

# VARIATION IN RESPONSE OF CHANNEL CATFISH TO Henneguya SP. INFECTIONS (PROTOZOA: MYXOSPORIDEA)

Author: McCRAREN, JOSEPH P.

Source: Journal of Wildlife Diseases, 11(1): 2-7

Published By: Wildlife Disease Association

URL: https://doi.org/10.7589/0090-3558-11.1.2

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at <u>www.bioone.org/terms-of-use</u>.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

# VARIATION IN RESPONSE OF CHANNEL CATFISH TO Henneguya SP. INFECTIONS (PROTOZOA: MYXOSPORIDEA)

JOSEPH P. McCRAREN, I U.S. Fish & Wildlife Service, Warm Water Biologist Center, National Fish Hatchery, Tishomingo, Oklahoma 73460

MARSHA L. LANDOLT 2 and GLENN L. HOFFMAN, 3 U.S. Fish & Wildife Service, Eastern Fish Disease Laboratory, Leetown, (P.O. Kearneysville), West Va. 25430

FRED P. MEYER, I U.S. Fish & Wildlife Service, Fish Farming Experiment Station, Stuttgart, Arkansas 72160

Abstract: Infections in channel catfish (Ictalurus punctatus, Rafinesque) induced by the sporozoan Henneguya (Protozoa: Myxosporidea) result in seven known and diverse disease manifestations. Most outstanding is an interlamellar branchial form responsible for significant losses among immature catfish, and a unique papillomatous form. The question of whether or not the species of Henneguya involved in these cases is H. exilis remains to be resolved.

#### INTRODUCTION

Probably the best known pathogenic Myxosporidians of North American freshwater fish are *Myxosoma cerebralis* and *Ceratomyxa shasta.*<sup>2,0,\*</sup> Each of these species is an important pathogen of salmonids. They are unique, however, as damage inflicted by most sporozoans has not been extensively studied. This paper describes seven manifestations of *Henneguya* infections in channel catfish, including a form which inflicts serious losses among immature individuals.

Seventeen species of *Henneguya* have been described from North American freshwater fishes representing several widely diversified fish families.<sup>1</sup> Most species are histozoic and localize in specific tissues. Infections may appear as white cysts within gills, barbels, adipose fins, the skin, gall bladder, connective tissue of the head, subcutaneous tissues, or in the sclera and muscles of the eye. Spores of *Henneguya* grossly resemble spermatozoa, possess two anterior polar capsules and have an elongate posterior process which may or may not be separated along the sutural plane (Fig. 1). Mode of transmission has not as yet been demonstrated. No means of chemical control is known.

In channel catfish, *Henneguya* infections are broadly categorized with respect to the tissue parasitized and site of spore formation. Two types of branchial (gill) infections are known: an intralamellar form that develops cysts within lamellae, and an interlamellar form that develops cysts between lamellae. Three cutaneous forms have been recognized: a papillomatous form that causes large tumor-like lesions; a skin infection that causes formation of cysts on the external body surface; and a rare adipose fin form that localizes solely within the tissue of that fin. Two other forms are also known: one in connective

2 Presently, Dept. of Pathology, National Zoological Park, Washington, D.C. 20009.

T Presently, Fish Culture Development Center, San Marcos, Texas 78666.

<sup>3</sup> Presently, Fish Farming Experimental Station, Stuttgart, Arkansas 72160.

I Presently, Fish Control Research Laboratory, La Crosse, Wisconsin 54601.



FIGURE 1. Spores of Henneguya. Unstained. 1000 X.

tissue surrounding the mandibular teeth; the other in the gall bladder.

Mature spores from the aforementioned infections are indistinguishable on the basis of shape and dimensions. They closely resemble H. exilis described by Kudo from channel catfish.<sup>3</sup> Further study is needed to determine if H. exilis is the species involved, or if sub-species or other species are implicated.

### BRANCHIAL FORMS

Intralamellar. The intralamellar form is commonly observed among cultured channel catfish. Spore development occurs within capillaries of gill lamellae or blood vessels of gill filaments (Fig. 2).



FIGURE 2. Cysts of intralamellar branchial form of **Henneguya** containing numerous spores. H. and E. 400 X.



The resultant opaque, spore-filled cysts may be found in large numbers and are readily observed in wet mounts. The role of this form as a debilitating agent is suspected but unproven.

Interlamellar. This form of Henneguya develops spores among basal cells between gill lamellae (Fig. 3). As spore formation progresses, lamellar tissue becomes greatly hypertrophied and gill tissue may lose all of its normal appearance. Large masses of spores may be noted among affected cells.

This form, in contrast to the intralamellar form, has been observed to be the causative agent of mass losses among very young cultured channel catfish. Epizootics involving losses of 95% among fingerlings less than 2 weeks old have been documented.<sup>4</sup> Fish up to 25.4 cm in total length have been observed in severe distress when heavily parasitized.

Nearly complete loss of respiratory function accompanies infection. Fish exhibit signs of anoxia, swimming at the surface of ponds with flared gill operculae. Affected fish do not tolerate handling, and most attempts to treat with parasiticides have caused additional loss.

