

Pasteurella anatipestifer INFECTION IN MIGRATING WHISTLING SWANS

Authors: Wobeser, G., and WARD, G. E.

Source: Journal of Wildlife Diseases, 10(4): 466-470

Published By: Wildlife Disease Association

URL: https://doi.org/10.7589/0090-3558-10.4.466

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at www.bioone.org/terms-of-use.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

Pasteurella anatipestifer INFECTION IN MIGRATING WHISTLING SWANS

G. WOBESER, Department of Veterinary Pathology, and

G. E. WARD, Department of Veterinary Microbiology, Western College of Veterinary Medicine, University of Saskatchewan, Saskatchewan, Saskatchewan, Canada S7N 0W0

Abstract: Pasteurella anatipestifer was isolated from five of seven juvenile whistling swans found sick or dead on two lakes in Saskatchewan, during the 1973 autumn migration. More than 100 sick or dead birds had been reported on one of these lakes. Pathologic lesions were similar to those reported in Pasteurella anatipestifer infection in other species of waterfowl. The possible epizootiology of the disease is discussed.

INTRODUCTION

Pasteurella anatipestifer causes septicemic disease in domestic ducklings, 3,9,10 and has also been reported to cause disease in domestic geese,1 turkeys11 and pheasants.1 Outbreaks of the disease have been reported in semi-domestic black ducks (Anas rubripes)4 and in several species of captive wild waterfowl. Donahue and Olson² isolated the organism from nasal swabs of clinically normal wild Canada geese (Branta canadensis) in Missouri. The only published reports of this disease in free flying wild birds describe an outbreak in black swans (Cygnus atratus) in Tasmania.7,8 The present report describes the disease in whistling swans (Olor columbianus) congregated on lakes near Saskatoon, Saskatchewan during the 1973 autumn migration.

CASE HISTORIES AND METHODS

On October 28, 1973 the senior author found a recently dead, juvenile whistling swan (Bird 1) at the edge of a small, shallow unnamed lake located approximately 35 km southeast of Saskatoon, Saskatchewan. At the time approximately 75 whistling swans; 1,000 ducks, primarily mallards (Anas platyrhinchos); and

25 Canada geese were present on the lake. No other dead birds were noted; however, the entire shoreline was not searched. By November 2, the date of the next visit, the ground was snow covered, and the lake was almost totally frozen. About 50 swans, 100 ducks and 1 Canada goose were present on the ice, but no dead birds were found.

On November 7, three dead juvenile whistling swans (Birds 2-4) were submitted to the diagnostic laboratory. The consignor had visited Goose Lake, (a shallow intermittent lake, measuring approximately 5 x 10 km and located about 75 km southwest of Saskatoon), on November 3, at which time approximately 2,000 swans were present. On November 4, only about 500 swans remained around the small amount of open water available. Of the 500 birds, at least 100 were exhibiting varying degrees of weakness, and at least 15 dead swans were observed. The consignor stated, that with the possible exception of one bird, all of the sick and dead birds were juveniles. Sixteen of the sick swans were collected by the consignor and taken to a site for care. The three birds submitted November 7 had died shortly after collection and had been frozen until submission. On November 14, 3 of the remaining 13 birds were submitted (Birds 5-7). Two of these birds had died within 3 days of capture, while the third had survived for an unspecified length of time. No further specimens were submitted and the fate of the remaining 10 birds was not determined.

Complete necropsies were performed on all birds. Tissues collected for histopathologic examination were fixed in 10% neutral buffered formalin, processed routinely, sectioned at 6 μ m and stained with hematoxylin-eosin. A variety of tissues from each bird were cultured for the presence of bacteria. Swabs taken aseptically from the meninges were cultured in six of the seven cases. Routine cultures were performed using blood agar and MacConkey agar incubated aerobically at 37C.

