *Salmonella* is unclear, although the military mission at Moody Air Force Base requires field training in gopher tortoise habitat, suggesting a human-influenced environment. These serovars may represent those from the environmental niche, habitat and/or diet components of the gopher tortoise (Briane et al., 2004). Results presented suggest that those individuals who handle wild tortoises should emphasize sanitary precautions.

We thank P. Barber, M. Brown, R. Shaw, and A. Evans for assistance with collection of samples. This work was supported by Department of Defense grant DAMD17-00-2-0034.

#### LITERATURE CITED

- BOZEMAN, J. R. 1971. A sociologic and geographic study of the sand ridge vegetation in the Coastal Plain of Georgia. PhD Dissertation, University of North Carolina, Chapel Hill, North Carolina. 244 pp.
- BRIANES, V., S. TÉLLEZ, J. GOYACHE, C. BALLESTEROS, M. DEL PILAR LANZAROT, L. DOMÍNGUEZ, AND J. F. FERNÁNDEZ-GARAYZÁBEL. 2004. *Salmonella* diversity associated with wild reptiles and amphibians in Spain. *Environmental Microbiology* 6: 868–871.
- CDC. 2003. Reptile-associated salmonellosis-selected states, 1998–2002. *Morbidity and Mortality Weekly Report* 49: 1206–1208.
- . 2004. *Salmonella* Annual Summary, 2004. Public Health Laboratory Information System. Centers for Disease Control and Prevention, Atlanta, Georgia. 95 pp.
- CIESLAK, P. R., F. J. ANGULO, E. L. SUEGER, E. K. MALONEY, AND D. L. SWERDLOW. 1994. Leapin' lizards: A jump in the incidence of reptile-associated salmonellosis (abstract J226). In Program and abstracts of the 34th Interscience Conference on Antimicrobial Agents and Che-

- motherapy, Orlando, Florida, 4–7 October 1994; American Society for Microbiology Washington, D.C.
- CORRENTE, M., A. MADIO, K. G. FRIEDRICH, G. GRECO, C. DESARIO, S. TAGLIABUE, M. D'INCAU, M. CAMPLO, AND C. BUONAVOLGIA. 2004. Isolation of *Salmonella* strains from reptile faeces and comparison of different culture media. *Journal of Applied Microbiology* 96: 709–715.
- DICKINSON, V. M., T. DUCK, C. R. SCHWALBE, J. L. JARCHOW, AND M. H. TRUEBLOOD. 2001. Nasal and cloacal bacteria in free-ranging desert tortoises from the western United States. *Journal of Wildlife Diseases* 37: 252–257.
- ERNST, C. H., R. W. BARBOUR, AND J. E. LOVICH. 1994. *Turtles of the United States and Canada*. Smithsonian Institution Press, Washington, D.C.
- FUNK, J. A., P. R. DAVIES, AND M. A. NICHOLS. 2000. The effect of fecal sample weight on detection of *Salmonella enterica* in swine feces. *Journal of Veterinary Diagnostic Investigation* 12: 412–418.
- GEORGIA DEPARTMENT OF HUMAN RESOURCES, DIVISION OF PUBLIC HEALTH. 2001. Outbreaks of foodborne disease in Georgia, 2000. *Georgia Epidemiology Report* 21(8): 1–3.
- GEORGIA DEPARTMENT OF HUMAN RESOURCES, DIVISION OF PUBLIC HEALTH. 2005. Reptile associated salmonellosis in Georgia residents. *Georgia Epidemiology Report* 17(6): 1–2.
- GONZALEZ, C. M., P. ATANCE, J. S. MARTIN, F. J. PALLARES, AND L. L. VIZCAINO. 2005. Granulomatous hepatitis caused by *Salmonella typhimurium* in a spur-thighed tortoise (*Testudo graeca*). *Veterinary Record* 157: 236.
- HIDALGO-VILA, J., C. DÍAZ-PANIAGUA, C. DE FRUTOS-ESCOBAR, C. JIMÉNEZ-MARTÍNEZ, AND N. PÉREZ-SANTIGOSA. 2007. *Salmonella* in free living terrestrial and aquatic turtles. *Veterinary Microbiology* 119: 311–315.
- HIGGINS, R., A. DESILETS, M. CANTIN, S. MESSIER, R. KHAKHRIA, J. ISMAIL, M. R. MULVEY, D. DAIGNAULT, AND H. CARON. 1997. Outbreak of *Salmonella* GIVE in the province of Quebec. *Canadian Veterinary Journal* 38: 780–781.
- HINSHAW, W. R., AND E. MCNEIL. 1945. *Salmonella* types isolated from snakes. *American Journal of Veterinary Research* 6: 264–266.
- JACKSON, E. R., AND E. G. MILSTREY. 1989. The fauna of gopher tortoise burrows, pp. 86–98. In J. E. Diemer, D. R. Jackson, J. L. Landers, J. N. Layne and D. A. Wood (eds.), *Gopher tortoise relocation symposium proceedings*. Technical report 5. Florida Game and Fresh Water Fish Commission, Tallahassee, Florida.
- LIPS, K. R. 1991. Vertebrates associated with tortoise (*Gopherus polyphemus*) burrows in tour habitats in south-central Florida. *Journal of Herpetology* 25: 477–481.
- MCNEIL, E., AND W. R. HINSHAW. 1946. *Salmonella* from Galapagos turtles, a Gila monster, and an iguana. *American Journal of Veterinary Research* 7: 63–68.
- MEAD, P. S., J. SLUTSKER, V. DIETZ, L. F. MCCAIG, J. S. BRESEE, C. SHAPIRO, P. M. GRIFFIN, AND R. V. TAUXE. 1999. Food-related illness and death in the United States. *Emerging Infectious Diseases* 5: 607–625.
- MERMIN, J., B. HOAR, AND F. J. ANGULO. 1997. Iguanas and *Salmonella* Marina infection in children: A reflection of the increasing incidence of reptile-associated salmonellosis in the United States. *Pediatrics* 99: 399–402.
- , L. HUTWAGNER, D. VUGIA, S. SHALLOW, P. DAILY, J. BENDER, J. KOEHLER, R. MARCUS, AND F. J. ANGULO. 2004. Reptiles, amphibians, and human *Salmonella* infection: A population-based, case-control study. *Clinical Infectious Diseases* 38: S253–S261.
- NIETFELD, J. C., B. KELLY, S. S. DRITZ, I. FEDER, AND J. C. GALLAND. 1998. Comparison of conventional and delayed secondary enrichment for isolation of *Salmonella* spp. from swine samples. *Journal of Veterinary Diagnostic Investigation* 10: 285–287.
- NOSS, R. F. 1989. Longleaf pine and wiregrass: Keystone components of an endangered ecosystem. *Natural Areas Journal* 9: 211–213.
- OLSEN, S. J., R. BISHOP, F. W. BRONNER, T. H. ROELS, N. BEAN, R. V. TAUXE, AND L. SLUTSKER. 2001. The changing epidemiology of *Salmonella*: Trends in serotypes isolated from humans in the United States, 1987–1997. *The Journal of Infectious Diseases* 183: 753–761.
- WHARTON, C. H. 1978. The natural environments of Georgia. Georgia Department of Natural Resources, pp. 277.
- WITZ, B. W., D. S. WILSON, AND M. D. PALMER. 1991. Distribution of *Gopherus polyphemus* and its vertebrate symbionts in three burrow categories. *American Midland Naturalist* 126: 152–158.

Received for publication 17 May 2007.