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Antibodies to *Mycobacterium bovis* in Wild Carnivores from Doñana National Park (Spain)

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ABSTRACT: We conducted a retrospective serologic survey for antibodies against the MPB70 protein of Mycobacterium bovis in wild carnivores from Doñana National Park (southwestern Spain). Serum samples from 118 red foxes (Vulpes vulpes), 39 Iberian lynx (Lynx pardinus), 31 Eurasian badgers (Meles meles), five Egyptian mongoose (Herpestes ichneumon), four European genet (Genetta genetta), and one Eurasian otter (Lutra lutra) were analyzed using an indirect competitive enzyme-linked immunoassay. Antibodies against the MPB70 protein of \dot{M} . bovis were detected in seven badgers, five foxes, and one lynx. The frequency of positive animals was significantly higher in badger (23%) than in lynx (3%) and fox (4%). Antibodies were not detected in other species. Annual antibody frequency peaked at 38% in badgers and 11% for red fox. These species may contribute to persistence of bovine tuberculosis in Doñana.

Key words: Badger, Doñana, epidemiology, Iberian lynx, MPB70, *Mycobacterium bovis*, red fox, wild carnivores.

Wild animals infected with Mycobacterium bovis not only represent a public health risk (Van Tiem, 1997) but also can impact wildlife conservation (Tessaro et al., 1990; Bengis et al., 1996). The transmission of M. bovis between wildlife and domestic livestock can be a major obstacle to bovine tuberculosis (BT) eradication in many countries (Clifton-Hadley et al., 1995b; O'Neil and Pharo, 1995; White & Harris, 1995). Bovine tuberculosis has been reported in wild carnivore species and it is well established that European badgers (Meles meles) are susceptible (Ragg et al., 1995; Keet et al., 1996; Gavier-Widen et al., 2001; Pérez et al., 2001).

In Doñana National Park (DNP), *M. bovis* has been confirmed in wild boar

(Sus scrofa), fallow deer (Dama dama) (León-Vizcano et al., 1991), Iberian lynx (Lynx pardinus) (Briones et al., 2000; Pérez et al., 2001), and red fox (Vulpes vulpes) (Martn-Atance et al., 2005). The objective of this study was to describe the serological status of wild carnivores to determine the extent of prior exposures to *M. bovis*.

Doñana National Park is located in southwestern Spain $(36^{\circ}11'10''-36^{\circ}47'30''N)$ and $6^{\circ}49'10''-6^{\circ}09'05''W)$ covering an area of approximately 2,750 km²; within this area is a 550-km² biological reserve (BR), which receives complete protection. Human activities are restricted to traditional, nonintensive use such as extensive grazing of cattle. Wild ungulates reach high densities in some areas of the park; in the ecotone between marshland and scrubland in the BR, for example, densities of up to 54, 48, and 22 individuals per km² can occur for red deer, wild boar, and fallow deer, respectively.

Carnivores were captured and marked with a microchip between 1988 and 1998 in seven geographical areas within DNP. These included the BR, Marismillas, Acebuche, Torrecuadros, Sotos, Coto del Rey (CR), and other areas not registered; all sampling was concentrated in BR and CR. Blood samples were obtained from the femoral, jugular, or cephalic veins and 247 different sera were obtained from 198 animals: 133 samples from 118 red fox, 68 samples from 39 Iberian lynx, 36 samples from 31 Eurasian badgers, five samples from five Egyptian mongoose (*Herpestes*) ichneumon), four samples from four European genet (Genetta genetta), and one

