



Genetically-Determined Polymorphism of Nonspecific Esterases and Phosphoglucomutase in Eight Introduced Breeds of the Silkworm, *Bombyx mori*, Raised in Bulgaria

Author: Staykova, Teodora

Source: Journal of Insect Science, 8(18) : 1-8

Published By: Entomological Society of America

URL: <https://doi.org/10.1673/031.008.1801>

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at www.bioone.org/terms-of-use.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.



Genetically-determined polymorphism of nonspecific esterases and phosphoglucomutase in eight introduced breeds of the silkworm, *Bombyx mori*, raised in Bulgaria

Teodora Staykova

Department of Developmental Biology, section of Genetics, Faculty of Biology, University of Plovdiv, 4000 Plovdiv, Bulgaria

Abstract

Isoenzymes are very suitable markers for the study of the inter-breed diversity of the silkworm *Bombyx mori* L. (Lepidoptera: Bombycidae). More than 250 breeds are raised in Bulgaria, which are not very well studied with regard to their isoenzymic polymorphism. Polymorphism of nonspecific esterases from pupal haemolymph was analyzed, as well as of phosphoglucomutase from different organs of larvae, pupae and imago, from eight introduced breeds. Electrophoresis in polyacrylamide gels was used. A polylocus control of nonspecific esterases, and possible monocus control of phosphoglucomutase was ascertained. Biallele and triallele polymorphism of phosphoglucomutase locus and in three of the esterase loci was determined. The allelic frequencies of the polymorphic loci in each breed were analyzed. Inter-breed differences were found in different allelic frequencies, different heterozygosity and polymorphism.

Keywords: isozymes

Correspondence: tstoikova@yahoo.com

Received: 3 February 2007 | **Accepted:** 28 May 2007 | **Published:** 11 March 2008

Copyright: This is an open access paper. We use the Creative Commons Attribution 3.0 license that permits unrestricted use, provided that the paper is properly attributed.

ISSN: 1536-2442 | Volume 8, Number 18

Cite this paper as:

Staykova T. 2008. Genetically-determined polymorphism of nonspecific esterases and phosphoglucomutase in eight introduced breeds of the silkworm, *Bombyx mori*, raised in Bulgaria. 8pp. *Journal of Insect Science* 8:18, available online: insectscience.org/8.18

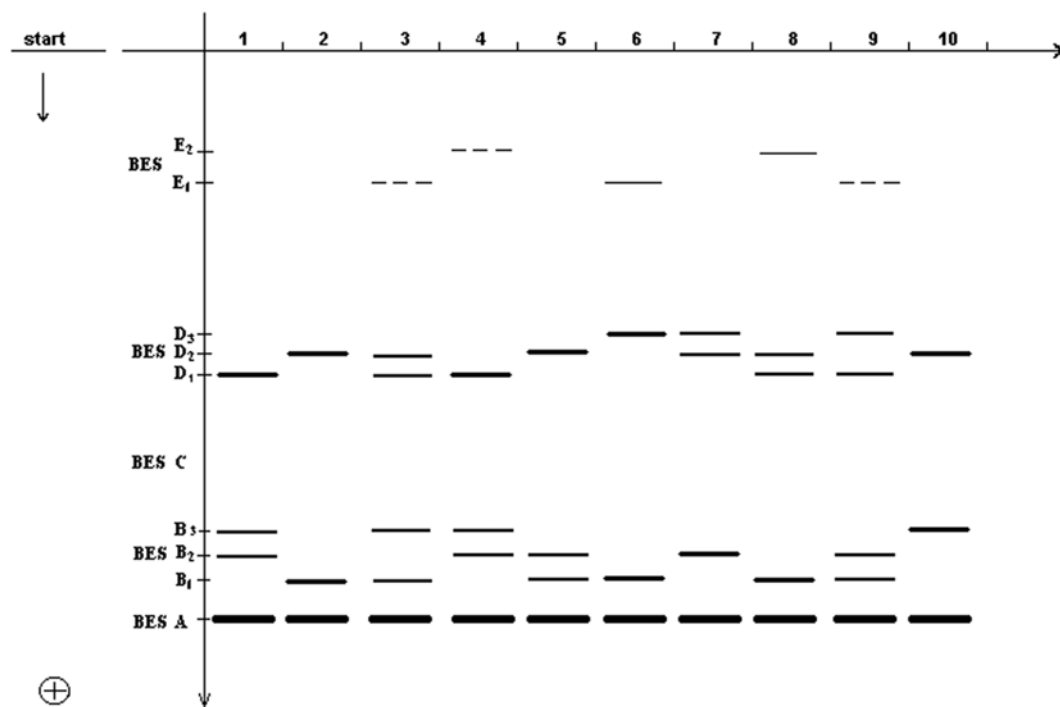


Figure 1. Esterase spectra of *Bombyx mori* haemolymph in 7.5% PAGE: 1–10, pupae 1–2 day.