Unfortunately, the interlamellar form appears to be spreading throughout the south-central states due to the unrestricted sale and transport of infected channel catfish fingerlings. It has been observed in five states and is likely to be found in others. It is the form of *Henneguya* that most seriously affects cultured channel catfish. Minchew has described the branchial forms in great detail.<sup>5</sup>

#### CUTANEOUS FORMS

Papillomatous. Tumor-like sporozoaninduced growths are known to occur in fish.<sup>6,7</sup> Growths attributable to *Henneguya* in channel catfish are white and granular (Fig. 4). They have been observed on dorsal, anal, and caudal fins of cultured catfish. The caudal peduncle may also be affected. The largest lesion we have observed measured 1 cm in dia-



FIGURE 3. Interlamellar branchial form of **Henneguya**. Note the large numbers of spores between gill lamellae. 440 X.

meter and completely engulfed the dorsal fin. Hemorrhages are often associated with the lesions. Examination of tissue from affected areas revealed the presence of large numbers of trophozoites, sporoblasts, and spores of *Henneguya*.

Histologically, the growths consist of hyperplastic squamous epithelium and

numerous goblet cells (Fig. 5). Islets partially walled off by condensation from epithelial cells are dispersed throughout the growths, and contain large numbers of *Henneguya* spores. Pronounced inflammation has not been observed. No connective tissue stroma or extensive vascular supply is associated with the



FIGURE 4. Channel catfish infected with papillomatous form of Henneguya.



FIGURE 5. Papillomatous growth in X-section showing cysts containing  $\ensuremath{\text{Henneguya}}$  spores. H. and E. 1000 X.

growths. Tissue proliferation is superficial and does not extend beyond the basement membrane.

Cutaneous cysts. This type of infection is characterized by the presence of visible cysts in the skin. Cysts with diameters of 2-4 mm have been recorded. The condition has been noted in wild and cultured catfish.

Cysts are located in the dermal connective tissue of the dermis (Fig. 6). No evidence of cellular reaction is associated with the condition. Outer layers of cysts contain a germinative layer 100-140  $\mu$ m thick. No recognizable trophozoites or sporoblasts have been noted, suggesting that the cysts represent a mature condition.

Adipose fin. On rare occasions, cultured fingerling channel catfish have been observed with long, narrow white cysts in the adipose fin. The cysts measure approximately  $3 \times 1$  mm and are apparently peculiar to this tissue.



FIGURE 6. Section through cutaneous cyst. H. and E. 37.5 X.

# MANDIBULAR FORM

In June of 1973 the senior author received a preserved portion of mandible collected from a wild channel catfish adult near Stillwater, Oklahoma (Richard D. Spall, personal communication). Eight cysts measuring approximately 1 mm in diameter were visible in the connective tissue surrounding the mandibular teeth. When several cysts were ruptured, and examined microscopically, they were found to contain numerous *Henneguya* spores. Histologically, tissue adjacent to the cysts was found to be necrotic.

# GALL BLADDER FORM

On one occasion, distressed adult channel catfish (domesticated) were examined (3X) and spores of *Henneguya* were found in the gall bladder. The gall bladders were greatly enlarged. Obstruction of the bile duct was evident and the bile was dark green. No other contributing cause to the condition of the fish could be determined. These fish were also infected with the intralamellar branchial form of *Henneguya*.

#### Acknowledgements

Our thanks to Mr. John J. Pulliam who first brought the papillomatous form to our attention, to Dr. Richard D. Spall for the mandibular material, to Messrs. Charlie E. Smith, Roy M. Jones, Richard D. Ivarie, and LTC James L. Stookey (USAMRIID) of Ft. Detrick, Md. for their photographic assistance, and to Mr. Kermit E. Sneed for the use of Fig. 3.

# LITERATURE CITED

- 1. HOFFMAN, G. L. 1967. Parasites of North American Freshwater Fishes. Univ. of Calif. Press. Berkley and Los Angeles.
- HOFFMAN, G. L., C. E. DUNBAR and A. BRADFORD. 1962. Whirling disease of trouts caused by *M. cerebralis* in the United States. Bur. Sport Fish. and Wildl. Spec. Sci. Rep. No. 427: 1-15.
- 3. KUDO, R. R. 1929. Histozoic Myxosporidia found in freshwater fishes of Illinois, USA Arch. Protistenk. 65: 364-378.
- 4. MEYER, F. P. 1969. *Henneguya* infections. *In:* Progress in Sport Fishery Research. Bur. Sport Fish. and Wildl. Resource Publ. No. 88, p. 60.
- MINCHEW, C. DOUGLAS. 1972. Identification and frequency of occurrence of four forms of *Henneguya* sp. found in channel catfish. Proc. 26th Ann. Conf. So. E. Fish and Game Comm. pp. 336-340.
- 6. NIGRELLI, ROSS F. 1953. Tumors and other atypical cell growths in temperate freshwater fishes of North America. Trans. Am. Fish Soc. 83: 262-296.
- NIGRELLI, R. F. and G. M. SMITH. 1938. Tissue responses of Cyprinodon variegatus to the myxosporidian parasite, Myxobolus lintoni, Gurley, Zoologica. N.Y. Zoo. Soc. Vol. 23 (Part 2). pp. 195-202.
- 8. SCHAEFER, W. E. 1968. Studies on the epizootiology of the myxosporidan *Ceratomyxa shasta*, Noble. Calif. Fish and Game. 54: 90-99.
- 9. WALES, J. and H. WOLF. 1955. Three protozoan diseases of trout in California. Calif. Fish and Game. 41: 183-187.

Received for publication 12 January 1973