	Serum samples						Percentage of competition						
				Animals		Seronegatives				Seropositives			
Specie	n	AF ^a	RF ^b	n	AF	RF	Mean	SD	CI_{95}	Range	Mean SD	CI_{95}	Range
Red fox	133	5	4%	118	5	4%	5.37	1.64	5.37 ± 0.3	8.7-1.9	68.9 2.8	68.9 ± 2.5	72.5-65.9
Iberian lynx	68	3	4%	39	1	3%	5.56	1.56	$5.56 {\pm} 0.38$	9.8 - 2.1	$65.2 \ 4.1$	65.2 ± 4.7	71.4-62.6
Eurasian badger	36	7	19%	31	7	$23\%^{\rm c}$	5.47	2.15	$5.47\!\pm\!0.78$	9.8 - 2.1	$64.8 \ 6.5$	64.8 ± 5.3	66.9-57.1
Egytian mon- goose	5	0	_	5	0	_	6.92	1.90	6.92 ± 1.90	8.7–4.3		—	
European genet	4	0		4	0		7.00	0.55	7.00 ± 0.55	7.6 - 6.5			—
Eurasian otter	1	0		1	0	—	5.40	0	0	—		—	—

TABLE 1. Prevalence of seropositive animals and percentages of competition both in the negative and positive sera for the six carnivore species included in the survey.

^a AF Absolute frequency (number seropositive samples or animals).

^b RF: Relative frequency (positives/number of serum samples of animals tested).

^c Significant difference (P < 0.05), SD = standard deviation, CI = confidence interval.

sample from a Eurasian otter (*Lutra lutra*). Sera were frozen at -80 C until tested.

Antibodies to M. bovis antibodies were detected using the indirect competition enzyme-linked immunosorbent assay (ELISA) technique. This was developed according to the method described by Harboe et al. (1990) with modifications (Acosta et al., 2000). Antigen consisted of MPB70 protein (Harboe and Nagai, 1984) and an anti-*M. bovis* goat serum was used to measure antigen captured by test and control sera. Positive and negative control serum samples were from red fox; positive serum was obtained from a fox that was experimentally vaccinated with *M. bovis* BCG. The anti-M. bovis goat serum, with which the carnivore sera competed, had an optical density (OD) value of 0.110. The negative/positive cutoff value for ELISA with MPB70 antigen the was set at an OD value of 0.018 based on the mean+3 SD of OD values obtained from tuberculosis-negative goat sera (Acosta et al., 2000).

Antibody prevalence was calculated based on the number of serum samples or animals tested during the study period for each species. Prevalence estimates for Eurasian badgers, red foxes, and Iberian lynxes were compared using either the chi-square or Fisher's exact test, at a level of significance of $P \leq 0.05$ (Sokal and Rohlf, 1994). Invariable categorical analyses were used to estimate standardized odds ratios, relating potential risk indicators to the occurrence of antibodies; Cornefield's method was applied to calculate 95% confidence intervals for the odds ratio.

Antibodies were detected in 4%, 4%, and 19% of the total samples collected from Iberian lynx, red fox, and Eurasian badger, respectively; all other species tested negative. It should be noted that the three positive lynx samples (Table 1) were taken from the same animal, which was captured in the 1989, 1990, and 1991; antibody levels were maintained in this animal throughout this period with percentage of competition in the ELISA ranging from 62.6% to 71.4%. Samples from all other recaptures tested negative throughout the study.

The prevalence of seropositive animals was significantly higher in badger (23%) than in lynx (3%) and fox (4%) (Table 1). The competition range for positive sera was not significant between these species (Table 1); 95% confidence intervals of competition values also were very similar between these species.