Introduction

The study of biodiversity in the silkworm is important for selection of useful traits. In order to find out the inter-breed variability of *Bombyx mori* L. (Lepidoptera: Bombycidae), enzyme markers have been found to be especially suitable (Patnaik and Datta 1995). With respect to their genetic structure enzymes are less changeable between individuals than other biochemical constituents of haemolymph, and other tissues (Etebari et al. 2005). This characteristic makes them good biochemical markers. Biodiversity among the more than 250 breeds of mulberry silkworm kept in Bulgaria (Petkov et al. 2006) has been studied mainly on the basis of the most important economic selection characters of quality and quantity. Isoenzymic polymorphism has been less studied in *B. mori* (Shabalina 1990; Stoykova et al. 2003), than genetically determined polymorphism in different enzymes (Eguchi et al. 1965; Hara et al. 1984; Egorova et al. 1985; Eguchi et al. 1988; Takeda et al. 1990; Takeda et al. 1993; Eguchi 1995; Sugimoto et al. 1995; Yamamoto et al. 2000). Within the breeds of *B. mori* raised in Bulgaria, the degree of variability of genetically determined isoenzymic polymorphism has not been previously studied. The objective of the present study was to examine

the polymorphism of nonspecific esterases and phosphoglucosmutase and on that basis to analyze the allelic frequency and degree of genetic heterogeneity with different breeds of mulberry silkworm introduced into Bulgaria from different ecological-geographic areas.

Materials and methods

Individual samples from haemolymph, fat body, silk glands, gut, testes and ovaries of 480 female and male individuals taken from eight breeds of *B. mori* cultured in Bulgaria and known as Almaz, Asahi, Gindga 8, China 23, Kinshu, Maiak 5, Tokai and Showa. The breeds Almaz, Gindga 8 and Maiak 5 were introduced from Azerbaijan, the breeds Asahi, Kinshu, Tokai and Showa from Japan, and China 23 from China. Using 7.5% PAGE the spectrum of nonspecific esterases from different tissues and organs of 1–2 day old pupae were analyzed. Using 7.5% and 6% PAGE the spectrum of phosphoglucosmutase of larvae, pupae and imago was studied. Polyacrylamide gel electrophoresis and isolation of haemolymph and organs were carried out according to Stoykova et al. (2003) and Staykova et al. (2004). Methods of Spencer et al. (1964) and Shaw and Prasad (1970) were used to visualize the phosphoglucosmutase and the esterases respectively. Allele frequencies,

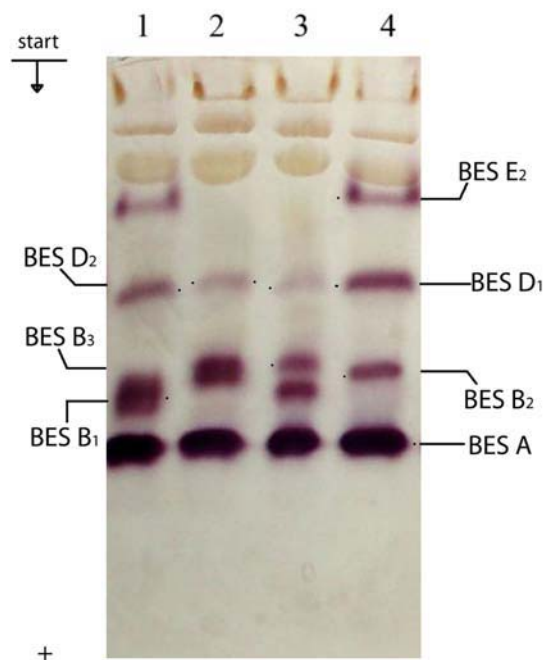


Figure 2. Esterase spectra of *Bombyx mori* haemolymph in 7.5% PAGE: 1–4, Maiak 5; pupae 1st day.

heterozygosity (H) and polymorphism (P) were calculated after Ayala and Kiger (1987).