PATHOLOGY

The seven swans examined were all juveniles in good body condition with large amounts of subcutaneous and intraabdominal fat. The sex, and weight of the birds, together with gross lesions observed, are summarized in Table 1. The most consistent gross lesion was a swollen

liver which was an unusual brick-red color. Fibrinous exudate was prominent in several sites in most birds (Table 1). The entire alimentary tract was empty of ingesta in all birds. A thick collection of fibrinous debris was present in the trachea of two birds, but the lungs appeared normal in all. The spleen was of normal size and pale. The histopathologic lesions observed are summarized in Table 2. The lesions were similar in all birds, but varied in extent and in type of inflammatory cells present. Heterophils were the most numerous cell type in periportal cellular infiltrates in the liver and in the meningeal exudate of Bird 1. In Birds 5 and 7 the inflammatory response was primarily mononuclear, while in the remaining birds a combination of heterophils and mononuclear cells were present. Vascular thrombosis was a common finding in many organs, as was endothelial proliferation and reticulo-endothelial (R.E.) cell activation as described by Karstad et al. Septic thrombi were found in the glomeruli of two birds, and the small foci of myocarditis present in some birds were assocated with septic thrombosis of small vessels.

TABLE 1. Sex, weight and gross lesions observed in juvenile whistling swans with Pasteurella anatipestifer infection.

	Bird No.								
		1	2	3	4	5	6	7	
Lesions	Sex Weight (kg)	F 5.4	F 6.3	M 6.3	M 6.3	F 5.3	M 5.9	M 5.5	
Liver swollen, brick red		+*	+	+	+	+		+	
Fibrinous perihepatitis		**	+	+	_	+	+	_	
Fibrinous pericarditis		_	+		_	+	+	+	
Fibrinous airsaccu	ılitis		+		_		_	_	
Fibrinous tracheitis						+		+	

^{*+} lesion present

^{**--} lesion not present

TABLE 2. Occurrence of histopathologic lesions in juvenile whistling swans with Pasteurella anatipestifer infection.

	Lesion	Bird No.							
Organ		1	2	3	4	5	6	7	
Liver	Fibrinous perihepatitis	_*	+	+	_	+	+	_	
	Periportal cellular infiltration	+**	+	+	+	+	_	+	
	Vacuolar degeneration	+	+	+	+	+	+	+	
	Focal necrosis	_	_	+	_	_			
Heart	Fibrinous epicarditis		+	_		+	+	+	
	Subepicardial edema		+	_		_	_	+	
	Myocarditis		+		_	+	+	+	
Kidney	Embolic glomerulitis	_	_	_	+		_	+	
Spleen	Fibrinous perisplenitis	_	+		_		_	+	
	Lymphoid depletion	+	+	+	+	+	+	+	
	R.E. cell hyperplasia	+	+	+	+	+	+	+	
	Focal necrosis	+	+		+	+	_	+	
Lung	Focal pneumonia		_	_		+	_		
	Fibrinous airsacculitis	_	+	_	_	_	_	_	
Trachea	Fibrinous tracheitis		_	_		+	_	+	
Brain	Fibrinous meningitis	+ -	_+	+	+	+		+	

^{*--} lesion not present

BACTERIOLOGY

Organisms which resembled *P. anatipestifer* were isolated from five of the seven birds examined (Table 3).

On blood agar, the organism produced small transparent, non-hemolytic colonies which were composed of small gramnegative coccobacilli. The organism was cytochrome oxidase positive, catalase positive, indol negative, did not grow on MacConkey agar, and did not ferment

common carbohydrate media. Subcultures of four of the isolates (Birds 1, 2, 5, 7) were sent to Dr. Jessie I. Price, Duck Research Laboratory, Cornell University, Eastport, New York. Dr. Price confirmed the identification of the isolates as *P. anatipestifer*, but found that the isolates failed to agglutinate with any sera available. She reported that the isolate from Bird 2 was tested for pathogenicity in domestic ducklings, and produced lesions in 14 of 19 ducklings inoculated.

^{**+} lesion present

TABLE 3. Results of bacteriologic culture for Pasteurella anatipestifer from seven juvenile whistling swans.