All of the positive animals were de-

		Sera co	llected	Positi	ves	Relative seropositive frequency			
Species	Year	BR^{a}	CR^{a}	BR	CR	BR	CR	Total	
Badger	1991	7	1	2	1	29%	100%	38%	
	1992	4	0	1		25%		$17\% \ (1/6)^{\rm b}$	
	1993	0	1	0			0%	0%	
	1994	7	0	2		29%		$25\% (2/8)^{\rm b}$	
	1995	0	4	0	1	0%	25%	25%	
	1996	0	4		0		0%	0%	
	1997	3	2	0	0	0%	0%	0%	
Red Fox	1988	0	1		0		0%	0%	
	1990	1	0	0		0%		0%	
	1991	15	3	1	1	7%	33%	11% (2/19) ^b	
	1992	27	0	1		4%		3% (1/32) ^b	
	1993	0	5	_	0		0%	0%	
	1994	22	4	1	1	5%	25%	8%	
	1995	0	10		0		0%	0%	
	1996	0	16		0		0%	0%	
	1997	8	7	0	0	0%	0%	0%	
	1998	0	1	_	0		0%	0%	
Lynx	1989	8	0	1		13%		$11\% (1/9)^{\rm b}$	
	1990	4	0	1		25%		25%	
	1991	9	3	1	0	11%	0%	8%	
	1992	1	1	0	0	0%	0%	$0\%~(0/7)^{\rm b}$	
	1993	1	4	0	0	0%	0%	0%	
	1994	1	4	0	0	0%	0%	$0\% (0/6)^{\rm b}$	
	1995	0	7	_	0	—	0%	0%	
	1996	0	5		0	_	0%	$0\% (0/6)^{\rm b}$	
	1997	0	12	_	0	_	0%	0%	

TABLE 2. Relative frequency of animals seropositive to *Mycobacterium bovis* MPB70 in different capture zones per year.

^a BR = biological reserve; CR = Coto del Rey.

^b Includes samples collected in Marismillas, Acebuche, Torrecuadros, Sotos, and unregistered areas.

tected in CR and BR. The prevalence of seropositive red fox was similar between these areas. For badgers, antibody prevalence was lower in CR than in BR despite the fact that badger density is much higher in CR (Table 2). The low number of detected seropositive animals found did not allow calculating the odds ratio as a function of sex or age group. In Table 2 the annual frequencies of positive sera are shown for the different species in the different years and capture zones.

The indirect ELISA with MPB70 antigen (Harboe et al., 1990) has a high level of sensitivity in goats (93.8%) (Acosta et al., 2000), but is not as sensitive in opossum (21%) (Buddle et al., 1995). Sensitivity for the competitive ELISA (cELISA) with the same antigen ranges from 85% to 89% for cattle and bison (*Bison bison*) (Sugden et al., 1997). Using the MPB83 antigen in the indirect ELISA, sensitivity for badgers is not high (40.7%), even in animals with visible lesions (62%) (Goodger et al., 1994; Clifton-Hadley et al., 1995a). All of these techniques have high specificity. The cELISA with MPB70 has not yet been validated in any of the carnivore species studied in this paper, but it is logical to assume that our results underestimate the actual prevalence of *M. bovis* infection in DNP.

The random group sampling method implies a certain lack of exactness, although it is an adequate technique for epidemiologic surveys such as this one. This deficiency could have been counteracted had it been feasible to carry out a random or systematic selection of the whole population, because the prevalence values of a disease tend to be more variable between groups than within one group (Thrusfield, 1995).

The relative frequency values may be useful to determine which species are at risk for *M. bovis* infection in DNP; for example, prevalence of *M. bovis* MPB70 antibodies in badgers is very high compared to red fox. In the case of the lynx, only one seropositive animal was detected and another lynx died with a generalized *M. bovis* infection (Pérez et al., 2001). These observations suggest that lynx may represent an incidental host for *M. bovis*.

In BR, cases of clinical M. bovis infection in wild boar and fallow deer have been reported and M. bovis has been isolated from fallow deer (1.3%) and red deer (2.5%) fecal samples (León-Vizcano, 1990; León-Vizcano et al., 1991). Ungulate densities are extremely high in BR. Although wild ungulate densities are lower in the CR, both areas are shared with domestic cattle. The frequency of seropositive badgers follows the same trend; however, badger density is higher in CR than in BR $(0.85 \text{ vs. } 0.28 \text{ individuals/km}^2)$. Badgers in DNP feed during the summer on dung beetles associated with ungulate dung, as well as scavenging on ungulate carcasses. Such behavior renders them highly likely to come into contact with M. bovis from infected ungulates. It has been suggested that there may be no relation between the population density of badgers and the prevalence of M. bovis infection (Cheeseman et al., 1998). Given the sparse badger population in DNP, the risk of ungulates being infected by contact with infected badgers should be relatively low.

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