Results and Discussion

By means of comparative electrophoretic analysis of different tissues and organs of *B. mori*, it was found that for the study of inter-breed and intra-breed polymorphism hemolymph was most suitable for nonspecific esterases, and all tissues except the haemolymph were suitable for phosphoglucomutase.

Nonspecific esterases

Nine esterase fractions belonging to four of the five previously described esterase zones (Stoykova et al. 2003) were found in haemolymph as shown in Figure 1 (BES A; BES B₁, B₂, B₃; BES D₁, D₂, D₃; BES E₁ and E₂). Esterases from zone BES C were not determined.

Breed specificity in the expression of the esterases from zones BES B, BES D and BES E was determined as follows:

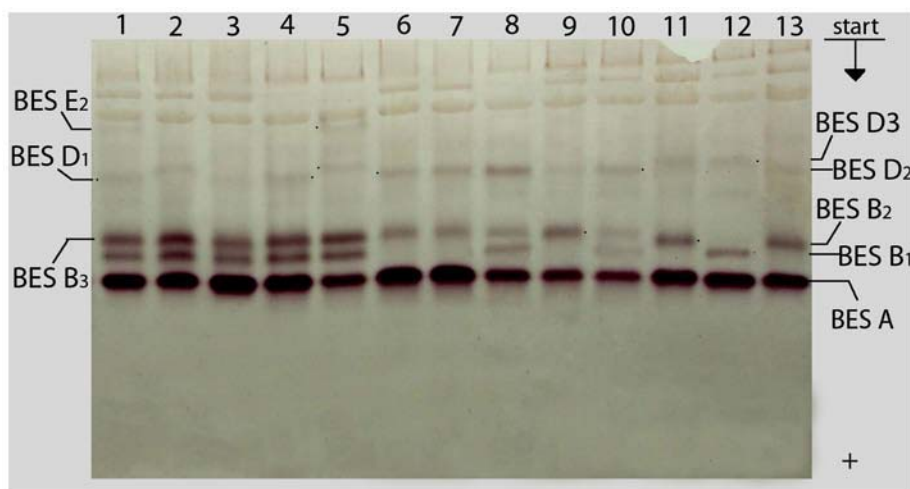


Figure 3. Esterase spectra of *Bombyx mori* haemolymph in 7.5% PAGE: 1–5, Kinshu; 6–10 – China 23; 11–13 – Giandga 8; pupae 1st day.

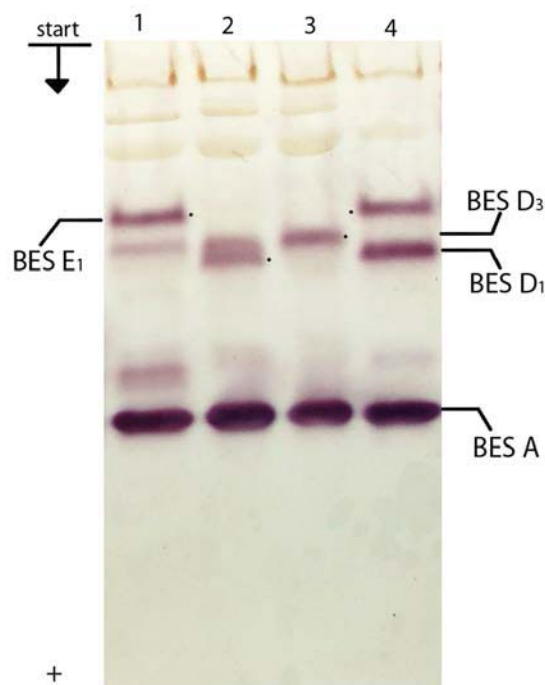


Figure 4. Esterase spectra of *Bombyx mori* haemolymph in 7.5% PAGE: 1–4, Giandga 8; pupae 2nd day.

In zone BES B the bands BES B₁, BES B₂ and BES B₃, were visualized each one separately or in pairs in the breeds Asahi, Gindga 8, China 23 and Maiak 5 (Figures 1, 2, 3). In the Tokai breed, only the bands BES B₁ and BES B₂ were present. In the Kinshu and Showa breeds, only BES B₁ and BES B₃ were present. In the haemolymph from the Almaz breed only fraction BES B₂ was present.