		Bird No.							
	1	2	3	4	5	6	7		
Liver	*	+**	* —	_	_		+		
Spleen	NC**	·* +	_	NC	_	_	+		
Kidney	+	+	_		_		+		
Meninges	NC	+	_	+	+	_	+		
Intestine		_	_		_				
Lung	_	+		_	_	_	+		
Trachea	NC	NC	NC	NC	_	_	NC		
Thoracic airsac	NC	+	NC	NC	NC	NC			
Pericardium	NC	NC	NC	NC	NC	NC			
Nasal cavity	NC	NC	NC		NC	NC			
Abdominal cavity	NC	NC	NC	_	NC	NC	_		

^{*—} P. anatipestifer not isolated

DISCUSSION

There are many similarities between the present cases and the outbreak of P. anatipestifer infection in black swans.7.8 Both occurred in the autumn of the year among birds concentrated during migration, and almost exclusively young birds were involved in both instances. Munday speculated that the causative organism, (at that time unidentified), might be enzootic in swans, only producing disease among birds in overcrowded situations. Under crowded condition, large numbers of organisms might be present in the environment and disease in juvenile birds might be the result of an overwhelming initial infection. Prior to the present outbreak, an unusually large number of swans (over 19,000) were present on Goose Lake. I Similarly, Munday reported overcrowding of black swan prior to the outbreak in Tasmania.

The isolation of *P. anatipestifer* from normal Canada Geese² suggests that the organism may be present in waterfowl in a sub-clinical form. Recently we have isolated *P. anatipestifer* from the nasal cavity of a 4-year-old captive whistling swan, from a local waterfowl collection, which had died of amyloidosis. This isolate was sent to Dr. Price and was found to agglutinate with sera to an avirulent *P. anatipestifer* strain isolated from domestic ducklings in New York. This supports the occurrence of inapparent infections in waterfowl and also suggests that

^{**+} P. anatipestifer isolated

^{***}NC Not cultured

[[]I] Gollop, J. B. 1973. Personal communication. Prairie Migratory Bird Research Centre, Canadian Wildlife Service, Environment Canada, Saskatoon, Saskatchewan.

strains of varying pathogenicity may be present. The pathology of the disease in whistling swans is very similar to that reported in other species^{5,6,9}; however, isolation of the organism is necessary for diagnosis of the disease. *P. anatipestifer* is often difficult to isolate from infected

birds, 6.9.10 perhaps explaining the failure to find the organism in Birds 3 and 6.

The organism was recovered more frequently from the meninges than from other organs; however, the results from the present cases indicate the importance of culturing from several organs.

LITERATURE CITED

- 1. BRUNER, D. W., C. I. ANGSTROM and J. I. PRICE. 1970. Pasteurella anatipestifer infection in pheasants. A case report. Cornell Vet. 60: 491-494.
- DONAHUE, J. M. and L. D. OLSON. 1969. Survey of wild ducks and geese for Pasteurella spp. Bull. Wildl. Dis. Ass. 5: 201-205.
- 3. DOUGHERTY, E. III, L. Z. SAUNDERS and E. H. PARSONS, JR. 1955. The pathology of infectious serositis of ducks. Am. J. Path. 31: 475-487.
- HILBERT, K. F. and W. F. WITTER. 1936. Report of the poultry disease laboratory at Farmingdale, Long Island. Rep. New York State Vet. Coll. 1934-35: 38.
- 5. JORTNER, B. S., R. PORRO and L. LEIBOVITZ. 1969. Central-nervous system lesions of spontaneous *Pasteurella anatipestifer infection* in ducklings. Avian Dis. 13: 27-35.
- KARSTAD, L., P. LUSIS and J. R. LONG. 1970. Pasteurella anatipestifer as a cause of mortality in captive wild waterfowl. J. Wildl. Dis. 6: 408-413.
- MUNDAY, B. L. 1966. "Diseases of Tasmanias' Free-living Animals". Agric. Res. Bull., Tasm. No. 5. 29 p.
- 8. MUNDAY, B. L., A. CORBOULD, K. L. HEDDLESTON and E. G. HARRY. 1970. Isolation of *Pasteurella anatipestifter* from black swan (Cygnus atratus). Aust. Vet. J. 46: 322-325.
- PICKRELL, J. A. 1966. Pathologic changes associated with experimental Pasteurella anatipestifer infection in ducklings. Avian Dis. 10: 281-288.
- PRICE, J. 1959. Studies of Pasteurella anatipestifer infections in White Pekin ducks. Avian Dis. 3: 486-487.
- 11. ZEHR, W. J. and J. OSTENDORF, Jr. 1970. Pasteurella anatipestifer in turkeys. Avian Dis. 14: 557-560.

Received for publication 17 July 1974