In the Showa breed, only BES D₁ was present. Fractions BES D₁ and BES D₂ were present separately or in combination in the breed Tokai, while BES D₂ and BES D₃ were present in the Asahi breed. Among the pupae studied from all the rest of breeds, fractions BES D₁, BES D₂ and BES D₃ were present each one separately or in pairs (Figures 1, 2, 3, 4).

In zone BES E, in some individuals from Asahi

breed, fractions BES E₁ and BES E₂ were present but showed different intensity (Figure 1). Fraction BES E₁ was present in some individuals from the Almaz and Gindga 8 breeds (Figure 4), and BES E₂, in some individuals from the Kinshu and Maiak 5 breeds (Figures 2, 3). In other individuals from these breeds, esterase bands from zone BES E were absent. With all studied individuals from China 23, Tokai and Showa, the absence of BES E esterase activity was also determined.

The results obtained from this study confirmed the polymorphism established earlier for three of the esterase loci, marked as Bes B, Bes D and Bes E (Stoykova et al. 2003). The allele composition and the frequencies of different alleles was analyzed on the basis of this polymorphism. Breed specificity was ascertained (Table 1).

Table 1. Allele frequencies at polymorphic esterase and phosphoglucosmutase loci in the studied breeds of *Bombyx mori*.

Breed	Allele frequency											
	Bes B			Bes D			Bes E			Pgm A		
	Bes B ₁	Bes B ₂	Bes B ₃	Bes D ₁	Bes D ₂	Bes D ₃	Bes E ₀	Bes E ₁	Bes E ₂	Pgm A ₁	Pgm A ₂	Pgm A ₃
Almaz	0	1	0	0.5	0.44	0.056	0.86	0.14	0	0.32	0.43	0.25
Asahi	0.53	0.18	0.28	0	0.43	0.57	0.47	0.45	0.08	0.11	0.82	0.07
Gindga 8	0.44	0.4	0.16	0.34	0.43	0.23	0.89	0.11	0	0	0.83	0.17
China 23	0.48	0.09	0.43	0.37	0.36	0.27	1	0	0	0.1	0.73	0.17
Kinshu	0.4	0	0.6	0.65	0.25	0.1	0.73	0	0.27	0	0.47	0.53
Maiak 5	0.5	0.15	0.35	0.41	0.18	0.41	0.44	0	0.56	0.37	0.63	0
Tokai	0.46	0.54	0	0.73	0.27	0	1	0	0	0.35	0.58	0.07
Showa	0.3	0	0.7	1	0	0	1	0	0	0.1	0.73	0.17

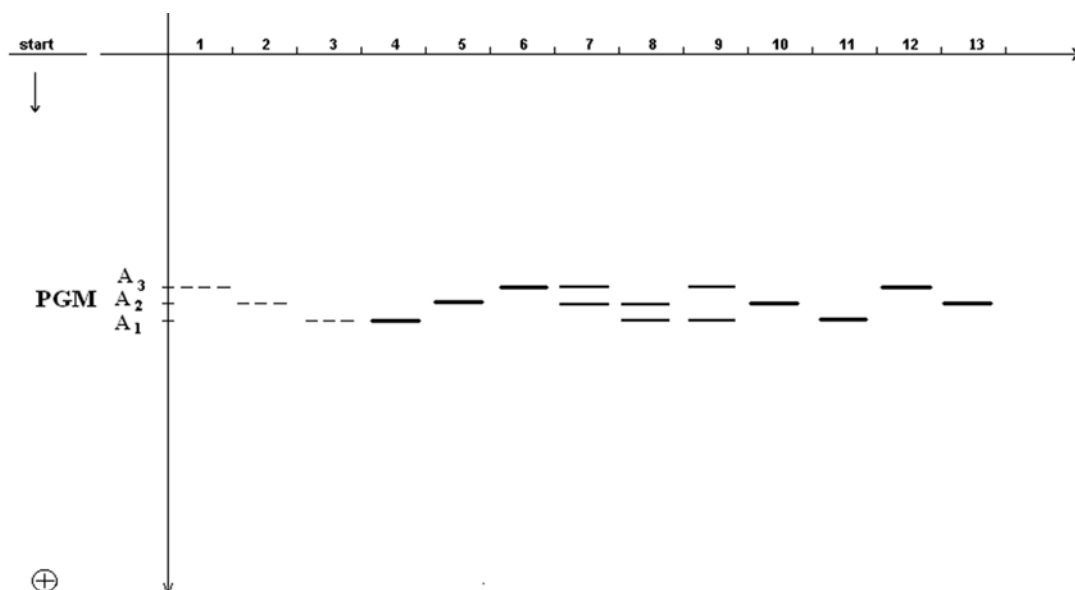


Figure 5. Phosphoglucomutase spectra of *Bombyx mori* tissues in 6% PAGE: 1, 2 and 3, haemolymph; 4–13, fat body, silk glands, testes and ovaries.

In the Asahi, Gindga 8, China 23 and Maiak 5 breeds, the three alleles in locus Bes B - Bes B₁, Bes B₂ and Bes B₃ were present (Table 1). In the Tokai breed the alleles Bes B₁ and Bes B₂ were present, and in the Kinshu and Showa breeds, Bes B₁ and Bes B₃ were present. In the case of Almaz breed only allele Bes B₂ was present. He (1995) and Stoykova et al. (2003) describe biallele polymorphism with codominant alleles on locus Bes B. The results obtained in this study when analyzing the breeds Kinshu, Tokai and Showa confirm these data. A third allele was found in the 5 remaining breeds, except Almaz.

He (1995) determined a triallele polymorphism of the esterases from zone BES D, and Egorova et al. (1985) found monomorphism in the same zone. The results obtained in this study suggest that triallele polymorphism in the locus Bes D with codominant alleles (Bes D₁, Bes D₂ and Bes D₃) is present in the most analyzed breeds (Table 1). Only Showa is monomorphic in locus Bes D. In the Tokai breed, the alleles Bes D₁ and Bes D₂ were present, and in the Asahi Bes D₂ and Bes D₃ were present

The expression of the esterases from zone BES E, only in some individuals, and their absence in other individuals, as well as their different intensity of expression, suggests a triallele polymorphism in locus Bes E with a presence of a null allele (Bes E₀, Bes E₁ and Bes E₂). These results confirm the earlier described

polymorphism with null alleles in the same locus (Stoykova et al. 2003). Though, in some of the breeds analyzed in the present study, a third allele - Bes E₂ was present. Among the studied individuals from the breeds China 23, Tokai and Showa only homozygotes in the null allele were present (Bes E₀). In Almaz and Gindga 8 the allele Bes E₁ was present, while in Kinshu and Maiak 5 the allele Bes E₂ was present. In the case of the breed Asahi all three alleles were present (Table 1).

Sex-dependent expression of haemolymph esterases was not established.

Phosphoglucomutase

There was a lack of sex and stage specificity in the expression of the phosphoglucomutase of the various tissues and organs of larvae, pupae and adult of *B. mori*. Phosphoglucomutase is expressed in the fat body, the silk glands, the gut and the testes and ovaries. The expression of this enzyme in the haemolymph is low during all stages of ontogenesis (Figure 5).

The comparative electrophoretic analysis of different specimens of the breeds studied showed the expression of three phosphoglucomutase fractions in all organs, labeled PGM A₁, PGM A₂ and PGM A₃ in order of mobility (Figures 5, 6). These three bands were expressed independently or in pairs in the various individual spectra. The 6% polyacrilamide gel was found to be more

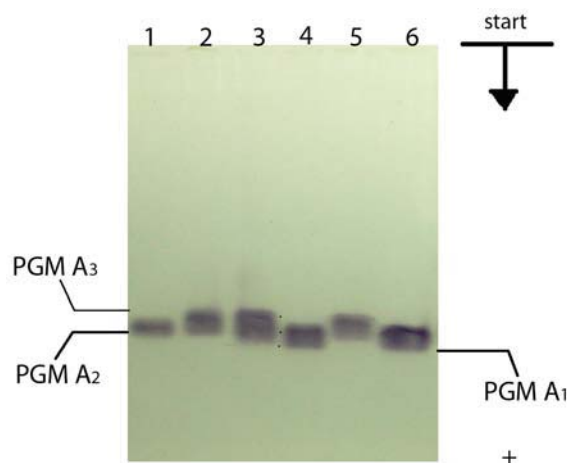


Figure 6. Phosphoglucosmutase spectra of *Bombyx mori* in 6% PAGE - silk glands of larvae 5th instar 7th day: 1–2, Gindga 8; 3–6, Almaz.

appropriate for analysis of the polymorphism of phosphoglucosmutase for *B. mori*.

The intensity and distribution of the three phosphoglucosmutase bands in individuals suggest monolocus control with three codominant alleles, respectively Pgm A₁, Pgm A₂ and Pgm A₃. Well-expressed inter-breed and intra-breed polymorphism was established in the Pgm A locus. The presence of the three alleles was established in Almaz, Asahi, China 23, Tokai and Showa (Table 1). The alleles Pgm A₁ and Pgm A₂ were present in Maiak 5, and Pgm A₂ and Pgm A₃ were present in Gindga 8 and Kinshu. There was breed specificity in differences in allelic composition, and differences in the frequencies of Pgm A alleles (Table 1). Interestingly, the allele Pgm A₂ was present in all breeds studied, and in seven of these breeds this allele was present with the highest frequency. It was only in Kinshu that the frequency of the allele Pgm A₃ was higher than the frequency of Pgm A₂.

Phosphoglucosmutase has not been previously used for a study on the biodiversity of *B. mori*. The polymorphism established provides evidence

that this enzyme is a very suitable marker for the analysis of the inter-breed and intra-breed polymorphism.

Heterozygosity and polymorphism

One of the indicators for the degree of genetic variability, which registers the average frequency of the heterozygote specimens in the population, is heterozygosity (H). The average heterozygosity (observed H_o and expected H_e) for every breed was established on the basis of the polymorphism determined and the allele and genotype frequencies calculated for each locus (Table 2). The heterozygosity observed had the highest value for Kinshu and the lowest value for China 23. The value of H_o was lowest in comparison with H_e for Maiak 5, and closest to the expected ones for Tokai. For all breeds studied the average value of H_o was lower than H_e, which is most likely due to the effects of inbreeding.

Another indicator of the level of genetic variability is its polymorphism (P). On the basis of the results obtained, the level of polymorphism for each of the breeds studied was calculated as well (Table 2). The highest value of P was found for

Table 2. Polymorphism (P) and average heterozygosity (H) in the studied breeds of *Bombyx mori*: H_o – observed heterozygosity; H_e – expected heterozygosity.

Breed	P	H		H _o - H _e
		H _o	H _e	
Almaz	0.6	0.124	0.205	-0.081
Asahi	0.8	0.105	0.196	-0.091
Gindga 8	0.8	0.154	0.219	-0.065
China 23	0.6	0.099	0.185	-0.086
Kinshu	0.8	0.238	0.313	-0.075
Maiak 5	0.8	0.109	0.275	-0.166
Tokai	0.6	0.236	0.285	-0.049
Showa	0.4	0.143	0.212	-0.069

Asahi, Gindga 8, Kinshu and Maiak 5, for which four polymorphic loci were described. The lowest value of P was found for the breed Showa.

Conclusions

On the basis of the results obtained, the following conclusions can be made:

The nonspecific esterases of the haemolymph of *Bombyx mori* are under polygenic control and do not show sex specificity in expression.

Three of the esterase loci are polymorphic, and in two of the breeds studied additional alleles occurred, which has not been previously shown.

Phosphoglucomutase is expressed in all organs of *B. mori* and is possibly controlled by a polymorphic gene and does not show sex specificity.

Esterase and phosphoglucomutase are very suitable markers for analysis of the inter-breed and intra-breed polymorphism for the mulberry silkworm, and for determining the level of genetic variability.

There is a presence of breed specificity in allelic frequency and the heterozygosity of the breeds studied.

The polymorphisms shown for the nonspecific esterases and phosphoglucomutase, and their genetic variability can be expected to be present genetically different parents and could be used during selection to improve strains, (Konicheva et al. 1975; Egorova et al. 1985; Chatterjee et al. 1993; Patnaik & Datta 1995).

Acknowledgments

This work has been financed by "Fund for Scientific Researches" at Ministry of Education and Science – contract VU – AN – 2/2005.

References

Ayala F, Kiger J. 1987. *Modern Genetics*. Zemizdat, Sofia.

Chatterjee SN, Rao C, Chatterjee GK, Ashwath SK, Patnaik AK. 1993. Correlation between yield and biochemical parameters in the mulberry silkworm, *Bombyx mori* L. *Theoretical Applications of Genetics* 87: 385-391.

Egorova T, Naletova E, Nasirillaev Y. 1985. Polymorphic system of silkworm haemolymph esterases as a criterion to make programs for parental specimens crossing. *Biochemistry of Insects - Moscow* 54-62.

Eguchi M, Yoshitake N, Kai H. 1965. Type and inheritance of blood esterase in the silkworm, *Bombyx mori* L. *Japan Journal of Genetics* 40: 15-19.

Eguchi M, Takahama Y, Ikeda M, Horii S. 1988. A novel variant of acid phosphatase isozyme from hemolymph of the silkworm, *Bombyx mori*. *Japan Journal of Genetics* 63: 149-157.

Eguchi M. 1995. Alkaline phosphatase isozymes in insects and comparison with mammalian enzyme. *Comparative Biochemistry and Physiology* 111B: 151-162.

Etebari K, Mirhoseini SZ, Matindoost L. 2005. A study on interspecific biodiversity of eight groups of silkworm (*Bombyx mori*) by biochemical markers. *Insect Science* 12: 87-94.

Hara W, Fujii H, Sakaguchi B. 1984. Activity and polymorphism of amylases in gut contents in different lines of *Bombyx mori*. *Journal of Sericultural Sciences of Japan* 53: 496-500.

He JL. 1995. Studies on the inheritance of a deletion type of the blood esterase isoenzymes in the silkworm, *Bombyx mori* L. *Sericologia* 35: 17-24.

Konicheva A, Egorova T, Philippovich Y. 1975. Study of some enzymes of the haemolymph in different races of silkworm. *Collection of Researches for Moscow State Pedagogical Institute "Lenin"* 18: 157-171.

Patnaik A, Datta RK. 1995. Amylase – its genetics and prospects as a marker in silkworm breeding. *Indian Journal of Sericulture* 34: 82 - 89.

Petkov NI, Tzenov PI, Petkov ZM, Natcheva YS, Vasileva YB. 2006. Silkworm, *Bombyx mori* L. germplasm resources in Bulgaria. National Centre for Agrarian Sciences – Sofia. *Sericultural Experiment Station, Vratza* 264

Shabalina A. 1990. Esterase genetic polymorphism in haemolymph of larvae *Bombyx mori* L. *Comptes rendus de l'Academie bulgare des Sciences* 43: 105-108.

Shaw C, Prasad R. 1970. Starch gel electrophoresis of enzymes – A compilation of Recipes. *Biochemistry Genetics* 4: 297 - 320.

Spencer N, Hopkinson DA, Harris H. 1964. Phosphoglucomutase polymorphism in man. *Nature* 204: 742-745.

Staykova T, Grekov D, Panayotov M. 2004. Electrophoretic analysis of nonspecific esterases in silkworm (*Bombyx mori* L.) female genital organs and eggs. *International Journal of Industrial Entomology* 9: 59-63.

- Stoykova T, Popov P, Dimitrov B. 2003. Electrophoretic analysis of non-specific haemolymph esterases during silkworm (*Bombyx mori* L.) ontogenesis. *Sericologia* 43: 153-162.
- Sugimoto Y, Nagaoka S, Nishimura F, Kusakabe T, Hori K, Koga K. 1995. Interchange in the type of aldolases during embryogenesis in *Bombyx mori* L. *Journal of Sericultural Sciences of Japan* 61: 165-171.
- Takeda S, Azuma M, Eguchi M. 1990. Alkaline phosphatase isozymes of the silkworm midgut: Polymorphism in the enzyme activity, antigen and electroforetogram. *Journal of Sericultural Sciences of Japan* 59: 127-134.
- Takeda S, Azuma M, Itoh M, Eguchi M. 1993. The strain difference and analysis of polymorphic nature of membrane-bound alkaline phosphatase in the midgut epithelium of the silkworm, *Bombyx mori*. *Comp. Biochemistry and Physiology* 104B: 81-89.
- Yamamoto K, Yakiyama M, Fujii H, Kusakabe T, Koga K, Aso Y, Ishioguro M. 2000. Expression of prophenoloxidase m-RNA during hemocyte development. *Bioscience, Biotechnology and Biochemistry* 64: 1197-1202.